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**BIOMARKERS, FUNCTIONAL
EXPLORATION, and GENOTYPE-
PHENOTYPE CORRELATIONS
(SCN1A GENE SEQUENCING) in
FEBRILE SEIZURES in CHILDREN
SUMMARY**

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PUBLISHED PAPERS

1. **Costea, R.M.**; Maniu, I.; Dobrota, L.; Pérez-Elvira, R.; Agudo, M.; Oltra-Cucarella, J.; Dragomir, A.; Bacilă, C.; Banciu, A.; Banciu, D.D.; Cipăian, C.R.; Crișan, R.; Neamtu, B. Exploring Inflammatory Status in Febrile Seizures Associated with Urinary Tract Infections: A Two-Step Cluster Approach. *Brain Sci.* **2021**, *11*, 1168. <https://doi.org/10.3390/brainsci11091168>
2. Maniu, I.; **Costea, R.**; Maniu, G.; Neamtu, B.M. Inflammatory Biomarkers in Febrile Seizure: A Comprehensive Bibliometric, Review and Visualization Analysis. *Brain Sci.* **2021**, *11*, 1077. <https://doi.org/10.3390/brainsci11081077>
3. Pérez-Elvira, R.; Oltra-Cucarella, J.; Carrobles, J.A.; Moltó, J.; Flórez, M.; Parra, S.; Agudo, M.; Saez, C.; Guarino, S.; **Costea, R.M.**; Neamtu, B. Enhancing the Effects of Neurofeedback Training: The Motivational Value of the Reinforcers. *Brain Sci.* **2021**, *11*, 457. <https://doi.org/10.3390/brainsci11040457>
4. Maniu, I.; **Costea, R.M.**; Dobrota, L.; Bacila, C.; Neamtu, B.M. Exploring Inflammatory Status in Febrile Seizures Associated with Urinary Tract Infections: A Two-Step Cluster Approach. *Proceedings* **2021**, *71*, 3. <https://doi.org/10.3390/IECBS-08108>
5. Maniu, I.; **Costea, RM.**; Maniu, G.; Neamtu, B.M. Inflammatory Biomarkers in Febrile Seizure: A Comprehensive Bibliometric, Review and Visualization Analysis. *Proceedings* **2021**, *71*, 1. <https://doi.org/10.3390/IECBS-08539>
6. **Costea RM**, Maniu I, Dobrota L, Neamtu B. Stress Hyperglycemia as Predictive Factor of Recurrence in Children with Febrile Seizures. *Brain Sciences*. **2020**; 10(3):131. <https://doi.org/10.3390/brainsci10030131>
7. Maniu I, **Costea RM**, Neamtu BM, "Cut-off Values for Biomarkers. A Review of Statistical Methods and an Application Study on the Association Between UTI and CRP in Febrile Seizure," **2020** International Conference on e-Health and Bioengineering (EHB), 2020, pp. 1-4, doi: 10.1109/EHB50910.2020.9280288.

8. **Costea RM**, Revenco N, Hadjiu S, Calcîi C, Sprincean M, Maniu I, Neamțu BM, Dobrotă L. Prevalența infecțiilor de tract urinar în cazul copiilor cu convulsiile febrile, Buletin De Perinatologie – Journal of Research and Practice, 2(87), 2020
9. **Costea RM**, Maniu I, Neamtu BM, Glycemic Status in Febrile Seizures, Acta Medica Transilvanica December 2019;24(4):58-61 DOI: 10.2478/amtsb-2019-0018 Online ISSN 2285-7079
10. **Costea RM**, Maniu I, Dragomir A, Banciu D.D, Neamtu BM, "Cluster Analysis a Profiling Tool in Children with Febrile Seizures," 2019 E-Health and Bioengineering Conference (EHB), 2019, pp. 1-4, doi: 10.1109/EHB47216.2019.8969915.
11. **Costea RM**, Neamtu ML; Neamtu MB. Antioxidanții (Resveratrolul) - posibile terapii adjuvante în profilaxia recurenței convulsiilor febrile. Buletin de Perinatologie. 2017, nr. 1(73), pp. 54-59. ISSN 1810-5289
12. Maniu I, Maniu G, Visa GA, **Costea RM**, Neamtu BM, Frequent Pattern Mining of Risk Factors Predicting Neonatal Seizures Outcomes. Brain – Broad Research in Artificial Intelligence and Neuroscience Volume 9, Issue 4 (November 2018), ISSN 2067-3957
13. **Costea RM**, Neamtu ML, Neamtu MB, The Opportunity of the Electroencephalography in the Diagnostic Approach of Febrile Seizures, Acta Medica Transilvanica, Volume 21, no 3, Sept 2016, pages 50-53
14. Neamtu MB, **Costea RM**, The Laboratory and Neuroimaging Investigations in the Diagnostic Approach of Febrile Seizures, Acta Medica Transilvanica, Volume 21, no 3, Sept 2016, pages 79-83
15. **Costea RM**, Neamtu MB, Maniu GC, Pathological EEG Pattern, Predictive Factor for Recurrence or Epilepsy in Febrile Seizures? Acta Medica Transilvanica, Volume 21, no 4, Dec 2016, pages 22-26
16. **Costea RM**, Neamtu ML. Iron Deficiency -Potential Risk Factor for Febrile Seizures. ESPGHAN- Journal of Pediatric Gastroenterology and Nutrition Vol.62, Supplement 1, May 2016
17. **Costea RM**, Neamtu ML, Febrile Seizures - EEG Study and Risk Factors for Recurrence and for Epilepsy, EAPS, Geneva- Eur J Pediatrics 175 (11), 1616-1617, 2016
18. **Costea RM**, Visa G, Maniu GC, Neamtu BM. Particular Features of Neonatal Seizures Populational Study. World Journal of Research and Review. 2016. 3(6), pp. 45-50

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APPENDIX

- ❖ DNA = deoxyribonucleic acid
- ❖ FH = family history
- ❖ ASLO = antistreptolisin O antibodies
- ❖ BBB = blood-brain barrier
- ❖ Ca = calcium
- ❖ FS = febrile seizures
- ❖ CFS = complex febrile seizures
- ❖ SFS = simple febrile seizures
- ❖ MCHC = mean corpuscular hemoglobin concentration
- ❖ Cl = chloride
- ❖ EEG = electroencephalogram
- ❖ SAP = serum acid phosphatase
- ❖ GEFS+ = genetic epilepsy with febrile seizures plus
- ❖ Hb = hemoglobin
- ❖ HCO_3^- = bicarbonate
- ❖ MCH = mean corpuscular hemoglobin
- ❖ IL = interleukin
- ❖ IL-R = interleukin receptor
- ❖ IL-Ra = interleukin receptor antagonist
- ❖ MRI = magnetic resonance imaging
- ❖ K = potassium
- ❖ Ly = absolute number of lymphocytes
- ❖ Na = sodium
- ❖ pCO_2 = partial pressure of carbon dioxide
- ❖ CRP = C-reactive protein
- ❖ PCT = plateletrit

- ❖ PDW = platelet distribution width
- ❖ pH = hydrogen potential
- ❖ P-LCR = platelet large cell ratio
- ❖ PLT = platelets
- ❖ EVP = endovenous perfusion
- ❖ RBC = red blood cells
- ❖ RDW = red cell distribution width
- ❖ RDW- CV = red cell distribution width – coefficient of variation
- ❖ RDW- SD = red cell distribution width – standard deviation
- ❖ NLR = neutrophil / lymphocyte ratio
- ❖ CNS = central nervous system
- ❖ SNP = single nucleotid polymorphism
- ❖ SNV = single nuclotid variant
- ❖ T^0 = temperature
- ❖ AST = aspartate amino transferase (oxalic glutamic transferase)
- ❖ ALT = alanine amino transferase (pyruvic glutamic transferase)
- ❖ TNF = tumor necrosis factor
- ❖ MCV = medium corpuscular volum
- ❖ MPV = medium platelet volume

INTRODUCTION

The study of febrile seizures, the most common types of seizures, is a continuous and current concern.

The awareness of a particular and persistent inflammatory process in the pathogenesis of FS possibly involved in promoting epileptogenesis arouses the special interest of researchers.

In this sense, the study of these mechanisms and the identification of new predictive potential biomarkers for recurrence or epilepsy are not only attractive research concepts but also essential elements for individualized therapy of patients.

The anxiety-inducing potential of FS for the entourage, concerning the risk of recurrence and/or epilepsy, the potential impact on cognitive and behavioral status, further motivate the complex approach to FS and promote the interest of researchers.

In addition, if some biomarkers have been researched as individual factors for prognosis and differentiation of FS types, the innovation element of the study is their interdependent approach and evaluation.

Another window of opportunity was represented by the identification of certain anamnesis and clinical features and biomarkers in the case of the first FS, relevant both for the pathogenesis of FS and for their prognosis.

Moreover, we aimed at the integration in clinical practice of SCN1A gene mutations, in individualized cases, from the perspective of the diagnostic, therapeutic approach and the prognosis.

At the same time, I aimed to highlight the effect of SCN1A gene mutations in association with phenotypes other than Dravet and the correlations between mutation type and phenotype.

In the GENERAL PART, the **first chapter** contains details about the history, updated definitions, epidemiological data, etiopathogenetic mechanisms in febrile seizures, a particular interest being addressed to risk factors for recurrence of FS, epilepsy or comorbidities, all elements that constitute the research premises.

Febrile seizures are systemic, multifactorial events, secondary to the interaction between 1.) the genetic factors determining the alteration of the inflammatory response, hypothalamic thermoregulation, respectively of the cortical hyperexcitability and, 2.) between the precipitating environmental factors of the infectious-inflammatory context, represented mainly by viral infections with neurotrophic properties, promoting inflammation or metabolic imbalances [453]. The results of studies on the involvement of oligoelements in promoting FS are contradictory [314, 356].

Literature data support the complex interaction between the central nervous system, the vegetative nervous system, endocrine and immune mechanisms in the genesis of febrile seizures. Studies place certain IL-gene polymorphisms, the imbalance between prostaglandins and cytokines involved in both fighting infection and pathological inflammation (predominantly IL-1 β) and anti-inflammatory cytokines (predominantly IL-1RA), the alteration of blood-brain barrier and imbalance in neuroinhibition in the etiopathogenesis of FS. At the same time, neuroinflammation, secondary to altered blood-brain barrier and activation of the inflammatory cascade, is incriminated in epileptogenesis associated with prolonged febrile seizures, therefore placing the recurrence of FS in association with an increased risk of comorbidities [453, 422].

In **the second chapter**, the focus is on the impact of the SCN1A gene in medical practice, starting from details on the structure and function of the SCN1A gene and the encoded Nav1.1 channels, continuing with the phenotypes associated with the SCN1A gene and genotype-phenotype correlations and concluding with details on the impact of genetic diagnosis in optimizing therapy and prognosis.

The clinical picture secondary to the alteration of the function of Nav1.1 channels in the CNS varies from epilepsy [659], mesial sclerosis [658], movement disorders, gait and posture abnormalities [138, 183, 221, 302, 479, 601, 700], migraine [181, 211, 299], alteration of cell proliferation and tumor migration [295, 372, 684, 701], autism [53, 261, 376, 563], mental retardation [43, 604, 691], Alzheimer's dementia [420], sleep disorders / circadian rhythm or thermoregulation, up to cardiac arrhythmias and sudden death syndrome in epilepsy [213, 245, 303, 431, 504, 198]. Psychiatric comorbidities have varied clinical expression from stereotypical behavior, social integration disorders, to spatial memory deficiencies and attention deficit [213, 539].

Pathogenic variants of SCN1A have been documented in a wide range of epileptic syndromes and especially with severe childhood myoclonic encephalopathy (Dravet syndrome). Mutations in the SCN1A gene have autosomal dominant inheritance, with

incomplete penetrance and variable expressiveness [216]. In general, mutations with protein truncation and missense mutations in the pore-forming regions of sodium channels are determinants of a severe phenotype (Dravet syndrome) therefore that missense mutations in other regions are more frequently associated with the spectrum of FS-FS plus- Genetic Epilepsy with FS plus [137, 246, 416, 418]. However, the predictive value of the type of mutation for the severity of the phenotype has not been confirmed, probably due to other modifying genes and the polygenic determinism of the FS-FS plus-Dravet spectrum [203, 418, 459].

The SPECIAL PART includes four chapters dedicated to personal research, respectively aspects related to the motivation, objectives and methodology of the research, the results and discussions of the study and finalizing with the conclusions and the global evaluation of the research project.

PERSONAL RESEARCH

Motivation

The present research aimed both the individual evaluation and especially, as an element of innovation, the interdependent evaluation of the inflammatory, metabolic, martial and electroencephalographic context as differentiating factors between FS types and as risk factors for FS recurrence, as well as the identification of certain anamnesis and clinical features and biomarkers for first FS relevant to their pathogenesis. The study aimed to identify easy, cost-effective biomarkers that would require minimally invasive maneuvers with practical applicability for establishing risk models and for prognosis.

If some biomarkers have been researched as individual factors for prognosis and differentiation of FS types, their interdependent approach and evaluation have not been performed so far.

This window of opportunity was decisive in choosing the research topic.

On the other hand, in the second part of the study, the integration of FS in a new concept, of the FS-FS plus-Dravet syndrome spectrum, the perspective of identifying new mutations of the SCN1A gene with the FS-FS plus-Dravet spectrum or with other types of epilepsy not yet associated with this gene and current trends in individualized therapy based on genomic

factors, have been a strong motivation for research. Gene exploration of subjects with the FS-FS plus-Dravet phenotype could have practical applicability in the early optimization of antiepileptic therapy and the improvement of late prognosis (both for seizures and for possible comorbidities).

The main objectives of the research were represented by:

- complex, individual and interdependent assessment of the inflammatory, metabolic, martial and electroencephalographic context as factors of discrimination between FS types and as risk factors for late recurrence;
- identification of the presence of SCN1A mutations in association with phenotypes other than Dravet and of correlations between the type of mutation and the phenotype;
- highlighting the differences between the efficiency of the SCN1A sequencing in the case of subjects with a typical phenotype associated with SCN1A mutations (FS-FS plus-Dravet spectrum) and in the case of subjects with another phenotype;
- identification of genetic testing criteria relevant to the etiological diagnosis involving the patient and his family, including aspects related to prognosis, family planning and therapeutic intervention.

As secondary objectives, we aimed to identify certain anamnestic-clinical features and biomarkers in the case of the first CF, relevant both for the pathogenesis of CF and for their prognosis.

Methodology

I proposed and performed a prospective study conducted in the Sibiu Pediatric Clinical Hospital, between October 2013 and January 2019.

Patients were included in the study in 2 stages, following the objectives of the study stated above.

In the first stage exclusively subjects with FS were included in the study represented by 208 hospitalized children with 305 FS. In the second stage, 54 subjects with FS-FS plus-Dravet phenotype and 25 subjects with other epilepsy phenotypes and history of fever-induced seizures were selected and included in the study group.

All patients with a history of febrile seizures were included in the study.

In the first stage of the study, the following patients were excluded from the study group: patients with CNS infections, with possible traumatic and metabolic causes suggestive of symptomatic acute seizures, patients with a history of unprovoked seizures, those with more than 2 hours postictal laboratory samples, who received pharmacological treatment and/or ENP with the potential to change the metabolic status, those who received emergency antiepileptics who modified the EEG trace or who underwent laboratory tests with different reference values in another medical unit.

From the second stage of the study (study group with SCN1A sequencing) were excluded patients who did not meet the quality criteria of DNA extraction.

In the first stage of the study, the entire study group included 208 patients with 305 seizures divided into study group and control group according to the rank of the seizure, the FS type and the presence of FS recurrence.

In the second stage of the study, 79 subjects who benefited from SCN1A sequencing were integrated into the whole study group, distributed in the study group and control group according to the presence of SCN1A mutations and phenotype.

Finally, the following groups resulted: study group that included patients with first FS (196 cases), compared to a control group with patients with other FS rank (109 cases); study group including patients with complex first febrile seizures (CFS) (30 cases) compared to patients with first simple febrile seizures (SFS) (166 cases); study group that included CFS patients (69) compared to SFS patients (236 seizures) from the total number of patients admitted; study group with first FS with late recurrence (82 seizures) compared to first FS without late recurrence (114 seizures); study group with SCN1A mutations (68 subjects) compared to a control group without SCN1A mutations (11 subjects) and study group dedicated to patients who benefited from SCN1A sequencing with phenotype from the FS-FS plus-Dravet spectrum (54 subjects) and group control consisting of patients with other phenotypes (25 cases).

In the first stage of the study for the whole group of subjects with FS, the anamnestic, clinical, biological and electroencephalographic evaluations were performed.

The anamnestic-clinical evaluation allowed the collection of data related to the family history of FS, epilepsy or diabetes, personal physiological and pathological history (ante-, peri- and postnatal: evolution of pregnancy, gestational age, type of birth, Apgar score, birth weight, days of hospitalization in maternity), and also allowed assessments on nutritional status, the neurological status of patients, psychomotor development, psychiatric

comorbidities and details about the febrile-critical context (etiological classification of the disease-causing fever, body temperature at the time of seizure, interval from the onset of the fever to the onset of the FS and the clinical aspect of the FS, the classification of the FS (CFS / SFS), the clinical pattern of patients with complex febrile seizures), the length of hospitalization as an indicator of the severity of the disease.

The biological evaluation aimed at determining / dosing inflammatory parameters (CRP, NLR, platelet count, PCT, MPV, PDW, P-LCR), metabolic (pH, pCO₂, HCO₃⁻, lactate, blood glucose, electrolytes - Na, K, Cl, ionic Ca, ASLO) and martial (RBC, Ht, Hb, MCV, MCH, MCHC, RDW-SD, RDW-CV, serum iron, ferritin). The following methods of determination were used: for CRP, ASLO immunoturbidimetric method, for other biochemical parameters (ALAT, ASAT, urea, creatinine, glycemia, calcium, ALP, etc.) the spectrophotometric method or analysis by dry biochemistry, for full blood count and flow cytometry, for the ionogram the potentiometric method ISE directly.

In the second stage of the study, the general, anamnestic-clinical, electroencephalographic and imaging parameters pending the genetic study were represented by age, gender; phenotype and defining characteristics for phenotypic classification, respectively family history of FS or epilepsy, age and aspect of the first seizure, the interval between the first seizure and recurrence, personal history of febrile seizures, neurological status before the onset of seizures and dynamics, EEG appearance at first seizure and in dynamics, brain MRI anomalies, reactivity to the regimen or drug resistance and prognosis.

The genetic exploration involved sequencing SCN1A with the nomination of the type of genetic variant depending on the impact on the protein sequence (missense, frameshift), the effect on structure (deletion, insertion and SNV), prediction of pathogenicity: probably pathogenic, possibly pathogenic and benign.

Genomic DNA was extracted from leukocytes in peripheral blood using an automated robotic DNA isolation system, using the MagCore Super HF 16 nucleic acid extractor and the MagCore Genomic DNA Large Volume Whole Blood extraction kit. The amplification of the 26 exons covering the coding regions of SCN1A and the intron-exon binding areas by polymerase chain reaction (PCR) was performed, followed by the cyclic sequencing on both chains, exclusively exonic, using the MiSeq sequencing system and the MiSeq research (Illumina, San Diego, CA, USA). The classification of the variants was performed according to the annotated and updated ACMG criteria (American College of Medical Genetics, 2015) [538].

Statistical analysis

In the data analysis process, for the variables under study, multiple elements of descriptive statistics were determined and statistical tests were used taking into account the level of significance/probability of error $p < 0.05$.

In the case of quantitative data, the normality of the data was first determined, using Shapiro-Wilk, Kolmogorov-Smirnov tests, histograms, QQ plot charts, and skewness (asymmetry) and kurtosis (excess) shape indicators. Depending on the distribution and the variant (Levene test) we applied parametric tests (Gaussian distribution) such as Student t-test (for equal variants) and Welch's t-test (for equal or unequal variants), both for independent samples and for paired samples, or nonparametric tests - Mann-Whitney test, respectively for more than two groups, the ANOVA analysis of variance test and the Kruskal-Wallis test were used.

To highlight groups that differ on average, we used Dunn-Bonferroni post-hoc analysis as a multiple comparison technique.

In the case of qualitative data, contingency tables were used, the data is analyzed using the Pearson-chi-square test or the Fisher test to verify the existence of a relationship/association between variables (qualitative). Z-tests with Bonferroni correction were performed to detail the results obtained after the contingency tables [404, 443]. To detect changes in the EEG appearance in the dynamics we used the McNemar test, which verifies the significance of the difference between two dependent samples when the variable is a dichotomous one.

In the associative analysis of the data, the following coefficients were calculated: the Pearson correlation coefficient (r) [404, 443], for dichotomous variables the / nominal Phi / Cramers V coefficients, and for ordinal variables the nonlinear association coefficient.

To predict the occurrence of late recurrence we used the logistic regression model, in which we integrated all laboratory parameters from the study that individually had a significant association with the change in the frequency of late recurrence.

Decision trees generated by the CRT (Classification and Regression Tree) method were used to study the interconnections between different markers [252].

After the individual analysis of the laboratory parameters, we used the two-step grouping procedure, in the exploratory mode, to identify subgroups of patients with FS based

on the association of the most frequently used metabolic, inflammatory and defining markers for anemia status.

Graphical representation of the analyzed data was performed using histogram diagrams, or boxplot [404, 443]. Statistical analysis was performed using IBM SPSS Statistics 25 and Microsoft Office Excel / Word 2013.

Results and discussions

Regarding the group of first FS compared to the group of recurrent FS, we found that the group with first FS is characterized by:

- ❖ lower percentages of seizures triggered by T^o less than 38°C and higher percentages of seizures lasting between 5 and 14.9 minutes;
- ❖ significantly lower median values of CRP, NLR, MCV, MCH and HCO_3^- and higher absolute number of lymphocytes;
- ❖ lower percentages of those with low PDW values and mild hypokalemia, respectively higher percentage of those with microcytosis;
- ❖ a higher percentage of clusters defined by iron deficiency anemia using as segmentation variables: $\text{Hb} + \text{MCV} + \text{MCH} + \text{MCHC}$ or $\text{MCV} + \text{RDW-CV}$;
- ❖ relatively higher chance than in the case of recurrent FS to fit in the cluster defined by the normal median values of CRP and NLR.

The association of the first FS with moderately high values of CRP, respectively significantly lower median values of CRP, the higher risk of inclusion in the cluster defined by normal median values of CRP and NLR and lower percentages of subjects with low values of PDW compared to recurrent FS, indicates a distinct inflammatory pattern probably correlated with the genetically determined low seizure threshold.

About the statistically significantly higher absolute number of higher lymphocytes respectively lower NLR in the group with first FS may be related to an increased inflammation dependent on neutrophils and on a lymphocyte-mediated anti-inflammatory response, both with probable genetic determinism, although the complex underlying mechanism has not been fully elucidated [262, 386].

The higher percentage of subjects with iron deficiency anemia in the group of first FS could on the one hand suggest the involvement of iron in the etiopathogenesis of FS through its function in neurotransmission, neural metabolic activity and immunity, and on the other

hand, it can be explained by the concordance of the maximum incidence age of iron deficiency anemia with that of the first FS [217, 240, 314].

Regarding the first CFS group compared to the first SFS group, we found that the first CFS group is characterized by:

- ❖ statistically significantly higher values of the duration of hospitalization, higher percentages of subjects with a family history of epilepsy and lower frequency of seizures occurred in sleep;
- ❖ higher percentages of subjects with stress hyperglycemia, the probability of identifying a first CFS being higher in the case of these subjects than in the absence of stress hyperglycemia;
- ❖ significantly higher percentages of the first CFS in the clusters defined by the presence of stress hyperglycemia compared to the other clusters;
- ❖ significantly higher median values of serum iron level and lower probability of low serum iron level.

The statistically significantly higher percentages of subjects with early CFS and stress hyperglycemia supports seizure activity as the main risk factor for stress hyperglycemia, but the exact contribution of FS activity or febrile/infectious context in the genesis of stress hyperglycemia remains uncertain. The results are consistent with data from the literature that associate stress hyperglycemia with prolonged seizures [368].

Statistically significant differences were identified between the degree of fever and the aspect of FS, both in the group of first FS and in the whole study group of FS: CFSs are predominantly precipitated by T° between 40-41°C, and SFS by T° between 39-40°C.

The percentages of first CFS were significantly higher in the cluster characterized by the highest median values of PDW and the lowest median values of PLT compared to the clusters defined by:

- ❖ median values of PDW by the lower normal limit and increased PLT values;
- ❖ low median values of PDW and normal median values of PLT;
- ❖ median values of PDW and normal PLT;

respectively in the case of subjects with inflammatory status defined by the highest median values of PDW, the lowest values of PLT (both within physiological limits), the highest median values of P-LCR (towards the upper limit of normal) and moderate increased CRP median values, suggestive of platelet activation.

If individually the platelet parameters have been the subject of a small number of studies aimed at differentiating FS types, the integrated "clustering" analysis offers a new perspective including the relationship with the inflammatory cascade in the case of CFS versus SFS.

Regarding the CFS group versus the SFS group, the CFS are characterized by:

- ❖ higher values of hospitalization duration, higher percentages of subjects with a family history of epilepsy and lower frequency of seizures in sleep;
- ❖ higher percentages of subjects with stress hyperglycemia which determines a higher probability of identifying a CFS in the case of these subjects than in the absence of stress hyperglycemia;
- ❖ lower percentage of subjects with normocalcemia;
- ❖ higher risk of an FS to be complex if the seizure occurs in less than an hour than in more than an hour from fever onset

The percentage of CFS was significantly higher in the cluster characterized by normal median values of platelet indices (PLT, PCT, P-LCR, MPV, PDW), slightly increased CRP and NLR compared to the cluster characterized by normal median values of PLT and PCT, low MPV, towards the lower limit of PDW, slightly higher CRP and NLR, results that reveal a distinct inflammatory pattern of FS depending on their type.

Although we did not identify statistically significant differences between the individual acid-base parameters to the FS type, we found a higher percentage of patients with acidosis in the CFS group, possibly due to the longer duration of the seizure.

Regarding the group of first FS with late recurrence versus the group of first FS without late recurrence, the present study reconfirms the known risk factors for FS recurrence: low age, family history of FS, the time interval between fever onset and seizure less than one hour and low degree of T° [587].

The late recurrence group is characterized by median values: lower CRP and pH, respectively higher pCO₂ and ionic Calcium.

In the differentiation of subjects at risk of late recurrence, moderately increased cut off point values of CRP were identified, pH cut off values pH at the limit between mild and moderate alkalosis, pCO₂ at the limit of mild/moderate hypocapnia, lactate values at the threshold of severe hyperlactatemia and lactic acidosis and cut off point of 170mg / dL for blood glucose, compatible with stress hyperglycemia.

The combined analysis of demographic and anamnestic-clinical parameters by decision trees showed that:

- ❖ the strongest risk indicators for recurrence are T^0 for the age category greater than or equal to 20 months and family history of FS for the age of less than 20 months;
- ❖ in the absence of a family history of FS, T^0 is a stronger recurrence risk factor for SFS than for CFS;
- ❖ in the presence of febrile seizure family history and T^0 less than 38 C°, CFS was a stronger risk factor for recurrence;
- ❖ for both types of seizures the late recurrence was indirectly proportional to the time interval between fever onset and seizure, and T^0 ;
- ❖ the model of association T^0 with a family history of FS and time interval between fever and seizure is the best in assessing the risk of late recurrence;

About the inflammatory status, the risk of late recurrence was significantly higher in the cluster (2) defined by normal median values of CRP and lower values of NLR than in cluster (1) with increased median values of CRP and higher NLR values; the number of FS recurrences was significantly higher for subjects in cluster 2 compared to those in cluster 1.

The risk of late recurrence was significantly higher in cluster 2 defined by normal median values of CRP, elevated PCT, slightly lower RDW-SD and median NLR lower than in cluster 1 defined by elevated median CRP and PCT, normal RDW-SD and higher NLR values.

Regarding metabolic status, the risk of late recurrence was significantly higher in the cluster with a compensated respiratory alkalosis (normal median pH values, moderately low pCO₂ and slightly low HCO₃⁻) versus the cluster with a decompensated respiratory alkalosis (high median values of pH, severely low pCO₂ and slightly low HCO₃⁻).

The risk of late recurrence was significantly higher in the cluster (1) with compensated respiratory alkalosis, moderate hyperlactatemia and the tendency to stress hyperglycemia compared to cluster (2) with decompensated respiratory alkalosis with hyperlactatemia and mild hyperglycemia

Using decision trees to identify patterns of risk of recurrence related to the metabolic status and EEG appearance we found the maximum risk of late recurrence for the association of early epileptiform abnormalities (EEG) with the cluster - respiratory alkalosis compensated with moderate hyperlactatemia and the tendency to stress hyperglycemia. The absence of recurrence was predominantly recorded for subjects in the cluster with

decompensated respiratory alkalosis with mild hyperlactatemia and hyperglycemia and without epileptiform abnormalities at both records.

The use of the decision tree (CRT type) allows, as an element of novelty, the identification of a risk model for the presence or absence of late recurrence depending on inflammatory parameters (CRP, NLR, PCT, PDW, RDW-SD, Fe), a model that increases and decreases in depending on the association and value of these parameters.

The results of the study are in line with data from the literature that support the strong modulation of the excitability of neural circuits by pH changes [349, 643]: increased pH being associated with neuronal hyperexcitability, seizure inhibition being facilitated by acidosis. Lower pH values, respectively higher pCO₂ values in the group with late recurrence compared to the group without late recurrence, the association of the risk of recurrence especially with the cluster defined by compensated respiratory alkalosis, moderate hyperlactatemia and the tendency to stress hyperglycemia indicate a metabolic status, particularly of subjects at risk of late recurrence. The results support the involvement of the common mechanisms of hyperlactatemia and stress hyperglycemia in promoting the recurrence of febrile seizures. Thus, in FS, abnormal synchronized discharge of neurons leads to a hypermetabolic status, a high need for oxygen and energy, promoting aerobic metabolism of blood glucose and anaerobic glycolysis. Although the mechanisms of FS are still under debate, immune activity may overlap with the counter-regulation of hormonal activity and proinflammatory status described in stress hyperglycemia. A complex interaction between fever, seizure and infection may be key, in the form of combined stressors, leading to a cumulative, synergistic interaction between proinflammatory cytokines and stress-associated hormones (interleukins, growth hormone, insulin, glucagon [413].

In addition to their important role in hemostasis and thrombosis, studies show that platelets contribute to the inflammatory process and microbial defense of the host by the release of cytokines, antibacterial peptides and microbiocidal proteins, angiogenesis and remodeling [630].

Simultaneous measurement of all platelet indices and their interdependent analysis in association with other inflammatory markers, by clustering or decision trees, can provide a valid tool for assessing disease severity and a perspective on the etiology of changes in platelet indices and prognosis.

Thus, the association of the risk of late recurrence with only moderately high values of CRP, respectively with the clusters that define a mild inflammatory syndrome, in the stage

of pre-activation or early platelet activation, suggests the importance of the genetic factor in promoting FS.

Genetic study

For the genetic evaluation, the study group included 68 subjects - with SCN1A mutations and the control group 11 subjects - without SCN1A mutations.

No significant differences were identified between groups regarding anamnestic, clinical and paraclinical data.

However, subjects in the study group compared to the control group presented: a higher percentage of family history of FS, epilepsy or FS and epilepsy; normal neurological status before and after the seizure onset; clinical picture more frequently consistent with the phenotype of FS or FS plus or Dravet vs. phenotype of FS or other epilepsies not included in the spectrum febrile seizures-febrile seizures plus-Dravet; respectively higher percentages of cases with severe phenotype.

We identified 24 distinct types of mutations and 3 other synonymous variants grouped in 1 to 4 associations in the case of one of the 79 subjects with mutations.

Thus, the most frequent mutations were: NM_001202435.1: c.4793A> T, NM_001202435.1: c.182T> C, NM_001202435.1: c.3778delA and NM_001202435.1: c.4385delA; most of the mutations were of the missense type, SNVs, with a predominantly probably pathogenic clinical significance, the rest were frameshift (deletions and insertions). Pathogenic significance was reported in approximately 45% of patients.

Most mutations are found in the spectrum FS-FS plus-Dravet, the most representative mutations being: NM_001202435.1: c.4793A> T, NM_001202435.1: c.182T> C, NM_001202435.1: c.3778delA and NM_001202435.1 : c.4385delA;

The most common SCN1A mutations in subjects with epilepsy not included in the spectrum of FS-FS plus-Dravet were: NM_001202435.1: c.4793A> T, NM_001202435.1: c.182T> C, NM_001202435.1: c.4766d and NM_001202435.1: c.1024G> C.

The mutation NM_001202435.1: c.4906dupC was identified exclusively with the FS-FS plus-Dravet spectrum.

The percentage of subjects with SNVs was similar between FS-FS plus and other epilepsies; most subjects with deletions presented FS or FS plus phenotype, and those with insertions other types of epilepsy.

Significantly more frequently, subjects with deletions had a normal EEG trace, probably in the context of identifying deletion-type mutations, mainly with the phenotype of FS or FS plus.

Significantly more frequently, insertion-type mutations were associated with a severe phenotype while deletions were more frequently associated with the absence of the severe phenotype.

We found the presence of the same mutations in association with different phenotypes (FS, FS plus, Dravet). We detected SCN1A mutations with other types of genetic epilepsy not previously associated with SCN1A: benign rolandic epilepsy of childhood with centrot temporal spikes, epilepsy pf childhood with absence seizures and idiopathic genetic epilepsy (with adolescence onset) (genetic epilepsy with grand mal seizures on awakening, juvenile myoclonic epilepsy and Jeavons syndrome);

The lower percentage of family history of FS and the higher percentage of family history of epilepsy in the study group vs. the control group, the exclusive presence of a family history of epilepsy and FS in the study group suggests the main association of SCN1A with the GEFS spectrum plus and less with familial FS, generally correlated with other types of mutations.

Although subjects with the Dravet phenotype had more frequent frameshift mutations compared to subjects with FS or FS plus, the differences were not statistically significant due to the small number of cases with the Dravet phenotype.

We found the presence of brain MRI abnormalities in a small percentage of subjects with SCN1A mutations, however, it is difficult to establish a causal relationship of SCN1A with brain abnormalities.

Phenotypic variability found by the presence of the same mutations in association with different phenotypes, FS-FS plus-Dravet, would be explained by polygenic determinism, the existence of regulatory genes or epigenetics. Genome-wide sequencing, especially in families with different phenotypes, could provide additional explanations.

In the case of family history, the higher percentage of FS in the control group compared to the study group, the lower percentage of epilepsy in the control group compared to the study group, and the presence of a family history of epilepsy and FS exclusively in the study group could be suggestive of the association of SCN1A mutations predominantly with the GEFS spectrum plus and less with familial CF, generally correlated with other types of mutations [739].

Moreover, although data from the literature associate *SCN1A* mutations, predominantly with FS plus-Dravet spectrum and exceptionally with FS and other types of epilepsy, in this study *SCN1A* mutations were present in a small number of subjects with other types of genetic epilepsy a causal relationship could have not been certainly established.

Although subjects with the Dravet phenotype had more frequent frameshift mutations compared with FS subjects and FS plus subjects, this observation was consistent with literature associating mutations with protein truncation with severe phenotype and the variability of phenotype severity with missense mutations; the differences were not statistically significant due to the low number of cases with Dravet phenotype [137, 246, 416, 418].

Currently, it is difficult to establish a determinism between mutation and the presence of MRI abnormalities (bilateral hippocampal atrophy, gliosis, suprasellar arachnoid cyst, bilateral nodular heterotopias, frontal cortical dysplasia), in the context in which the origin of structural abnormalities (cortical focal atrophy, focal dysplasia, heterotopias and hippocampal sclerosis) reported in the literature in a minority of patients with *SCN1A* mutations remains unclear [232].

There does not appear to be any correlation between the presence of MRI abnormalities and the duration of epilepsy, the age at the onset of seizures, or the frequency of status epilepticus episodes. The few neuropathological studies available on Dravet's syndrome have provided inconsistent findings, including evidence of subtle brain malformations. However, the basic dysfunction of the *SCN1A* gene could give the brain a unique vulnerability profile whose consequences are not easily revealed by neuropathology and require specific experimental settings to be fully appreciated [232].

On the other hand, the study by Striano and colleagues in a group of patients with severe infantile myoclonic epilepsy reports MRI abnormalities more frequently in patients without *SCN1A* mutations [611], while Lee and the colleagues report large-scale brain changes in patients with epilepsy and *SCN1A* gene mutation, which may be associated with the underlying symptoms of patients [371].

Further longitudinal MRI studies with larger cohorts are needed to confirm the effect of *SCN1A* gene mutation on brain structural development.

Variants in the *SCN1A* gene are a common genetic cause for a wide range of epilepsy phenotypes, from febrile seizures to Dravet syndrome. Focal onset seizures and structural lesions may be present in these patients and the question arises as to whether epilepsy

surgery should be considered, given that data from the literature record postoperatively, in subjects with SCN1A mutations variable results and directly related to the anatomo-electroclinic phenotype of the patient. Patients with Dravet syndrome had unfavorable results, and those with focal epilepsy correlated with a single structural lesion good postoperative prognosis [668].

The interpretation of the data from the genetic study was integrated into the current informational context and reported to the currently available SCN1A databases. However, the process of classifying a genetic variant is continuous and involves the identification of the genetic variant in association with a certain pathology, confirmation association through functional studies that demonstrate not only the validity of the proposed hypothesis but also that clarify the molecular mechanism.

Current data from the literature have allowed the validation of the pathogenic character of certain variants identified in this study. For the new genetic variants, the classification was performed based on the functional prediction programs, but the data validation requires the performance of functional studies and/or re-evaluation in time by reference to the updated SCN1A databases.

At the same time, to validate the pathogenicity of the mutations, it would be advisable to investigate the SCN1A of the parents, especially in cases with a family history of epilepsy and/or FS, taking into account the autosomal dominant transmission with incomplete penetrance and phenotypic variability. The investigation of the parents could be the object of a new study and would also allow the classification of the hereditary or de novo character of the mutation.

Regarding the EEG

The study supports the variability of the detection rate of EEG abnormalities depending on the time of recording, age of subjects, type of febrile seizures and types of abnormalities monitored (epileptiform and/or nonspecific), consistent with data from the literature that mentions a variable prevalence of EEG abnormalities in FS subjects, between 2% and 86% [689].

Early EEG was predominantly associated with nonspecific abnormalities and late EEG with epileptiform abnormalities stating that the timing of recording did not decisively influence the presence of epileptiform abnormalities and that nonspecific abnormalities decreased in dynamics.

The value of EEG in the differentiation of CFS from SFS is limited regardless of the time of examination although pathological EEG predominantly defined by epileptiform abnormalities (frontal or central) is more common in the CFS group.

Regardless of the time of recording, pathological EEG defined by epileptiform (frontal) or nonspecific abnormalities is less common in the group of first FS compared to the group of recurrent FS.

The detection rate of the epileptiform pattern regardless of the time of recording was: higher in the elderly, family history of epilepsy, duration of FS greater than 15 minutes, early recurrence in the first 24 hours and lower in those with hyperpyrexia.

The risk of late recurrence of the first FS was higher correlated with the presence of epileptiform anomalies, especially those located frontally, in the context of mixed data available in the literature [81, 335].

The results of the study are consistent with those of most studies, which associate early recorded EEG in the first 7-10 days with the presence of nonspecific abnormalities, and late EEG with the presence of epileptiform abnormalities [292].

Our results are partially consistent with data from the literature, according to which EEG abnormalities are rarer in SFS compared to CFS [292]. In antithesis, such as data obtained by Maytal et al, other studies identify a similar rate of postictal abnormalities in the two types of FS. An explanation is provided by the inclusion in the study group only of patients with normal neurological examination and the interpretation of results based on a single sleep EEG examination (in the first postictal week) [292, 427, 716]. However, in the absence of an FS-specific EEG trace, in the context of an inconclusive history, the EEG value in differentiating CFS from SFS is limited.

The results are consistent with the data from the literature, according to which there is a low sensitivity of EEG in (unprovoked) seizures until the age of 3 years, and a lower detection rate of the epileptiform pattern at young ages [292, 316]. This result states a better EEG recording accuracy especially at older ages when the frequency of FS should be decreased. The association between the presence of localized late epileptiform abnormalities and family history of epilepsy supports the involvement of the genetic factor in the genesis of FS.

The results of this study converge, regardless of the neurological status of the subjects, with the recommendations of some authors to capitalize on EEG registration in the case of CFS, especially in the presence of other risk factors for epilepsy, which increase the detection rate of epileptiform pattern: atypical seizures, neurological abnormalities, extreme

age, family history of epilepsy [118]. So far described as predictive of EEG abnormalities are focal seizures lasting more than 15 minutes, multiple seizures (repeated within 24 hours) [292], abnormal psychomotor development, family history of epilepsy and the presence of more than one diagnosis criterion for CFS [506], delay in psychomotor development, neurological abnormalities, age under 1 year, seizures facilitated by low fever, atypical seizures and extreme ages [359].

Thus, although there are no class Ia studies about the value of EEG abnormalities for FS recurrence or epilepsy, the arguments that could recommend the appearance of EEG as a predictor for FS or epilepsy cannot be ignored. The absence of EEG abnormalities is described with a low frequency of seizures in the absence of fever so that the presence of epileptiform (focal) abnormalities, predominantly frontal, is a risk factor for later unprovoked seizures [279, 322, 335].

Conclusions

1. The present research supports the multifactorial etiology of FS, with the possible genetic factor determining the susceptibility and prognosis of FS, in the context in which the inflammatory process, through its particularities, cannot be considered a unique promoter element of FS. In the case of the first FS, especially in the case of subjects with late recurrence, there are lower values of inflammatory parameters consistent with the stage of preactivation or early platelet activation compared to recurrent FS and subjects without late recurrence. Moreover, the percentage of CFSs was statistically significantly higher in the cluster defined by values of platelet indices suggestive of initiating platelet activation.
2. The study partially supports the importance of iron in the genesis of CF and its properties to promote the growth and differentiation of immune cells and its interference with cytokine pathways, by identifying a higher percentage of clusters defined by iron deficiency anemia in the group of first FS compared to the group of recurrent FS. Its usefulness in discriminating the type of FS and as an inflammatory marker is supported by identifying a higher percentage of subjects with low serum iron levels (functional iron deficiency) in the CFS group.
3. The association of the risk of late recurrence especially with the cluster defined by compensated respiratory alkalosis, moderate hyperlactatemia and the tendency to

stress hyperglycemia indicates a particular metabolic status of subjects at risk of late recurrence, secondary to the complex interaction between proinflammatory cytokines and stress hormones.

4. The combined analysis of the demographic and anamnestic-clinical parameters, respectively of the inflammatory parameters through decisional trees allowed the elaboration of some risk models that require validation through multicenter studies.
5. The present research suggests the phenotypic variability of SCN1A by polygenic determinism, the existence of regulatory or epigenetic genes, and the association of SCN1A mutations predominantly with the GEFS spectrum plus and less with familial FS.
6. We detected the presence of SCN1A mutations with other types of epilepsy not associated so far with SCN1A mutations, but with "FS" onset; functional studies and SCN1A investigation of parents are to demonstrate the validity of the causal relationship.
7. SCN1A mutations give the brain a unique profile of vulnerability, which could also result in the presence of brain malformations.
8. The EEG exploration of subjects with CF should be individualized according to the presence of risk factors for recurrence, epilepsy or comorbidities, and the time of recording adapted to the objective. In the interpretation of EEG, the parameters that influence the detection rate of EEG anomalies must be taken into account.

Personal contributions. Limitations of the study. Recommendation

Overall evaluation of the study

The present personal research aimed at the individual as well as interdependent evaluation of the inflammatory, metabolic, martial and electroencephalographic parameters as risk factors for late recurrence and discrimination factors between FS types, as well as the integration in the FS evaluation of platelet indices, parameters explored less at the moment. The innovation element of the study is the interdependent evaluation of the above-mentioned parameters, using tree type methods and especially the Two-Step Cluster grouping algorithm.

Combined analysis of demographic and anamnestic-clinical parameters by trees

has identified different risk patterns for late-based recurrence different combinations of anamnestic-clinical parameters (age, family history of epilepsy or FS, fever-seizure onset interval, T°, FS type).

At the same time, the use of the decision tree allowed the identification of a risk model for the presence or absence of late recurrence depending on inflammatory parameters (CRP, NLR, PCT, PDW, RDW-SD, Fe), with increasing and decreasing risk depending on the association and the value of these parameters.

Finally, based on the decision trees, we identified some risk patterns of recurrence related to the metabolic status and EEG anomalies.

Moreover, the research was focused on facilitating the identification of certain anamnestic-clinical features and biomarkers in the case of the first FS (compared to other FS) relevant both for their pathogenesis and especially for prognosis.

The “genetics” chapter of the study aims to highlight the effect of SCN1A mutations in association with phenotypes other than Dravet and the correlations between mutation type and phenotype. At the same time, the identification of genetic testing criteria relevant to the etiological diagnosis would have implications for prognosis, family planning and therapeutic intervention, especially in the era of innovative gene therapies.

Regarding the genetic study in which we identified 24 distinct types of mutations and 3 other synonymous variants grouped in 1 to 4 associations in the case of one of the 79 subjects with mutations, as original findings we noted that:

- the percentage of subjects with SNVs was similar between FS-FS plus and other epilepsies;
- most subjects with deletions presented FS or FS plus phenotype, and those with insertions other types of epilepsy;
- significantly more frequently subjects with deletions had a normal EEG trace, probably in the context of identifying deletions predominantly in relationship with FS or FS plus phenotype;
- the presence of SCN1A mutations has been identified with other types of epilepsies not previously associated with SCN1A (rolandic epilepsy of childhood with centrotemporal spikes, epilepsy of childhood with absence seizures and idiopathic genetic epilepsy (in adolescence) (genetic epilepsy with grand mal seizures on awakening seizures, juvenile myoclonic epilepsy and Jeavons syndrome);
- the brain MRI abnormalities were present in a small percentage of subjects with SCN1A mutation.

We have identified several SCN1A variants not described so far in the literature, classified by the prediction software as pathogenic or possibly pathogenic.

Recommendations for research and general practice:

It would be advisable to perform multicenter studies to validate biomarkers identified as risk factors for late recurrence in the present study.

At the same time, the detection by interdependent analysis of a particular inflammatory status of the first FSs to other FSs should be explored by using more sensitive biomarkers involved in the pathogenesis of FS (cytokines).

In the case of subjects with FS in the evaluation and interpretation of EEG, the parameters that may influence the aspect of EEG must be taken into account: age, family history of epilepsy or FS, time and type of recording.

Determining the right time for EEG recording depends on the goal examination; for the differentiation of FS from other paroxysmal manifestations of nonepileptic type is potentially useful early EEG. The risk of recurrence regarding the EEG trace is not influenced by the time of recording, so that an EEG performed late (between 21-28 days postictal) does not alter the value of this instrument in evaluating the late prognosis.

Although the present study identifies a higher frequency of epileptiform abnormalities in CFSs compared to SFSs, the usefulness of EEG exploration in differentiating FS types is limited.

The benefit of EEG recording should be integrated into the clinical context and considered especially in subjects with CFSs, especially in the presence of other risk factors for late recurrence or epilepsy.

Monitoring the postictal EEG aspect in dynamics, having other temporal landmarks, as well as the integration in the analysis of the sleep EEG pathway, could be the subject of future studies on the FS prognosis.

Phenotypic variability by the presence of the same mutations in association with different phenotypes, FS-FS plus-Dravet, could be explained by polygenic determinism, the existence of genes regulatory or epigenetics. Genome-wide sequencing, especially in families with different phenotypes, could provide additional explanations. The investigation by SCN1A sequencing of the parents could be the object of a new study and would allow both the validation of the pathogenic character of the VUS type mutations and the classification of the hereditary or de novo character of the mutation.

Parental SCN1A investigation is especially recommended in cases of family history of epilepsy and/or FS, taking into account autosomal dominant inheritance with incomplete penetrance and phenotypic variability.

To be able to establish with certainty a cause-effect relationship between the genetic variants SCN1A identified in other types of epilepsy not previously associated with SCN1A mutations additional genetic explorations should be considered (SCN1A sequencing of parents, functional tests for framing the pathogenetic character of the variant, genome sequencing for proband and parents).

Considering the limitations of next-generation sequencing, CNV variants and the fact that deletions/duplications of one or more exons represent a small proportion of pathogenic variants, we recommend MLPA analysis in addition to sequencing SCN1A in cases without SCN1A mutations but with phenotype compatible with FS-FS plus-Dravet spectrum.

The interpretation of the data from the genetic study has been integrated with the current informational context and related to currently available SCN1A databases. However, it must be taken into account that the process of classifying a genetic variant is a continuous one and involves the identification of the genetic variant in association with a certain pathology, confirmation of association through functional studies that demonstrate not only the validity of the proposed hypothesis but also clarify the molecular mechanism is needed.

Current data from the literature have allowed the validation of the pathogenic character of certain variants identified in this study. For new genetic variants, the framing was performed on the basis of functional prediction programs, but data validation requires studies functional and/or reassessment over time by reference to updated SCN1A databases.

At the same time, to validate the pathogenic character of the mutations, it would be advisable to investigate SCN1A of parents especially in cases with a family history of epilepsy and/or FS, considering the autosomal dominant inheritance with incomplete penetrance and phenotypic variability. The investigation of the parents could be the subject of a new study and would also allow the classification of the hereditary or de novo character of the mutation.

In the context of identifying MRI abnormalities in association with SCN1A mutations, evaluation of the epilepsy phenotype, integrated with the understanding of the meaning of genetic variants, is essential in determining the benefit and identifying candidates for epilepsy surgery.

Thus, the value of epilepsy-oriented surgery in patients with an SCN1A variant is based on two aspects: the demonstration of the pathogenicity of the variant and the anatomo-electroclinic phenotype of the patient.

References

1. Abdul Wahab, S.A. et al.: Lack of meaningful genotype-phenotype association in SCN1A-related infantile-onset epileptic encephalopathies. *Neurol. Asia.* 22, 99–111 (2017).
2. Abou-Khalil, B. et al.: Temporal lobe epilepsy after prolonged febrile convulsions: excellent outcome after surgical treatment. *Epilepsia.* 34, 5, 878–883 (1993). <https://doi.org/10.1111/j.1528-1157.1993.tb02105.x>.
3. Abuhandan, M. et al.: Evaluation of Selenium Levels and Mean Platelet Volume in Patients with Simple Febrile Convulsion. *Iran. J. Pediatr.* 24, 4, 401–405 (2014).
4. Abuhandan, M. et al.: The oxidative and antioxidative status of simple febrile seizure patients. *JPMA J. Pak. Med. Assoc.* 63, 5, 594–597 (2013).
5. Adibi, P. et al.: Population-based platelet reference values for an Iranian population. *Int. J. Lab. Hematol.* 29, 3, 195–199 (2007). <https://doi.org/10.1111/j.1751-553X.2006.00843.x>.
6. Aicardi, J., Chevrie, J.J.: The significance of electroencephalographic paroxysms in children less than 3 years of age. *Epilepsia.* 14, 1, 47–55 (1973). <https://doi.org/10.1111/j.1528-1157.1973.tb03941.x>.
7. Akan, H. et al.: Thrombopoietic cytokines in patients with iron deficiency anemia with or without thrombocytosis. *Acta Haematol.* 103, 3, 152–156 (2000). <https://doi.org/10.1159/000041038>.
8. Al Morshedy, S. et al.: Interleukin-1 β and interleukin-1receptor antagonist polymorphisms in Egyptian children with febrile seizures: A case-control study. *Medicine (Baltimore).* 96, 11, e6370 (2017). <https://doi.org/10.1097/MD.0000000000006370>.
9. Allan, S.M. et al.: Interleukin-1 and neuronal injury. *Nat. Rev. Immunol.* 5, 8, 629–640 (2005). <https://doi.org/10.1038/nri1664>.
10. Andersen, L.W. et al.: Etiology and therapeutic approach to elevated lactate levels. *Mayo Clin. Proc.* 88, 10, 1127–1140 (2013). <https://doi.org/10.1016/j.mayocp.2013.06.012>.
11. Annegers, J.F. et al.: Factors prognostic of unprovoked seizures after febrile convulsions. *N. Engl. J. Med.* 316, 9, 493–498 (1987). <https://doi.org/10.1056/NEJM198702263160901>.
12. Annegers, J.F. et al.: Recurrence of febrile convulsions in a population-based cohort. *Epilepsy Res.* 5, 3, 209–216 (1990). [https://doi.org/10.1016/0920-1211\(90\)90040-3](https://doi.org/10.1016/0920-1211(90)90040-3).
13. Annesi, G. et al.: Two novel SCN1A missense mutations in generalized epilepsy with febrile seizures plus. *Epilepsia.* 44, 9, 1257–1258 (2003). <https://doi.org/10.1046/j.1528-1157.2003.22503.x>.
14. Armon, K. et al.: An evidence and consensus based guideline for the management of a child after a seizure. *Emerg. Med. J. EMJ.* 20, 1, 13–20 (2003). <https://doi.org/10.1136/emj.20.1.13>.
15. Arshi, S. et al.: A Study on the Relationship between Hypocapnia and Febrile Seizure at Hazrat Rasool Hospital in Iran during a Three-Year Period of 2013-2015. *Altern. Integr. Med.* 8, 2, 1–4 (2019).
16. Arzimanoglou, A.: Dravet syndrome: from electroclinical characteristics to molecular biology. *Epilepsia.* 50 Suppl 8, 3–9 (2009). <https://doi.org/10.1111/j.1528-1167.2009.02228.x>.
17. Aschemeyer, S. et al.: Structure-function analysis of ferroportin defines the binding site and an alternative mechanism of action of hepcidin. *Blood.* 131, 8, 899–910 (2018). <https://doi.org/10.1182/blood-2017-05-786590>.
18. Ashwal, S.: The Founders of child neurology. Norman Pub. in association with the Child Neurology Society, San Francisco (1990).
19. Ates, I. et al.: Association between high platelet indices and proteinuria in patients with hypertension. *Ann. Lab. Med.* 35, 6, 630–634 (2015). <https://doi.org/10.3343/alm.2015.35.6.630>.
20. Atwa, H. et al.: Possible Role of Iron Deficiency in Occurrence and Recurrence of Febrile Seizures in Children Aged 6-36 Months. *Pediatr. Res.* 70, 5, 150–150 (2011). <https://doi.org/10.1038/pr.2011.375>.

21. Audenaert, D. et al.: A novel GABRG2 mutation associated with febrile seizures. *Neurology.* 67, 4, 687–690 (2006). <https://doi.org/10.1212/01.wnl.0000230145.73496.a2>.
22. Avcil, S.: Evaluation of the neutrophil/lymphocyte ratio, platelet/lymphocyte ratio, and mean platelet volume as inflammatory markers in children with attention-deficit hyperactivity disorder. *Psychiatry Clin. Neurosci.* 72, 7, 522–530 (2018). <https://doi.org/10.1111/pcn.12659>.
23. Aydin, L. et al.: Zinc supplementation prolongs the latency of hyperthermia-induced febrile seizures in rats. *Physiol. Int.* 103, 1, 121–126 (2016). <https://doi.org/10.1556/036.103.2016.1.12>.
24. Azab, S.F. et al.: Interleukin-6 gene polymorphisms in Egyptian children with febrile seizures: a case-control study. *Ital. J. Pediatr.* 42, (2016). <https://doi.org/10.1186/s13052-016-0244-9>.
25. Azam, A.S. et al.: Relationship between serum selenium level and febrile seizure in children. *J Chem Pharm Res.* 7, 13–18 (2015).
26. Baek, S.-J. et al.: Risk of low serum levels of ionized magnesium in children with febrile seizure. *BMC Pediatr.* 18, 1, 297 (2018). <https://doi.org/10.1186/s12887-018-1271-z>.
27. Balosso, S. et al.: A novel non-transcriptional pathway mediates the proconvulsive effects of interleukin-1beta. *Brain J. Neurol.* 131, Pt 12, 3256–3265 (2008). <https://doi.org/10.1093/brain/awn271>.
28. Ban, E. et al.: Receptors for interleukin-1 (alpha and beta) in mouse brain: mapping and neuronal localization in hippocampus. *Neuroscience.* 43, 1, 21–30 (1991). [https://doi.org/10.1016/0306-4522\(91\)90412-h](https://doi.org/10.1016/0306-4522(91)90412-h).
29. Baram, T.Z. et al.: Febrile seizures: an appropriate-aged model suitable for long-term studies. *Brain Res. Dev. Brain Res.* 98, 2, 265–270 (1997). [https://doi.org/10.1016/s0165-3806\(96\)00190-3](https://doi.org/10.1016/s0165-3806(96)00190-3).
30. Barnard, M. et al.: Whole Blood Analysis of Leukocyte-Platelet Aggregates. *Curr. Protoc. Cytom. Editor. Board J Paul Robinson Manag. Ed. Al. Chapter 6, Unit 6.15* (2003). <https://doi.org/10.1002/0471142956.cy0615s24>.
31. Baroni, D. et al.: A mutation of SCN1B associated with GEFS+ causes functional and maturation defects of the voltage-dependent sodium channel. *Hum. Mutat.* 39, 10, 1402–1415 (2018). <https://doi.org/10.1002/humu.23589>.
32. Bar-Or, D. et al.: Stress Hyperglycemia in Critically Ill Patients: Insight Into Possible Molecular Pathways. *Front. Med.* 6, 54 (2019). <https://doi.org/10.3389/fmed.2019.00054>.
33. Bartfai, T. et al.: Interleukin-1 system in CNS stress: seizures, fever, and neurotrauma. *Ann. N. Y. Acad. Sci.* 1113, 173–177 (2007). <https://doi.org/10.1196/annals.1391.022>.
34. Bath, P.M., Butterworth, R.J.: Platelet size: measurement, physiology and vascular disease. *Blood Coagul. Fibrinolysis Int. J. Haemost. Thromb.* 7, 2, 157–161 (1996).
35. Battaglia, D. et al.: Cognitive decline in Dravet syndrome: is there a cerebellar role? *Epilepsy Res.* 106, 1–2, 211–221 (2013). <https://doi.org/10.1016/j.eplepsyres.2013.03.012>.
36. Baulac, S. et al.: A novel locus for generalized epilepsy with febrile seizures plus in French families. *Arch. Neurol.* 65, 7, 943–951 (2008). <https://doi.org/10.1001/archneur.65.7.943>.
37. Baulac, S. et al.: Fever, genes, and epilepsy. *Lancet Neurol.* 3, 7, 421–430 (2004). [https://doi.org/10.1016/S1474-4422\(04\)00808-7](https://doi.org/10.1016/S1474-4422(04)00808-7).
38. Bechi, G. et al.: Pure haploinsufficiency for Dravet syndrome Na(V)1.1 (SCN1A) sodium channel truncating mutations. *Epilepsia.* 53, 1, 87–100 (2012). <https://doi.org/10.1111/j.1528-1167.2011.03346.x>.
39. Beckh, S. et al.: Differential regulation of three sodium channel messenger RNAs in the rat central nervous system during development. *EMBO J.* 8, 12, 3611–3616 (1989).
40. Beguin, Y.: Erythropoietin and platelet production. *Haematologica.* 84, 6, 541–547 (1999).
41. Behmanesh, F. et al.: Evaluation of Interleukin 1 β in Febrile Convulsion. *Iran. J. Allergy Asthma Immunol.* 336–339 (2012).

42. Behnke, O., Forer, A.: From megakaryocytes to platelets: platelet morphogenesis takes place in the bloodstream. *Eur. J. Haematol.* 60, S61, 3–23 (1998). <https://doi.org/10.1111/j.1600-0609.1998.tb01052.x>.
43. Bender, A.C. et al.: Cognitive Deficits Associated with Nav1.1 Alterations: Involvement of Neuronal Firing Dynamics and Oscillations. *PLOS ONE*. 11, 3, e0151538 (2016). <https://doi.org/10.1371/journal.pone.0151538>.
44. Bender, R.A. et al.: Localization of HCN1 Channels to Presynaptic Compartments: Novel Plasticity That May Contribute to Hippocampal Maturation. *J. Neurosci.* 27, 17, 4697–4706 (2007). <https://doi.org/10.1523/JNEUROSCI.4699-06.2007>.
- 395
45. Berg, A.T. et al.: A prospective study of recurrent febrile seizures. *N. Engl. J. Med.* 327, 16, 1122–1127 (1992). <https://doi.org/10.1056/NEJM199210153271603>.
46. Berg, A.T.: Are febrile seizures provoked by a rapid rise in temperature? *Am. J. Dis. Child.* 1960. 147, 10, 1101–1103 (1993). <https://doi.org/10.1001/archpedi.1993.02160340087020>.
47. Berg, A.T. et al.: Predictors of recurrent febrile seizures: a metaanalytic review. *J. Pediatr.* 116, 3, 329–337 (1990). [https://doi.org/10.1016/s0022-3476\(05\)82816-1](https://doi.org/10.1016/s0022-3476(05)82816-1).
48. Berg, A.T. et al.: Predictors of recurrent febrile seizures. A prospective cohort study. *Arch. Pediatr. Adolesc. Med.* 151, 4, 371–378 (1997). <https://doi.org/10.1001/ARCHPEDI.1997.02170410045006>.
49. Berg, A.T. et al.: Revised terminology and concepts for organization of seizures and epilepsies: Report of the ILAE Commission on Classification and Terminology, 2005–2009. *Epilepsia*. 51, 4, 676–685 (2010). <https://doi.org/10.1111/j.1528-1167.2010.02522.x>.
50. Berg, A.T. et al.: Risk factors for a first febrile seizure: a matched case-control study. *Epilepsia*. 36, 4, 334–341 (1995). <https://doi.org/10.1111/j.1528-1157.1995.tb01006.x>.
51. Berg, A.T., Shinnar, S.: Complex febrile seizures. *Epilepsia*. 37, 2, 126–133 (1996). <https://doi.org/10.1111/j.1528-1157.1996.tb00003.x>.
52. Berg, A.T., Shinnar, S.: Unprovoked seizures in children with febrile seizures: short-term outcome. *Neurology*. 47, 2, 562–568 (1996). <https://doi.org/10.1212/wnl.47.2.562>.
53. Berkvens, J.J.L. et al.: Autism and behavior in adult patients with Dravet syndrome (DS). *Epilepsy Behav.* 47, 11–16 (2015). <https://doi.org/10.1016/j.yebeh.2015.04.057>.
54. Bertelsen, E.N. et al.: Childhood Epilepsy, Febrile Seizures, and Subsequent Risk of ADHD. *Pediatrics*. 138, 2, e20154654 (2016). <https://doi.org/10.1542/peds.2015-4654>.
55. Bessman, J.D. et al.: Mean platelet volume. The inverse relation of platelet size and count in normal subjects, and an artifact of other particles. *Am. J. Clin. Pathol.* 76, 3, 289–293 (1981). <https://doi.org/10.1093/ajcp/76.3.289>.
56. Bethune, P. et al.: Which child will have a febrile seizure? *Am. J. Dis. Child.* 1960. 147, 1, 35–39 (1993). <https://doi.org/10.1001/archpedi.1993.02160250037013>.
57. Bharat, D.K. et al.: Association between iron deficiency anemia and febrile seizures. *Pediatr. Rev. Int. J. Pediatr. Res.* 2, 4, 41–46 (2015). <https://doi.org/10.17511/ijpr.2015.i04.02>.
58. Bhat, M.A. et al.: Role of SCN1A and SCN2A Gene Polymorphisms in Epilepsy Syndromes-A Study from India. *J. Neurol. Neurosci.* 9, 1, (2018). <https://doi.org/10.21767/2171-6625.1000238>.
59. Bhutia, T.D. et al.: Abnormalities in glucose homeostasis in critically ill children. *Pediatr. Crit. Care Med. J. Soc. Crit. Care Med. World Fed. Pediatr. Intensive Crit. Care Soc.* 14, 1, e16-25 (2013). <https://doi.org/10.1097/PCC.0b013e3182604998>.
60. Bisulli, F. et al.: SCN1A mutations in focal epilepsy with auditory features: widening the spectrum of GEFS plus. *Epileptic Disord. Int. Epilepsy J. Videotape*. 21, 2, 185–191 (2019). <https://doi.org/10.1684/epd.2019.1046>.
61. Boia, M. et al.: Noțiuni practice de puericultură /. Editura Victor Babeș, Timișoara (2019).

62. Bonanni, P. et al.: Generalized epilepsy with febrile seizures plus (GEFS+): clinical spectrum in seven Italian families unrelated to SCN1A, SCN1B, and GABRG2 gene mutations. *Epilepsia*. 45, 2, 149–158 (2004). <https://doi.org/10.1111/j.0013-9580.2004.04303.x>.
63. Bozzi, Y., Borrelli, E.: The role of dopamine signaling in epileptogenesis. *Front. Cell. Neurosci.* 7, 157 (2013). <https://doi.org/10.3389/fncel.2013.00157>.
64. Braekkan, S.K. et al.: Mean platelet volume is a risk factor for venous thromboembolism: the Tromsø Study, Tromsø, Norway. *J. Thromb. Haemost. JTH.* 8, 1, 157–162 (2010). <https://doi.org/10.1111/j.1538-7836.2009.03498.x>.
65. Brewster, A. et al.: Developmental Febrile Seizures Modulate Hippocampal Gene Expression of Hyperpolarization-Activated Channels in an Isoform- and Cell-Specific Manner. *J. Neurosci. Off. J. Soc. Neurosci.* 22, 4591–9 (2002). <https://doi.org/10.1523/JNEUROSCI.22-11-04591.2002>.
66. Brinkman, J.E., Sharma, S.: Respiratory Alkalosis. In: StatPearls. StatPearls Publishing, Treasure Island (FL) (2021).
67. Brooks, B.R., Adams, R.D.: Cerebrospinal fluid acid-base and lactate changes after seizures in unanesthetized man. I. Idiopathic seizures. *Neurology*. 25, 10, 935–942 (1975). <https://doi.org/10.1212/wnl.25.10.935>.
68. Brown, N.J. et al.: Vaccination, seizures and “vaccine damage.” *Curr. Opin. Neurol.* 20, 2, 181–187 (2007). <https://doi.org/10.1097/WCO.0b013e3280555160>.
- 396
69. Buckmaster, P.S., Dudek, F.E.: Network properties of the dentate gyrus in epileptic rats with hilar neuron loss and granule cell axon reorganization. *J. Neurophysiol.* 77, 5, 2685–2696 (1997). <https://doi.org/10.1152/jn.1997.77.5.2685>.
70. Budak, Y.U. et al.: The use of platelet indices, plateletcrit, mean platelet volume and platelet distribution width in emergency non-traumatic abdominal surgery: a systematic review. *Biochem. Medica*. 26, 2, 178–193 (2016). <https://doi.org/10.11613/BM.2016.020>.
71. Bullock, G.C. et al.: Iron control of erythroid development by a novel aconitase-associated regulatory pathway. *Blood*. 116, 1, 97–108 (2010). <https://doi.org/10.1182/blood-2009-10-251496>.
72. Burhanoglu, M. et al.: Hypozincaemia in febrile convulsion. *Eur. J. Pediatr.* 155, 6, 498–501 (1996). <https://doi.org/10.1007/BF01955189>.
73. Burstein, S. et al.: Thrombocytopoiesis in normal and sublethally irradiated dogs: response to human interleukin-6. *Blood*. 80, 2, 420–428 (1992). <https://doi.org/10.1182/blood.V80.2.420.420>.
74. Calkosiński, I. et al.: [Characterization of an inflammatory response]. *Postepy Hig. Med. Doswiadczałnej Online*. 63, 395–408 (2009).
75. Camfield, P. et al.: Chapter 2 - Antecedents and Risk Factors for Febrile Seizures. In: Baram, T.Z. and Shinnar, S. (eds.) *Febrile Seizures*. pp. 27–36 Academic Press, San Diego (2002). <https://doi.org/10.1016/B978-012078141-6/50004-4>.
76. Camfield, P. et al.: What types of epilepsy are preceded by febrile seizures? A population-based study of children. *Dev. Med. Child Neurol.* 36, 10, 887–892 (1994). <https://doi.org/10.1111/j.1469-8749.1994.tb11779.x>.
77. Camfield, P., Camfield, C.: Febrile seizures and genetic epilepsy with febrile seizures plus (GEFS+). *Epileptic Disord. Int. Epilepsy J. Videotape*. 17, 2, 124–133 (2015). <https://doi.org/10.1684/epd.2015.0737>.
78. Cangemi, R. et al.: Platelet Activation Is Associated with Myocardial Infarction in Patients with Pneumonia. *J. Am. Coll. Cardiol.* 64, 18, 1917–1925 (2014). <https://doi.org/10.1016/j.jacc.2014.07.985>.
79. Canpolat, M. et al.: Investigating the prevalence of febrile convulsion in Kayseri, Turkey: An assessment of the risk factors for recurrence of febrile convulsion and for development of epilepsy. *Seizure*. 55, 36–47 (2018). <https://doi.org/10.1016/j.seizure.2018.01.007>.

80. Capovilla, G. et al.: Recommendations for the management of “febrile seizures”: Ad Hoc Task Force of LICE Guidelines Commission. *Epilepsia*. 50 Suppl 1, 2–6 (2009). <https://doi.org/10.1111/j.1528-1167.2008.01963.x>.
81. Cappellari, A.M. et al.: Predictive value of EEG for febrile seizure recurrence. *Brain Dev.* 40, 4, 311–315 (2018). <https://doi.org/10.1016/j.braindev.2017.12.004>.
82. Carlioglu, A. et al.: Increased mean platelet volume in papillary thyroid cancer. *Endocr. Abstr.* 35, (2014). <https://doi.org/10.1530/endoabs.35.P1114>.
83. Carranza Rojo, D. et al.: De novo SCN1A mutations in migrating partial seizures of infancy. *Neurology*. 77, 4, 380–383 (2011). <https://doi.org/10.1212/WNL.0b013e318227046d>.
84. Cartmell, T. et al.: Brain sites of action of endogenous interleukin-1 in the febrile response to localized inflammation in the rat. *J. Physiol.* 518 (Pt 2), 585–594 (1999). <https://doi.org/10.1111/j.1469-7793.1999.0585p.x>.
85. Cartmell, T. et al.: Interleukin-1 mediates a rapid inflammatory response after injection of adenoviral vectors into the brain. *J. Neurosci. Off. J. Soc. Neurosci.* 19, 4, 1517–1523 (1999).
86. Carvalho-Tavares, J. et al.: A role for platelets and endothelial selectins in tumor necrosis factor-alpha-induced leukocyte recruitment in the brain microvasculature. *Circ. Res.* 87, 12, 1141–1148 (2000). <https://doi.org/10.1161/01.res.87.12.1141>.
87. Carvill, G.L. et al.: Aberrant Inclusion of a Poison Exon Causes Dravet Syndrome and Related SCN1A-Associated Genetic Epilepsies. *Am. J. Hum. Genet.* 103, 6, 1022–1029 (2018). <https://doi.org/10.1016/j.ajhg.2018.10.023>.
88. Carvill, G.L. et al.: GABRA1 and STXBP1: novel genetic causes of Dravet syndrome. *Neurology*. 82, 14, 1245–1253 (2014). <https://doi.org/10.1212/WNL.0000000000000291>.
89. Caserta, M.T. et al.: Primary human herpesvirus 7 infection: a comparison of human herpesvirus 7 and human herpesvirus 6 infections in children. *J. Pediatr.* 133, 3, 386–389 (1998). [https://doi.org/10.1016/s0022-3476\(98\)70275-6](https://doi.org/10.1016/s0022-3476(98)70275-6).
- 397
90. Castro-Gago, M. et al.: Cerebrospinal fluid purine metabolites and pyrimidine bases after brief febrile convulsions. *Epilepsia*. 36, 5, 471–474 (1995). <https://doi.org/10.1111/j.1528-1157.1995.tb00488.x>.
91. Catarino, C.B. et al.: Dravet syndrome as epileptic encephalopathy: evidence from long-term course and neuropathology. *Brain J. Neurol.* 134, Pt 10, 2982–3010 (2011). <https://doi.org/10.1093/brain/awr129>.
92. Catterall, W.A.: Chapter 4 - Dravet Syndrome: A Sodium Channel Interneuronopathy. In: Pitt, G.S. (ed.) *Ion Channels in Health and Disease*. pp. 85–111 Academic Press, Boston (2016). <https://doi.org/10.1016/B978-0-12-802002-9.00004-2>.
93. Catterall, W.A.: Dravet Syndrome: A Sodium Channel Interneuronopathy. *Curr. Opin. Physiol.* 2, 42–50 (2018). <https://doi.org/10.1016/j.cophys.2017.12.007>.
94. Cendes, F., Andermann, F.: Chapter 6 - Do Febrile Seizures Promote Temporal Lobe Epilepsy? Retrospective Studies. In: Baram, T.Z. and Shinnar, S. (eds.) *Febrile Seizures*. pp. 77–86 Academic Press, San Diego (2002). <https://doi.org/10.1016/B978-012078141-6/50008-1>.
95. Cestèle, S. et al.: Nonfunctional NaV1.1 familial hemiplegic migraine mutant transformed into gain of function by partial rescue of folding defects. *Proc. Natl. Acad. Sci. U. S. A.* 110, 43, 17546–17551 (2013). <https://doi.org/10.1073/pnas.1309827110>.
96. Cetica, V. et al.: Clinical and genetic factors predicting Dravet syndrome in infants with SCN1A mutations. *Neurology*. 88, 11, 1037–1044 (2017). <https://doi.org/10.1212/WNL.0000000000003716>.
97. Ceulemans, B.P.G.M. et al.: Clinical correlations of mutations in the SCN1A gene: from febrile seizures to severe myoclonic epilepsy in infancy. *Pediatr. Neurol.* 30, 4, 236–243 (2004). <https://doi.org/10.1016/j.pediatrneurol.2003.10.012>.

98. Chandrashekhar, V.: Plateletcrit as a Screening Tool for Detection of Platelet Quantitative Disorders. *J. Hematol.* 2, 22–26 (2013). <https://doi.org/10.4021/jh70w>.
99. Chapman, A.G. et al.: Cerebral metabolic changes during prolonged epileptic seizures in rats. *J. Neurochem.* 28, 5, 1025–1035 (1977). <https://doi.org/10.1111/j.1471-4159.1977.tb10665.x>.
100. Chen, K. et al.: Long-Term Plasticity of Endocannabinoid Signaling Induced by Developmental Febrile Seizures. *Neuron.* 39, 4, 599–611 (2003). [https://doi.org/10.1016/S0896-6273\(03\)00499-9](https://doi.org/10.1016/S0896-6273(03)00499-9).
101. Chen, K. et al.: Persistently modified h-channels after complex febrile seizures convert the seizure-induced enhancement of inhibition to hyperexcitability. *Nat. Med.* 7, 3, 331–337 (2001). <https://doi.org/10.1038/85480>.
102. Chen, K. et al.: Prevention of plasticity of endocannabinoid signaling inhibits persistent limbic hyperexcitability caused by developmental seizures. *J. Neurosci. Off. J. Soc. Neurosci.* 27, 1, 46–58 (2007). <https://doi.org/10.1523/JNEUROSCI.3966-06.2007>.
103. Chen, Q. et al.: Association between interleukin-6 gene polymorphisms and febrile seizure risk: A meta-analysis. *Medicine (Baltimore)*. 98, 39, e17167 (2019). <https://doi.org/10.1097/MD.00000000000017167>.
104. Chen, R. et al.: Analysis of cytokines and trace elements in children with febrile seizures. *Transl. Pediatr.* 9, 6, 809–817 (2020). <https://doi.org/10.21037/tp-20-398>.
105. Chen, Y.H. et al.: Cloning, distribution and functional analysis of the type III sodium channel from human brain. *Eur. J. Neurosci.* 12, 12, 4281–4289 (2000).
106. Chen, Y.-J. et al.: Electrophysiological Differences between the Same Pore Region Mutation in SCN1A and SCN3A. *Mol. Neurobiol.* 51, 3, 1263–1270 (2015). <https://doi.org/10.1007/s12035-014-8802-x>.
107. Chesler, M., Kaila, K.: Modulation of pH by neuronal activity. *Trends Neurosci.* 15, 10, 396–402 (1992). [https://doi.org/10.1016/0166-2236\(92\)90191-a](https://doi.org/10.1016/0166-2236(92)90191-a).
108. Chiofalo, N. et al.: Prevalence of epilepsy in children of Melipilla, Chile. *Epilepsia.* 20, 3, 261–266 (1979). <https://doi.org/10.1111/j.1528-1157.1979.tb04803.x>.
109. Chiron, C.: Stiripentol for the treatment of seizures associated with Dravet syndrome. *Expert Rev. Neurother.* 19, 4, 301–310 (2019). <https://doi.org/10.1080/14737175.2019.1593142>.
110. Cho, J.H. et al.: The IL-1B Genetic Polymorphism Is Associated with Aspirin-Induced Peptic Ulcers in a Korean Ethnic Group. *Gut Liver.* 10, 3, 362–368 (2016). <https://doi.org/10.5009/gnl15129>.
111. Choi, B.H.: Oxygen, antioxidants and brain dysfunction. *Yonsei Med. J.* 34, 1, 1–10 (1993). <https://doi.org/10.3349/ymj.1993.34.1.1>.
- 398
112. Choi, J. et al.: Association Analysis of Interleukin-1 β , Interleukin-6, and HMGB1 Variants with Postictal Serum Cytokine Levels in Children with Febrile Seizure and Generalized Epilepsy with Febrile Seizure Plus. *J. Clin. Neurol. Seoul Korea.* 15, 4, 555–563 (2019). <https://doi.org/10.3988/jcn.2019.15.4.555>.
113. Choi, J. et al.: Increased levels of HMGB1 and pro-inflammatory cytokines in children with febrile seizures. *J. Neuroinflammation.* 8, 135 (2011). <https://doi.org/10.1186/1742-2094-8-135>.
114. Chou, H.-F. et al.: Utility of laboratory tests for children in the emergency department with a first seizure. *Pediatr. Emerg. Care.* 27, 12, 1142–1145 (2011). <https://doi.org/10.1097/PEC.0b013e31823aba17>.
115. Chou, I.-C. et al.: Interleukin (IL)-1beta, IL-1 receptor antagonist, IL-6, IL-8, IL-10, and tumor necrosis factor alpha gene polymorphisms in patients with febrile seizures. *J. Clin. Lab. Anal.* 24, 3, 154–159 (2010). <https://doi.org/10.1002/jcla.20374>.
116. Choy, M. et al.: Inflammatory processes, febrile seizures, and subsequent epileptogenesis. *Epilepsy Curr.* 14, 1 Suppl, 15–22 (2014). <https://doi.org/10.5698/1535-7511-14.s2.15>.

117. Chung, B., Wong, V.: Relationship between five common viruses and febrile seizure in children. *Arch. Dis. Child.* 92, 7, 589–593 (2007). <https://doi.org/10.1136/adc.2006.110221>.
118. Chung, S.: Febrile seizures. *Korean J. Pediatr.* 57, 9, 384–395 (2014). <https://doi.org/10.3345/kjp.2014.57.9.384>.
119. Claes, L. et al.: De novo mutations in the sodium-channel gene SCN1A cause severe myoclonic epilepsy of infancy. *Am. J. Hum. Genet.* 68, 6, 1327–1332 (2001). <https://doi.org/10.1086/320609>.
120. Cody, C.L. et al.: Nature and rates of adverse reactions associated with DTP and DT immunizations in infants and children. *Pediatrics.* 68, 5, 650–660 (1981).
121. Connolly, M.B.: Dravet Syndrome: Diagnosis and Long-Term Course. *Can. J. Neurol. Sci. J. Can. Sci. Neurol.* 43 Suppl 3, S3-8 (2016). <https://doi.org/10.1017/cjn.2016.243>.
122. Conti, B. et al.: Cytokines and fever. *Front. Biosci. J. Virtual Libr.* 9, 1433–1449 (2004). <https://doi.org/10.2741/1341>.
123. Coryell, M.W. et al.: Targeting ASIC1a reduces innate fear and alters neuronal activity in the fear circuit. *Biol. Psychiatry.* 62, 10, 1140–1148 (2007). <https://doi.org/10.1016/j.biopsych.2007.05.008>.
124. Costa Leite, T. et al.: Lactate favours the dissociation of skeletal muscle 6-phosphofructo-1-kinase tetramers down-regulating the enzyme and muscle glycolysis. *Biochem. J.* 408, Pt 1, 123–130 (2007). <https://doi.org/10.1042/BJ20070687>.
125. Costea, R.-M. et al.: Antioxidanții (resveratrolul) - posibile terapii adjuvante în profilaxia recurenței convulsiilor febrile. *Bul. Perinatol.* 73, 1, 54–59 (2017).
126. Costea, R.M. et al.: Stress Hyperglycemia as Predictive Factor of Recurrence in Children with Febrile Seizures. *Brain Sci.* 10, 3, (2020). <https://doi.org/10.3390/brainsci10030131>.
127. Costea, R.M. et al.: The Opportunity of the Electroencephalography in the Diagnostic Approach of the Febrile Seizures. *Acta Medica Transilv.* 21, 3, 50–53 (2016).
128. Costello, D.A. et al.: Interleukin-1alpha and HMGB1 mediate hippocampal dysfunction in SIGIRR-deficient mice. *J. Neurosci. Off. J. Soc. Neurosci.* 31, 10, 3871–3879 (2011). <https://doi.org/10.1523/JNEUROSCI.6676-10.2011>.
129. Cuestas, E.: Is routine EEG helpful in the management of complex febrile seizures? *Arch. Dis. Child.* 89, 3, 290 (2004). <https://doi.org/10.1136/adc.2003.048447>.
130. Cui, X. et al.: A novel SCN1A missense mutation causes generalized epilepsy with febrile seizures plus in a Chinese family. *Neurosci. Lett.* 503, 1, 27–30 (2011). <https://doi.org/10.1016/j.neulet.2011.08.001>.
131. Cummins, T.R. et al.: Nav1.3 sodium channels: rapid repriming and slow closed-state inactivation display quantitative differences after expression in a mammalian cell line and in spinal sensory neurons. *J. Neurosci. Off. J. Soc. Neurosci.* 21, 16, 5952–5961 (2001).
132. Cummins, T.R., Waxman, S.G.: Downregulation of tetrodotoxin-resistant sodium currents and upregulation of a rapidly repriming tetrodotoxin-sensitive sodium current in small spinal sensory neurons after nerve injury. *J. Neurosci. Off. J. Soc. Neurosci.* 17, 10, 3503–3514 (1997).
133. De Onis, M.: WHO Child Growth Standards - Length/Height-for-age, Weight-for-age, Weight-for-length, Weight-for-height and Body Mass Index-for age: Methods and Development. World Health Organization, Geneva (2006).
- 399
134. De Simoni, M.G. et al.: Inflammatory cytokines and related genes are induced in the rat hippocampus by limbic status epilepticus. *Eur. J. Neurosci.* 12, 7, 2623–2633 (2000). <https://doi.org/10.1046/j.1460-9568.2000.00140.x>.
135. Deng, L. et al.: Postvaccination Febrile Seizure Severity and Outcome. *Pediatrics.* 143, 5, (2019). <https://doi.org/10.1542/peds.2018-2120>.

136. Depienne, C. et al.: Mechanisms for variable expressivity of inherited SCN1A mutations causing Dravet syndrome. *J. Med. Genet.* 47, 6, 404–410 (2010). <https://doi.org/10.1136/jmg.2009.074328>.
137. Depienne, C. et al.: Spectrum of SCN1A gene mutations associated with Dravet syndrome: analysis of 333 patients. *J. Med. Genet.* 46, 3, 183–191 (2009). <https://doi.org/10.1136/jmg.2008.062323>.
138. Di Marco, R. et al.: Stabilometry in patients with Dravet Syndrome to quantitatively assess ataxia: A preliminary study. *Gait Posture.* 66, S15 (2018). <https://doi.org/10.1016/j.gaitpost.2018.07.123>.
139. Diachinsky, M.: Fever. In: Mahmoud, S.H. (ed.) *Patient assessment in clinical pharmacy: a comprehensive guide.* pp. 121–132 Springer (2019).
140. Dibbens, L.M. et al.: GABRD encoding a protein for extra- or peri-synaptic GABA_A receptors is a susceptibility locus for generalized epilepsies. *Hum. Mol. Genet.* 13, 13, 1315–1319 (2004). <https://doi.org/10.1093/hmg/ddh146>.
141. Dignass, A. et al.: Limitations of Serum Ferritin in Diagnosing Iron Deficiency in Inflammatory Conditions. *Int. J. Chronic Dis.* 2018, 9394060 (2018). <https://doi.org/10.1155/2018/9394060>.
142. Dinarello, C.A.: Biologic basis for interleukin-1 in disease. *Blood.* 87, 6, 2095–2147 (1996).
143. Djémié, T. et al.: Pitfalls in genetic testing: the story of missed SCN1A mutations. *Mol. Genet. Genomic Med.* 4, 4, 457–464 (2016). <https://doi.org/10.1002/mgg3.217>.
144. Djordjevic, D. et al.: Neutrophil-to-Lymphocyte Ratio, Monocyte-to-Lymphocyte Ratio, Platelet-to-Lymphocyte Ratio, and Mean Platelet Volume-to-Platelet Count Ratio as Biomarkers in Critically Ill and Injured Patients: Which Ratio to Choose to Predict Outcome and Nature of Bacteremia?, <https://www.hindawi.com/journals/mi/2018/3758068/>, last accessed 2021/03/13. <https://doi.org/10.1155/2018/3758068>.
145. Dogan, I. et al.: Relationship between serum neutrophil count and infarct size in patients with acute myocardial infarction. *Nucl. Med. Commun.* 30, 10, 797–801 (2009). <https://doi.org/10.1097/MNM.0b013e32832e3a16>.
146. Donati, D. et al.: Detection of human herpesvirus-6 in mesial temporal lobe epilepsy surgical brain resections. *Neurology.* 61, 10, 1405–1411 (2003). <https://doi.org/10.1212/01.wnl.0000094357.10782.f9>.
147. Dong, Z.-F. et al.: Transcription of the Human Sodium Channel SCN1A Gene Is Repressed by a Scaffolding Protein RACK1. *Mol. Neurobiol.* 50, 2, 438–448 (2014). <https://doi.org/10.1007/s12035-014-8633-9>.
148. Doose, H., Maurer, A.: Seizure risk in offspring of individuals with a history of febrile convulsions. *Eur. J. Pediatr.* 156, 6, 476–481 (1997). <https://doi.org/10.1007/s004310050643>.
149. Dravet, C., Oguni, H.: Dravet syndrome (severe myoclonic epilepsy in infancy). *Handb. Clin. Neurol.* 111, 627–633 (2013). <https://doi.org/10.1016/B978-0-444-52891-9.00065-8>.
150. Dreier, J.W. et al.: Evaluation of Long-term Risk of Epilepsy, Psychiatric Disorders, and Mortality Among Children With Recurrent Febrile Seizures: A National Cohort Study in Denmark. *JAMA Pediatr.* 173, 12, 1164–1170 (2019). <https://doi.org/10.1001/jamapediatrics.2019.3343>.
151. Dubé, C. et al.: Interleukin-1beta contributes to the generation of experimental febrile seizures. *Ann. Neurol.* 57, 1, 152–155 (2005). <https://doi.org/10.1002/ana.20358>.
152. Dubé, C. et al.: Temporal lobe epilepsy after experimental prolonged febrile seizures: prospective analysis. *Brain J. Neurol.* 129, Pt 4, 911–922 (2006). <https://doi.org/10.1093/brain/awl018>.
153. Dubé, C.M. et al.: Epileptogenesis provoked by prolonged experimental febrile seizures: mechanisms and biomarkers. *J. Neurosci. Off. J. Soc. Neurosci.* 30, 22, 7484–7494 (2010). <https://doi.org/10.1523/JNEUROSCI.0551-10.2010>.

154. Dubé, C.M. et al.: Febrile seizures: mechanisms and relationship to epilepsy. *Brain Dev.* 31, 5, 366–371 (2009). <https://doi.org/10.1016/j.braindev.2008.11.010>.
155. Duffy, J. et al.: Febrile Seizure Risk after Vaccination in Children One to Five Months of Age. *Pediatr. Neurol.* 76, 72–78 (2017). <https://doi.org/10.1016/j.pediatrneurol.2017.08.005>.
- 400
156. Duflocq, A. et al.: Nav1.1 is predominantly expressed in nodes of Ranvier and axon initial segments. *Mol. Cell. Neurosci.* 39, 2, 180–192 (2008). <https://doi.org/10.1016/j.mcn.2008.06.008>.
157. Dungan, K.M. et al.: Stress hyperglycaemia. *Lancet Lond. Engl.* 373, 9677, 1798–1807 (2009). [https://doi.org/10.1016/S0140-6736\(09\)60553-5](https://doi.org/10.1016/S0140-6736(09)60553-5).
158. Durá-Travé, T. et al.: Epilepsy in Children in Navarre, Spain: Epileptic Seizure Types and Epileptic Syndromes. *J. Child Neurol.* 22, 7, 823–828 (2007). <https://doi.org/10.1177/0883073807304207>.
159. Dzhala, V.I. et al.: NKCC1 transporter facilitates seizures in the developing brain. *Nat. Med.* 11, 11, 1205–1213 (2005). <https://doi.org/10.1038/nm1301>.
160. Ebach, K. et al.: SCN1A mutation analysis in myoclonic astatic epilepsy and severe idiopathic generalized epilepsy of infancy with generalized tonic-clonic seizures. *Neuropediatrics.* 36, 3, 210–213 (2005). <https://doi.org/10.1055/s-2005-865607>.
161. al-Eissa, Y.A.: Febrile seizures: rate and risk factors of recurrence. *J. Child Neurol.* 10, 4, 315–319 (1995). <https://doi.org/10.1177/088307389501000415>.
162. El-Harith, E.-H.A. et al.: Familial thrombocytosis caused by the novel germ-line mutation p.Pro106Leu in the MPL gene. *Br. J. Haematol.* 144, 2, 185–194 (2009). <https://doi.org/10.1111/j.1365-2141.2008.07430.x>.
163. Elmas, B. et al.: Thiol/disulfide homeostasis as a novel indicator of oxidative stress in children with simple febrile seizures. *Neurol. Sci. Off. J. Ital. Neurol. Soc. Ital. Soc. Clin. Neurophysiol.* 38, 11, 1969–1975 (2017). <https://doi.org/10.1007/s10072-017-3087-2>.
164. El-Masry, H.M.A. et al.: Metabolic profile of oxidative stress and trace elements in febrile seizures among children. *Metab. Brain Dis.* 33, 5, 1509–1515 (2018). <https://doi.org/10.1007/s11011-018-0258-7>.
165. Emsley, H.C.A. et al.: Variations in inflammation-related genes may be associated with childhood febrile seizure susceptibility. *Seizure.* 23, 6, 457–461 (2014). <https://doi.org/10.1016/j.seizure.2014.03.006>.
166. Engdahl, E. et al.: Human Herpesvirus 6B Induces Hypomethylation on Chromosome 17p13.3, Correlating with Increased Gene Expression and Virus Integration. *J. Virol.* 91, 11, (2017). <https://doi.org/10.1128/JVI.02105-16>.
167. England, S., de Groot, M.J.: Subtype-selective targeting of voltage-gated sodium channels. *Br. J. Pharmacol.* 158, 6, 1413–1425 (2009). <https://doi.org/10.1111/j.1476-5381.2009.00437.x>.
168. Epstein, L.G. et al.: Human herpesvirus 6 and 7 in febrile status epilepticus: the FEBSTAT study. *Epilepsia.* 53, 9, 1481–1488 (2012). <https://doi.org/10.1111/j.1528-1167.2012.03542.x>.
169. Erikson, K. et al.: Iron deficiency decreases dopamine D-1 and D-2 receptors in rat brain. *Pharmacol. Biochem. Behav.* 69, 409–18 (2001). [https://doi.org/10.1016/S0091-3057\(01\)00563-9](https://doi.org/10.1016/S0091-3057(01)00563-9).
170. Escayg, A. et al.: A novel SCN1A mutation associated with generalized epilepsy with febrile seizures plus--and prevalence of variants in patients with epilepsy. *Am. J. Hum. Genet.* 68, 4, 866–873 (2001). <https://doi.org/10.1086/319524>.
171. Escayg, A. et al.: Mutations of SCN1A, encoding a neuronal sodium channel, in two families with GEFS+2. *Nat. Genet.* 24, 4, 343–345 (2000). <https://doi.org/10.1038/74159>.
172. van Esch, A. et al.: Family history and recurrence of febrile seizures. *Arch. Dis. Child.* 70, 5, 395–399 (1994). <https://doi.org/10.1136/adc.70.5.395>.

173. Eschbach, K., Knupp, K.G.: Stiripentol for the treatment of seizures in Dravet syndrome. *Expert Rev. Clin. Pharmacol.* 12, 5, 379–388 (2019). <https://doi.org/10.1080/17512433.2019.1605904>.
174. Esiri, M.M., Kennedy, P.G.E.: Virus diseases of the nervous system. In: Graham, D.I. and Lantos, P.L. (eds.) *Greenfield's Neuropathology*. pp. 3–63 Edward Arnold, London (1997).
175. Eskandarifar, A. et al.: The risk factors in children with simple and complex febrile seizures: An epidemiological study. *Int. J. Pediatr.* 5, 5137–5144 (2017). <https://doi.org/10.22038/ijp.2017.22000.1840>.
176. Esposito, K. et al.: Inflammatory cytokine concentrations are acutely increased by hyperglycemia in humans: role of oxidative stress. *Circulation.* 106, 16, 2067–2072 (2002). <https://doi.org/10.1161/01.cir.0000034509.14906.ae>.
177. Esposito, L. et al.: Large-scale analysis of viral nucleic acid spectrum in temporal lobe epilepsy biopsies. *Epilepsia.* 56, 2, 234–243 (2015). <https://doi.org/10.1111/epi.12890>.
- 401
178. Estacion, M. et al.: A sodium channel mutation linked to epilepsy increases ramp and persistent current of Nav1.3 and induces hyperexcitability in hippocampal neurons. *Exp. Neurol.* 224, 2, 362–368 (2010). <https://doi.org/10.1016/j.expneurol.2010.04.012>.
179. Esteki-Zadeh, A. et al.: Human cytomegalovirus infection is sensitive to the host cell DNA methylation state and alters global DNA methylation capacity. *Epigenetics.* 7, 6, 585–593 (2012). <https://doi.org/10.4161/epi.20075>.
180. Fallah, R. et al.: Efficacy of zinc sulfate supplement on febrile seizure recurrence prevention in children with normal serum zinc level: A randomised clinical trial. *Nutr. Burbank Los Angel. Cty. Calif.* 31, 11–12, 1358–1361 (2015). <https://doi.org/10.1016/j.nut.2015.05.024>.
181. Fan, C. et al.: Early-onset familial hemiplegic migraine due to a novel SCN1A mutation. *Cephalgia Int. J. Headache.* 36, 13, 1238–1247 (2016). <https://doi.org/10.1177/0333102415608360>.
182. Farrington, P. et al.: A new method for active surveillance of adverse events from diphtheria/tetanus/pertussis and measles/mumps/rubella vaccines. *Lancet Lond. Engl.* 345, 8949, 567–569 (1995). [https://doi.org/10.1016/s0140-6736\(95\)90471-9](https://doi.org/10.1016/s0140-6736(95)90471-9).
183. Fasano, A. et al.: Antecollis and levodopa-responsive parkinsonism are late features of Dravet syndrome. *Neurology.* 82, 24, 2250–2251 (2014). <https://doi.org/10.1212/WNL.0000000000000521>.
184. Fattorusso, V. et al.: Non-Diabetic Hyperglycemia in the Pediatric Age: Why, How, and When to Treat? *Curr. Diab. Rep.* 18, 12, 140 (2018). <https://doi.org/10.1007/s11892-018-1115-0>.
185. Feenstra, B. et al.: Common variants associated with general and MMR vaccine-related febrile seizures. *Nat. Genet.* 46, 12, 1274–1282 (2014). <https://doi.org/10.1038/ng.3129>.
186. Feng, B. et al.: Transient increase of interleukin-1 β after prolonged febrile seizures promotes adult epileptogenesis through long-lasting upregulating endocannabinoid signaling. *Sci. Rep.* 6, 21931 (2016). <https://doi.org/10.1038/srep21931>.
187. Feng, B., Chen, Z.: Generation of Febrile Seizures and Subsequent Epileptogenesis. *Neurosci. Bull.* 32, 5, 481–492 (2016). <https://doi.org/10.1007/s12264-016-0054-5>.
188. Ferrari, S.L. et al.: Two promoter polymorphisms regulating interleukin-6 gene expression are associated with circulating levels of C-reactive protein and markers of bone resorption in postmenopausal women. *J. Clin. Endocrinol. Metab.* 88, 1, 255–259 (2003). <https://doi.org/10.1210/jc.2002-020092>.
189. Fetveit, A.: Assessment of febrile seizures in children. *Eur. J. Pediatr.* 167, 1, 17–27 (2008). <https://doi.org/10.1007/s00431-007-0577-x>.
190. Fischler, M.P., Reinhart, W.H.: [Fever: friend or enemy?]. *Schweiz. Med. Wochenschr.* 127, 20, 864–870 (1997).

191. Flamand, L. et al.: Human herpesvirus 6 induces interleukin-1 beta and tumor necrosis factor alpha, but not interleukin-6, in peripheral blood mononuclear cell cultures. *J. Virol.* 65, 9, 5105–5110 (1991). <https://doi.org/10.1128/JVI.65.9.5105-5110.1991>.
192. Forsgren, L. et al.: A prospective incidence study of febrile convulsions. *Acta Paediatr. Scand.* 79, 5, 550–557 (1990). <https://doi.org/10.1111/j.1651-2227.1990.tb11510.x>.
193. Fotheringham, J. et al.: Association of human herpesvirus-6B with mesial temporal lobe epilepsy. *PLoS Med.* 4, 5, e180 (2007). <https://doi.org/10.1371/journal.pmed.0040180>.
194. Fountain-Capal, J.K. et al.: When should clinicians order genetic testing for Dravet syndrome? *Pediatr. Neurol.* 45, 5, 319–323 (2011). <https://doi.org/10.1016/j.pediatrneurol.2011.08.001>.
195. Frampton, J.E.: Stiripentol: A Review in Dravet Syndrome. *Drugs.* 79, 16, 1785–1796 (2019). <https://doi.org/10.1007/s40265-019-01204-y>.
196. Francis, J.R. et al.: An observational study of febrile seizures: the importance of viral infection and immunization. *BMC Pediatr.* 16, 1, 202 (2016). <https://doi.org/10.1186/s12887-016-0740-5>.
197. Frantzen, E. et al.: Longitudinal EEG and clinical study of children with febrile convulsions. *Electroencephalogr. Clin. Neurophysiol.* 24, 3, 197–212 (1968). [https://doi.org/10.1016/0013-4694\(68\)90001-1](https://doi.org/10.1016/0013-4694(68)90001-1).
198. Frasier, C.R. et al.: Channelopathy as a SUDEP Biomarker in Dravet Syndrome Patient-Derived Cardiac Myocytes. *Stem Cell Rep.* 11, 3, 626–634 (2018). <https://doi.org/10.1016/j.stemcr.2018.07.012>.
199. Frelinger, A.L. et al.: Platelet-rich plasma stimulated by pulse electric fields: Platelet activation, procoagulant markers, growth factor release and cell proliferation. *Platelets.* 27, 2, 128–135 (2016). <https://doi.org/10.3109/09537104.2015.1048214>.
- 402
200. French, J.A. et al.: Characteristics of medial temporal lobe epilepsy: I. Results of history and physical examination. *Ann. Neurol.* 34, 6, 774–780 (1993). <https://doi.org/10.1002/ana.410340604>.
201. Freund, T.F. et al.: Simultaneous recording of local electrical activity, partial oxygen tension and temperature in the rat hippocampus with a chamber-type microelectrode. Effects of anaesthesia, ischemia and epilepsy. *Neuroscience.* 28, 3, 539–549 (1989). [https://doi.org/10.1016/0306-4522\(89\)90003-1](https://doi.org/10.1016/0306-4522(89)90003-1).
202. Frode Svartdal, Tord Mortensen: Effects of Reinforcer Value on Sensitivity to Non-Verbal Operant Contingencies in Humans. *Q. J. Exp. Psychol. Sect. A.* 46, 2, 347–364 (1993). <https://doi.org/10.1080/14640749308401050>.
203. Fujiwara, T. et al.: Mutations of sodium channel alpha subunit type 1 (SCN1A) in intractable childhood epilepsies with frequent generalized tonic-clonic seizures. *Brain J. Neurol.* 126, Pt 3, 531–546 (2003). <https://doi.org/10.1093/brain/awg053>.
204. Fukuma, G. et al.: Mutations of neuronal voltage-gated Na⁺ channel alpha 1 subunit gene SCN1A in core severe myoclonic epilepsy in infancy (SMEI) and in borderline SMEI (SMEB). *Epilepsia.* 45, 2, 140–148 (2004). <https://doi.org/10.1111/j.0013-9580.2004.15103.x>.
205. Fung, E.L.W. et al.: Value of EEG in management of complex febrile convolution. *Eur. J. Paediatr. Neurol.* 21, e99 (2017). <https://doi.org/10.1016/j.ejpn.2017.04.738>.
206. Gaily, E. et al.: Dravet syndrome: new potential genetic modifiers, imaging abnormalities, and ictal findings. *Epilepsia.* 54, 9, 1577–1585 (2013). <https://doi.org/10.1111/epi.12256>.
207. Gajecka, M.: Unrevealed mosaicism in the next-generation sequencing era. *Mol. Genet. Genomics MGG.* 291, 2, 513–530 (2016). <https://doi.org/10.1007/s00438-015-1130-7>.
208. Gallentine, W.B. et al.: Plasma cytokines associated with febrile status epilepticus in children: A potential biomarker for acute hippocampal injury. *Epilepsia.* 58, 6, 1102–1111 (2017). <https://doi.org/10.1111/epi.13750>.

209. Ganesh, R., Janakiraman, L.: Serum zinc levels in children with simple febrile seizure. *Clin. Pediatr. (Phila.)*. 47, 2, 164–166 (2008). <https://doi.org/10.1177/0009922807306165>.
210. Gao, L. et al.: Prognostic value of combination of preoperative platelet count and mean platelet volume in patients with resectable non-small cell lung cancer. *Oncotarget*. 8, 9, 15632–15641 (2017). <https://doi.org/10.18632/oncotarget.14921>.
211. Gargus, J.J., Tournay, A.: Novel mutation confirms seizure locus SCN1A is also familial hemiplegic migraine locus FHM3. *Pediatr. Neurol.* 37, 6, 407–410 (2007). <https://doi.org/10.1016/j.pediatrneurol.2007.06.016>.
212. Gasparyan, A.Y. et al.: Mean platelet volume: a link between thrombosis and inflammation? *Curr. Pharm. Des.* 17, 1, 47–58 (2011). <https://doi.org/10.2174/138161211795049804>.
213. Gataullina, S., Dulac, O.: From genotype to phenotype in Dravet disease. *Seizure*. 44, 58–64 (2017). <https://doi.org/10.1016/j.seizure.2016.10.014>.
214. Gatti, S. et al.: Mechanisms of Fever and Febrile Seizures: Putative Role of the Interleukin-1 System. In: *Febrile Seizures*. pp. 169–188 (2002).
215. Geier, D.A., Geier, M.R.: An evaluation of serious neurological disorders following immunization: a comparison of whole-cell pertussis and acellular pertussis vaccines. *Brain Dev.* 26, 5, 296–300 (2004). [https://doi.org/10.1016/S0387-7604\(03\)00169-4](https://doi.org/10.1016/S0387-7604(03)00169-4).
216. Gennaro, E. et al.: Somatic and germline mosaisms in severe myoclonic epilepsy of infancy. *Biochem. Biophys. Res. Commun.* 341, 2, 489–493 (2006). <https://doi.org/10.1016/j.bbrc.2005.12.209>.
217. Georgieff, M.K.: Iron in the Brain: Its Role in Development and Injury. *NeoReviews*. 7, 7, e344–e352 (2006). <https://doi.org/10.1542/neo.7-7-e344>.
218. Ghasemi, F. et al.: Iron-deficiency Anemia in Children with Febrile Seizure: A Case-Control Study. *Iran. J. Child Neurol.* 8, 2, 38–44 (2014).
219. Gholipoor, P. et al.: Prenatal stress potentiates febrile seizure and leads to long-lasting increase in cortisol blood levels in children under 2years old. *Epilepsy Behav.* 72, 22–27 (2017). <https://doi.org/10.1016/j.yebeh.2017.04.021>.
220. Giacomini, A. et al.: Platelet count and parameters determined by the Bayer ADVIA 120 in reference subjects and patients. *Clin. Lab. Haematol.* 23, 3, 181–186 (2001). <https://doi.org/10.1046/j.1365-2257.2001.00391.x>.
221. Gitiaux, C. et al.: Motor neuropathy contributes to crouching in patients with Dravet syndrome. *Neurology*. 87, 3, 277–281 (2016). <https://doi.org/10.1212/WNL.0000000000002859>.
- 403
222. Goksugur, S.B. et al.: Neutrophil-to-lymphocyte ratio and red blood cell distribution width is a practical predictor for differentiation of febrile seizure types. *Eur. Rev. Med. Pharmacol. Sci.* 18, 22, 3380–3385 (2014).
223. Golebiewska, E.M., Poole, A.W.: Platelet secretion: From haemostasis to wound healing and beyond. *Blood Rev.* 29, 3, 153–162 (2015). <https://doi.org/10.1016/j.blre.2014.10.003>.
224. Gonsales, M.C. et al.: Multimodal Analysis of SCN1A Missense Variants Improves Interpretation of Clinically Relevant Variants in Dravet Syndrome. *Front. Neurol.* 10, 289 (2019). <https://doi.org/10.3389/fneur.2019.00289>.
225. Gontko-Romanowska, K. et al.: The assessment of laboratory parameters in children with fever and febrile seizures. *Brain Behav.* 7, 7, e00720 (2017). <https://doi.org/10.1002/brb3.720>.
226. Gordon, K.E. et al.: Is temperature regulation different in children susceptible to febrile seizures? *Can. J. Neurol. Sci. J. Can. Sci. Neurol.* 36, 2, 192–195 (2009).
227. Gorter, J.A. et al.: Status epilepticus, blood-brain barrier disruption, inflammation, and epileptogenesis. *Epilepsy Behav.* EB. 49, 13–16 (2015). <https://doi.org/10.1016/j.yebeh.2015.04.047>.

228. Gradišnik, P. et al.: Predictive value of paroxysmal EEG abnormalities for future epilepsy in focal febrile seizures. *Brain Dev.* 37, 9, 868–873 (2015). <https://doi.org/10.1016/j.braindev.2015.02.005>.
229. Graves, R.C. et al.: Febrile seizures: risks, evaluation, and prognosis. *Am. Fam. Physician.* 85, 2, 149–153 (2012).
230. Griffin, A.L. et al.: Zebrafish studies identify serotonin receptors mediating antiepileptic activity in Dravet syndrome. *Brain Commun.* 1, 1, fcz008 (2019). <https://doi.org/10.1093/braincomms/fcz008>.
231. Guerrini, R. et al.: Lamotrigine and seizure aggravation in severe myoclonic epilepsy. *Epilepsia.* 39, 5, 508–512 (1998). <https://doi.org/10.1111/j.1528-1157.1998.tb01413.x>.
232. Guerrini, R. et al.: Neuroimaging and neuropathology of Dravet syndrome. *Epilepsia.* 52 Suppl 2, 30–34 (2011). <https://doi.org/10.1111/j.1528-1167.2011.02998.x>.
233. Guerrini, R. et al.: Variable epilepsy phenotypes associated with a familial intragenic deletion of the SCN1A gene. *Epilepsia.* 51, 12, 2474–2477 (2010). <https://doi.org/10.1111/j.1528-1167.2010.02790.x>.
234. Güneş, A. et al.: Correlation of mean platelet volume, neutrophil-to-lymphocyte ratio, and disease activity in children with juvenile idiopathic arthritis. *Int. J. Clin. Exp. Med.* 8, 7, 11337–11341 (2015).
235. Güneş, S. et al.: Oxidant Status in Children After Febrile Seizures. *Pediatr. Neurol.* 40, 1, 47–49 (2009). <https://doi.org/10.1016/j.pediatrneurol.2008.09.006>.
236. Gunnerson, K.J., Harvey, C.E.: Lactic Acidosis. (2018).
237. Gupta, S. et al.: Iron Deficiency as a Risk Factor for Febrile Seizures – A, <https://www.semanticscholar.org/paper/Iron-Deficiency-as-a-Risk-Factor-for-Febrile-%E2%80%93-A-Gupta-Agarwal/b2ea30cfaf82847fd86a1a7fb7102a87d301e427>, last accessed 2021/07/23.
238. Gupta, S. et al.: Serum Interleukin-6 Levels in Children with Febrile Seizures. *Indian Pediatr.* 55, 5, 411–413 (2018).
239. Ha, J. et al.: Interleukin-4 and tumor necrosis factor-alpha levels in children with febrile seizures. *Seizure.* 58, 156–162 (2018). <https://doi.org/10.1016/j.seizure.2018.04.004>.
240. Habibian, N. et al.: Association between Iron Deficiency Anemia and Febrile Convulsion in 3- to 60-Month-Old Children: A Systematic Review and Meta-Analysis. *Iran. J. Med. Sci.* 39, 6, 496–505 (2014).
241. Haerian, B.S. et al.: Contribution of GABRG2 Polymorphisms to Risk of Epilepsy and Febrile Seizure: a Multicenter Cohort Study and Meta-analysis. *Mol. Neurobiol.* 53, 8, 5457–5467 (2016). <https://doi.org/10.1007/s12035-015-9457-y>.
242. Hall, C.B. et al.: Human herpesvirus-6 infection in children. A prospective study of complications and reactivation. *N. Engl. J. Med.* 331, 7, 432–438 (1994). <https://doi.org/10.1056/NEJM199408183310703>.
243. Hambidge, S.J. et al.: Timely versus delayed early childhood vaccination and seizures. *Pediatrics.* 133, 6, e1492-1499 (2014). <https://doi.org/10.1542/peds.2013-3429>.
244. Hampers, L.C., Spina, L.A.: Evaluation and management of pediatric febrile seizures in the emergency department. *Emerg. Med. Clin. North Am.* 29, 1, 83–93 (2011). <https://doi.org/10.1016/j.emc.2010.08.008>.
- 404
245. Han, S. et al.: NaV1.1 channels are critical for intercellular communication in the suprachiasmatic nucleus and for normal circadian rhythms. *Proc. Natl. Acad. Sci.* 109, E368–E377 (2012). <https://doi.org/10.1073/pnas.1115729109>.
246. Harkin, L.A. et al.: The spectrum of SCN1A-related infantile epileptic encephalopathies. *Brain J. Neurol.* 130, Pt 3, 843–852 (2007). <https://doi.org/10.1093/brain/awm002>.

247. Harkin, L.A. et al.: Truncation of the GABA(A)-receptor gamma2 subunit in a family with generalized epilepsy with febrile seizures plus. *Am. J. Hum. Genet.* 70, 2, 530–536 (2002). <https://doi.org/10.1086/338710>.
248. Hartfield, D.S. et al.: The association between iron deficiency and febrile seizures in childhood. *Clin. Pediatr. (Phila.)* 48, 4, 420–426 (2009). <https://doi.org/10.1177/0009922809331800>.
249. Hartwig, J., Italiano, J.: The birth of the platelet. *J. Thromb. Haemost.* 1, 7, 1580–1586 (2003). <https://doi.org/10.1046/j.1538-7836.2003.00331.x>.
250. Haspolat, S. et al.: Interleukin-1beta, tumor necrosis factor-alpha, and nitrite levels in febrile seizures. *J. Child Neurol.* 17, 10, 749–751 (2002). <https://doi.org/10.1177/08830738020170101501>.
251. Hassan, B.J., Campos, M.P.: Iron Deficiency Thrombocytopenia: A Case Report and Review of the Literature. *J. Clin. Case Rep.* 08, 03, (2018). <https://doi.org/10.4172/2165-7920.10001090>.
252. Hastie, T. et al.: The Elements of Statistical Learning: Data Mining, Inference, and Prediction, Second Edition. Springer-Verlag, New York (2009). <https://doi.org/10.1007/978-0-387-84858-7>.
253. Hauser, W.A.: The prevalence and incidence of convulsive disorders in children. *Epilepsia.* 35 Suppl 2, S1-6 (1994). <https://doi.org/10.1111/j.1528-1157.1994.tb05932.x>.
254. Hauser, W.A. et al.: The risk of seizure disorders among relatives of children with febrile convulsions. *Neurology.* 35, 9, 1268–1268 (1985). <https://doi.org/10.1212/WNL.35.9.1268>.
255. Hawas, A. et al.: The impact of electrolytes in pathogenesis of simple febrile convulsions. *Med. J. Babylon.* 15, 12 (2018). https://doi.org/10.4103/MJBL.MJBL_4_18.
256. Heida, J.G. et al.: Febrile convulsions induced by the combination of lipopolysaccharide and low-dose kainic acid enhance seizure susceptibility, not epileptogenesis, in rats. *Epilepsia.* 46, 12, 1898–1905 (2005). <https://doi.org/10.1111/j.1528-1167.2005.00286.x>.
257. Heida, J.G. et al.: Lipopolysaccharide-induced febrile convulsions in the rat: short-term sequelae. *Epilepsia.* 45, 11, 1317–1329 (2004). <https://doi.org/10.1111/j.0013-9580.2004.13704.x>.
258. Heida, J.G. et al.: The role of interleukin-1beta in febrile seizures. *Brain Dev.* 31, 5, 388–393 (2009). <https://doi.org/10.1016/j.braindev.2008.11.013>.
259. Helbig, I.: Genetic Causes of Generalized Epilepsies. *Semin. Neurol.* 35, 3, 288–292 (2015). <https://doi.org/10.1055/s-0035-1552922>.
260. Helbig, I., Tayoun, A.A.N.: Understanding Genotypes and Phenotypes in Epileptic Encephalopathies. *Mol. Syndromol.* 7, 4, 172–181 (2016). <https://doi.org/10.1159/000448530>.
261. Henriksen, M.W. et al.: De novo mutations in SCN1A are associated with classic Rett syndrome: a case report. *BMC Med. Genet.* 19, 1, 184 (2018). <https://doi.org/10.1186/s12881-018-0700-z>.
262. Hernberg, M. et al.: The prognostic role of blood lymphocyte subset distribution in patients with resected high-risk primary or regionally metastatic melanoma. *J. Immunother.* Hagerstown Md 1997. 30, 7, 773–779 (2007). <https://doi.org/10.1097/CJI.0b013e31814e0898>.
263. Heron, S.E. et al.: De novo SCN1A mutations in Dravet syndrome and related epileptic encephalopathies are largely of paternal origin. *J. Med. Genet.* 47, 2, 137–141 (2010). <https://doi.org/10.1136/jmg.2008.065912>.
264. Heydarian, F. et al.: The First Febrile Seizure: An Updated Study for Clinical Risk Factors. *Iran. J. Pediatr.* 6, (2018). <https://doi.org/10.5812/ijp.69761>.
265. Higgins, C.: Central venous blood gas analysis, <https://acute caretesting.org/en/articles/central-venous-blood-gas-analysis>, last accessed 2019/10/23.
266. Higgins, C.: Lactate and lactic acidosis, <https://acute caretesting.org/en/articles/lactate-and-lactic-acidosis/>, last accessed 2019/10/23.
267. Hilber, K. et al.: The selectivity filter of the voltage-gated sodium channel is involved in channel activation. *J. Biol. Chem.* 276, 30, 27831–27839 (2001). <https://doi.org/10.1074/jbc.M101933200>.
268. Hildebrand, M.S. et al.: Loss of synaptic Zn²⁺ transporter function increases risk of febrile seizures. *Sci. Rep.* 5, 17816 (2015). <https://doi.org/10.1038/srep17816>.

269. Hirose, S. et al.: SCN1A testing for epilepsy: application in clinical practice. *Epilepsia*. 54, 5, 946–952 (2013). <https://doi.org/10.1111/epi.12168>.
- 405
270. Hjortebjerg, D. et al.: Exposure to traffic noise and air pollution and risk for febrile seizure: a cohort study. *Scand. J. Work. Environ. Health*. 44, 5, 539–546 (2018). <https://doi.org/10.5271/sjweh.3724>.
271. Hong, H. et al.: Steady increment of immature platelet fraction is suppressed by irradiation in single-donor platelet components during storage. *PLoS One*. 9, 1, e85465 (2014). <https://doi.org/10.1371/journal.pone.0085465>.
272. Hsiao, J. et al.: Upregulation of Haploinsufficient Gene Expression in the Brain by Targeting a Long Non-coding RNA Improves Seizure Phenotype in a Model of Dravet Syndrome. *EBioMedicine*. 9, 257–277 (2016). <https://doi.org/10.1016/j.ebiom.2016.05.011>.
273. Huang, C. et al.: Apolipoprotein 4 may increase viral load and seizure frequency in mesial temporal lobe epilepsy patients with positive human herpes virus 6B. *Neurosci. Lett*. 593, 29–34 (2015). <https://doi.org/10.1016/j.neulet.2014.12.063>.
274. Huang, C.C. et al.: Risk factors for a first febrile convulsion in children: a population study in southern Taiwan. *Epilepsia*. 40, 6, 719–725 (1999). <https://doi.org/10.1111/j.1528-1157.1999.tb00769.x>.
275. Huang, J. et al.: Association of single nucleotide polymorphisms of SCN1A gene with therapeutic effect of carbamazepine among ethnic Zhuang Chinese patients with epilepsy]. *Zhonghua Yi Xue Zi Chuan Xue Za Zhi Zhonghua Yixue Yichuanxue Zazhi Chin. J. Med. Genet.* 36, 3, 271–274 (2019). <https://doi.org/10.3760/cma.j.issn.1003-9406.2019.03.020>.
276. Huang, W.-X. et al.: TRPV1 promotes repetitive febrile seizures by pro-inflammatory cytokines in immature brain. *Brain. Behav. Immun.* 48, 68–77 (2015). <https://doi.org/10.1016/j.bbi.2015.01.017>.
277. Huczek, Z. et al.: Mean platelet volume on admission predicts impaired reperfusion and long-term mortality in acute myocardial infarction treated with primary percutaneous coronary intervention. *J. Am. Coll. Cardiol.* 46, 2, 284–290 (2005). <https://doi.org/10.1016/j.jacc.2005.03.065>.
278. Hurst, D.L.: Epidemiology of Severe Myoclonic Epilepsy of Infancy. *Epilepsia*. 31, 4, 397–400 (1990). <https://doi.org/10.1111/j.1528-1157.1990.tb05494.x>.
279. Hwang, G. et al.: Predictors of unprovoked seizure after febrile seizure: short-term outcomes. *Brain Dev*. 37, 3, 315–321 (2015). <https://doi.org/10.1016/j.braindev.2014.06.003>.
280. Idro, R. et al.: Iron Deficiency and Acute Seizures: Results from Children Living in Rural Kenya and a Meta-Analysis. *PLOS ONE*. 5, 11, e14001 (2010). <https://doi.org/10.1371/journal.pone.0014001>.
281. Imuekemhe, S.O. et al.: Cerebrospinal fluid/serum lactic acid in febrile convulsions. *East Afr. Med. J.* 66, 9, 589–593 (1989).
282. Indriani, A. et al.: Five Years Study of Recurrent Febrile Seizure Risk Factors. *Althea Med. J.* 4, 2, 282–285 (2017). <https://doi.org/10.15850/amj.v4n2.1086>.
283. Inoue, S., Johnston, M.: Pediatric Thrombocytosis Clinical Presentation. (2020).
284. Ishii, A. et al.: SCN1A.NET, <https://www.scn1a.net/>, last accessed 2021/03/24.
285. Ishizaki, Y. et al.: Interleukin-10 is associated with resistance to febrile seizures: genetic association and experimental animal studies. *Epilepsia*. 50, 4, 761–767 (2009). <https://doi.org/10.1111/j.1528-1167.2008.01861.x>.
286. Ito, M. et al.: Autosomal dominant epilepsy with febrile seizures plus with missense mutations of the (Na⁺)-channel ??1 subunit gene, SCN1A. *Epilepsy Res.* 48, 15–23 (2002). [https://doi.org/10.1016/S0920-1211\(01\)00313-8](https://doi.org/10.1016/S0920-1211(01)00313-8).
287. Izumi, Y. et al.: Hypozincemia during fever may trigger febrile convulsion. *Med. Hypotheses*. 32, 1, 77–80 (1990). [https://doi.org/10.1016/0306-9877\(90\)90073-n](https://doi.org/10.1016/0306-9877(90)90073-n).

288. Jan, M., Aquino, M.: The use of chloral hydrate in pediatric electroencephalography. *Neurosciences*. 6, 99–102 (2001).
289. Jang, H.N. et al.: Prospective case control study of iron deficiency and the risk of febrile seizures in children in South Korea. *BMC Pediatr.* 19, 1, 309 (2019). <https://doi.org/10.1186/s12887-019-1675-4>.
290. Janusz Wendorff, Krzysztof Zeman: Immunology of febrile seizures. *Neurol. Dziecięca*. 20, 40, 41–46 (2011).
291. Jehangir A Bhat et al.: Association of iron deficiency anemia with simple febrile seizures: a hospital-based observational case-control study. *Menoufia Med. J.* 33, 3, 882 (2020). https://doi.org/10.4103/mmj.mmj_37_19.
- 406
292. Jeong, K.A. et al.: Early postictal electroencephalography and correlation with clinical findings in children with febrile seizures. *Korean J. Pediatr.* 56, 12, 534–539 (2013). <https://doi.org/10.3345/kjp.2013.56.12.534>.
293. Jiang, T. et al.: Clinical and molecular analysis of epilepsy-related genes in patients with Dravet syndrome. *Medicine (Baltimore)*. 97, 50, e13565 (2018). <https://doi.org/10.1097/MD.00000000000013565>.
294. Johnson, W.G. et al.: Pedigree analysis in families with febrile seizures. *Am. J. Med. Genet.* 61, 4, 345–352 (1996). [https://doi.org/10.1002/\(SICI\)1096-8628\(19960202\)61:4<345::AID-AJMG8>3.0.CO;2-T](https://doi.org/10.1002/(SICI)1096-8628(19960202)61:4<345::AID-AJMG8>3.0.CO;2-T).
295. Joshi, A.D. et al.: Sodium ion channel mutations in glioblastoma patients correlate with shorter survival. *Mol. Cancer*. 10, 17 (2011). <https://doi.org/10.1186/1476-4598-10-17>.
296. Joshi, C. et al.: Do clinical variables predict an abnormal EEG in patients with complex febrile seizures? *Seizure*. 14, 6, 429–434 (2005). <https://doi.org/10.1016/j.seizure.2005.07.006>.
297. Jun, Y.S. et al.: Relationship between iron deficiency anemia and febrile convulsion in infants. *Korean J. Pediatr.* 53, 3, 392–396 (2010).
298. Kahle, K.T. et al.: The KCC2 Cotransporter and Human Epilepsy: Getting Excited About Inhibition. *Neurosci. Rev. J. Bringing Neurobiol. Neurol. Psychiatry*. 22, 6, 555–562 (2016). <https://doi.org/10.1177/1073858416645087>.
299. Kahlig, K.M. et al.: Divergent sodium channel defects in familial hemiplegic migraine. *Proc. Natl. Acad. Sci. U. S. A.* 105, 28, 9799–9804 (2008). <https://doi.org/10.1073/pnas.0711717105>.
300. Kajbaf, F. et al.: Mechanisms underlying stress-induced hyperglycemia in critically ill patients. *Clin. Pract.* 4, 1, 97 (2007).
301. Kajitani, T. et al.: Febrile convulsions and rolandic discharges. *Brain Dev.* 3, 4, 351–359 (1981). [https://doi.org/10.1016/s0387-7604\(81\)80063-0](https://doi.org/10.1016/s0387-7604(81)80063-0).
302. Kalume, F. et al.: Reduced sodium current in Purkinje neurons from Nav1.1 mutant mice: implications for ataxia in severe myoclonic epilepsy in infancy. *J. Neurosci. Off. J. Soc. Neurosci.* 27, 41, 11065–11074 (2007). <https://doi.org/10.1523/JNEUROSCI.2162-07.2007>.
303. Kalume, F. et al.: Sleep impairment and reduced interneuron excitability in a mouse model of Dravet Syndrome. *Neurobiol. Dis.* 77, 141–154 (2015). <https://doi.org/10.1016/j.nbd.2015.02.016>.
304. Kamath, S. et al.: Platelet activation: assessment and quantification. *Eur. Heart J.* 22, 17, 1561–1571 (2001). <https://doi.org/10.1053/euhj.2000.2515>.
305. Kanai, K. et al.: Effect of localization of missense mutations in SCN1A on epilepsy phenotype severity. *Neurology*. 63, 2, 329–334 (2004). <https://doi.org/10.1212/01.WNL.0000129829.31179.5B>.
306. Kanemoto, K. et al.: Increased frequency of interleukin-1beta-511T allele in patients with temporal lobe epilepsy, hippocampal sclerosis, and prolonged febrile convulsion. *Epilepsia*. 44, 6, 796–799 (2003). <https://doi.org/10.1046/j.1528-1157.2003.43302.x>.

307. Kanemura, H. et al.: EEG characteristics predict subsequent epilepsy in children with febrile seizure. *Brain Dev.* 34, 4, 302–307 (2012). <https://doi.org/10.1016/j.braindev.2011.07.007>.
308. Kang, J.-Q. et al.: Why does fever trigger febrile seizures? GABAA receptor gamma2 subunit mutations associated with idiopathic generalized epilepsies have temperature-dependent trafficking deficiencies. *J. Neurosci. Off. J. Soc. Neurosci.* 26, 9, 2590–2597 (2006). <https://doi.org/10.1523/JNEUROSCI.4243-05.2006>.
309. Kang, J.-Q., Macdonald, R.L.: Molecular Pathogenic Basis for GABRG2 Mutations Associated With a Spectrum of Epilepsy Syndromes, From Generalized Absence Epilepsy to Dravet Syndrome. *JAMA Neurol.* 73, 8, 1009–1016 (2016). <https://doi.org/10.1001/jamaneurol.2016.0449>.
310. Kao, A., Rao, P.M.: Chapter 13 - Idiopathic generalized epilepsies. In: Stefan, H. and Theodore, W.H. (eds.) *Handbook of Clinical Neurology.* pp. 209–224 Elsevier (2012). <https://doi.org/10.1016/B978-0-444-52898-8.00013-6>.
311. Kaputu Kalala Malu, C. et al.: [Epidemiology and characteristics of febrile seizures in children]. *Rev. Med. Liege.* 68, 4, 180–185 (2013).
312. Karatas, H. et al.: Investigation of HSV-1, HSV-2, CMV, HHV-6 and HHV-8 DNA by real-time PCR in surgical resection materials of epilepsy patients with mesial temporal lobe sclerosis. *J. Neurol. Sci.* 264, 1–2, 151–156 (2008). <https://doi.org/10.1016/j.jns.2007.08.010>.
313. Karimi, A. et al.: Evaluation of Viral (HHV6, Adenovirus, HSV1, Enterovirus) and Bacterial Infection in Children with Febrile Convulsion by Serum PCR and Blood Culture Mofid 407 Children’s Hospital , 2016 - 2017. *Arch. Pediatr. Infect. Dis.* In Press, (2018). <https://doi.org/10.5812/pedinfect.63954>.
314. Karimi, P. et al.: Association of iron deficiency anemia and febrile seizure in Asia: A systematic review and meta-analysis. *Iran. J. Neonatol.* 9, 42–52 (2018). <https://doi.org/10.22038/ijn.2018.22323.1264>.
315. Karimi, P., Rashtchizadeh, N.: Oxidative Versus Thrombotic Stimulation of Platelets Differentially activates Signalling Pathways. *J. Cardiovasc. Thorac. Res.* 5, 2, 61–65 (2013). <https://doi.org/10.5681/jcvtr.2013.013>.
316. Karimzadeh, P. et al.: The Best Time for EEG Recording in Febrile Seizure. *Iran. J. Child Neurol.* 8, 1, 20–25 (2014).
317. Kasperaviciute, D. et al.: Epilepsy, hippocampal sclerosis and febrile seizures linked by common genetic variation around SCN1A. *Brain J. Neurol.* 136, Pt 10, 3140–3150 (2013). <https://doi.org/10.1093/brain/awt233>.
318. Katona, I.: Cannabis and Endocannabinoid Signaling in Epilepsy. *Handb. Exp. Pharmacol.* 231, 285–316 (2015). https://doi.org/10.1007/978-3-319-20825-1_10.
319. Kauffman, M.A. et al.: Association study between interleukin 1 beta gene and epileptic disorders: a HuGe review and meta-analysis. *Genet. Med. Off. J. Am. Coll. Med. Genet.* 10, 2, 83–88 (2008). <https://doi.org/10.1097/GIM.0b013e318161317c>.
320. Kaushansky, K.: The molecular mechanisms that control thrombopoiesis. *J. Clin. Invest.* 115, 12, 3339–3347 (2005). <https://doi.org/10.1172/JCI26674>.
321. Kavčič, A., Rener-Primec, Z.: Predictive Value of Epileptiform Discharges for Subsequent Epilepsy After Febrile Seizures. *J. Child Neurol.* 33, 12, 772–775 (2018). <https://doi.org/10.1177/0883073818787064>.
322. Kavcic, V. et al.: The relationship between baseline EEG spectra power and memory performance in older African Americans endorsing cognitive concerns in a community setting. *Int. J. Psychophysiol. Off. J. Int. Organ. Psychophysiol.* 109, 116–123 (2016). <https://doi.org/10.1016/j.ijpsycho.2016.09.001>.

323. Kawai, A.T. et al.: Febrile Seizures After 2010–2011 Trivalent Inactivated Influenza Vaccine. *Pediatrics*. 136, 4, e848–e855 (2015). <https://doi.org/10.1542/peds.2015-0635>.
324. Kawai, T., Akira, S.: The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors. *Nat. Immunol.* 11, 5, 373–384 (2010). <https://doi.org/10.1038/ni.1863>.
325. Kelly, G.: Body temperature variability (Part 1): a review of the history of body temperature and its variability due to site selection, biological rhythms, fitness, and aging. *Altern. Med. Rev. J. Clin. Ther.* 11, 4, 278–293 (2006).
326. Kent, P. et al.: A comparison of three clustering methods for finding subgroups in MRI, SMS or clinical data: SPSS TwoStep Cluster analysis, Latent Gold and SNOB. *BMC Med. Res. Methodol.* 14, 113 (2014). <https://doi.org/10.1186/1471-2288-14-113>.
327. Khadria, S.S. et al.: Association between serum selenium levels and febrile seizures in Indian children: a case control study. *Int. J. Contemp. Pediatr.* 4, 2, 553–556 (2017). <https://doi.org/10.18203/2349-3291.ijcp20170708>.
328. Khajeh, A. et al.: Serum Zinc Level in Children With Febrile Convulsion, <https://sites.kowsarpub.com/zjrms/articles/5881.html#abstract>, last accessed 2021/03/13.
329. Khan, W.A. et al.: Central Nervous System Manifestations of Childhood Shigellosis: Prevalence, Risk Factors, and Outcome. *Pediatrics*. 103, 2, e18–e18 (1999). <https://doi.org/10.1542/peds.103.2.e18>.
330. Khoshdel, A. et al.: Selenium and leptin levels in febrile seizure: a case-control study in children. *Korean J. Pediatr.* 56, 2, 80–85 (2013). <https://doi.org/10.3345/kjp.2013.56.2.80>.
331. Kilicaslan, B. et al.: Association between hypocapnia and febrile seizures. *J. Child Neurol.* 29, 5, 599–602 (2014). <https://doi.org/10.1177/0883073813513070>.
332. Killeen, P.R., Jacobs, K.W.: Coal Is Not Black, Snow Is Not White, Food Is Not a Reinforcer: The Roles of Affordances and Dispositions in the Analysis of Behavior. *Behav. Anal.* 40, 1, 17–38 (2017). <https://doi.org/10.1007/s40614-016-0080-7>.
333. Kim, C.H. et al.: An increase in mean platelet volume from baseline is associated with mortality in patients with severe sepsis or septic shock. *PLoS One*. 10, 3, e0119437 (2015). <https://doi.org/10.1371/journal.pone.0119437>.
334. Kim, D.-A., Kim, T.-Y.: Controversies over the interpretation of changes of mean platelet volume in rheumatoid arthritis. *Platelets*. 22, 1, 79–80 (2011). <https://doi.org/10.3109/09537101003663758>.
- 408
335. Kim, H. et al.: Clinical and EEG risk factors for subsequent epilepsy in patients with complex febrile seizures. *Epilepsy Res.* 105, 1–2, 158–163 (2013). <https://doi.org/10.1016/j.eplepsyres.2013.02.006>.
336. Kim, J.A., Connors, B.W.: High temperatures alter physiological properties of pyramidal cells and inhibitory interneurons in hippocampus. *Front. Cell. Neurosci.* 6, 27 (2012). <https://doi.org/10.3389/fncel.2012.00027>.
337. Kim, K. et al.: Analysis of plasma multiplex cytokines and increased level of IL-10 and IL-1Ra cytokines in febrile seizures. *J. Neuroinflammation*. 14, 1, 200 (2017). <https://doi.org/10.1186/s12974-017-0974-7>.
338. Kim, M.J. et al.: Comparison of platelet parameters in thrombocytopenic patients associated with acute myeloid leukemia and primary immune thrombocytopenia. *Blood Coagul. Fibrinolysis Int. J. Haemost. Thromb.* 25, 3, 221–225 (2014). <https://doi.org/10.1097/MBC.0000000000000027>.
339. Kira, R. et al.: Genetic susceptibility to febrile seizures: case-control association studies. *Brain Dev.* 32, 1, 57–63 (2010). <https://doi.org/10.1016/j.braindev.2009.09.018>.

340. Kisacik, B. et al.: Mean platelet volume (MPV) as an inflammatory marker in ankylosing spondylitis and rheumatoid arthritis. *Joint Bone Spine.* 75, 3, 291–294 (2008). <https://doi.org/10.1016/j.jbspin.2007.06.016>.
341. Kılıç, B.: Clinical Features and Evaluation in Terms of Prophylaxis of Patients With Febrile Seizures. *Sisli Etfal Hastan. Tip Bul.* 53, 3, 276–283 (2019). <https://doi.org/10.14744/SEMB.2019.30633>.
342. Klein, N.P. et al.: Measles-containing vaccines and febrile seizures in children age 4 to 6 years. *Pediatrics.* 129, 5, 809–814 (2012). <https://doi.org/10.1542/peds.2011-3198>.
343. Klein, N.P. et al.: Measles-Mumps-Rubella-Varicella Combination Vaccine and the Risk of Febrile Seizures. *Pediatrics.* (2010). <https://doi.org/10.1542/peds.2010-0665>.
344. Kloiber, O. et al.: Effect of hypoxia on bicuculline seizures of rat: NMR spectroscopy and bioluminescence imaging. *NMR Biomed.* 6, 5, 333–338 (1993). <https://doi.org/10.1002/nbm.1940060509>.
345. Knudsen, F.U.: Frequent febrile episodes and recurrent febrile convulsions. *Acta Neurol. Scand.* 78, 5, 414–417 (1988). <https://doi.org/10.1111/j.1600-0404.1988.tb03678.x>.
346. Knudsen, F.U.: Recurrence risk after first febrile seizure and effect of short term diazepam prophylaxis. *Arch. Dis. Child.* 60, 11, 1045–1049 (1985). <https://doi.org/10.1136/adc.60.11.1045>.
347. Kondo, K. et al.: Association of Human Herpesvirus 6 Infection of the Central Nervous System with Recurrence of Febrile Convulsions. *J. Infect. Dis.* 167, 5, 1197–1200 (1993). <https://doi.org/10.1093/infdis/167.5.1197>.
348. Korniluk, A. et al.: Mean Platelet Volume (MPV): New Perspectives for an Old Marker in the Course and Prognosis of Inflammatory Conditions, <https://www.hindawi.com/journals/mi/2019/9213074/>, last accessed 2021/03/13. <https://doi.org/10.1155/2019/9213074>.
349. Kovács, R. et al.: Bioenergetic Mechanisms of Seizure Control. *Front. Cell. Neurosci.* 12, 335 (2018). <https://doi.org/10.3389/fncel.2018.00335>.
350. Krishnan, R. et al.: Iron status among under-five children with first febrile convolution and subsequent febrile convolution. *Indian J. Child Health.* 5, 6, 397–401 (2018). <https://doi.org/10.32677/IJCH.2018.v05.i06.002>.
351. Kuang, Y.-Q. et al.: Epileptiform Discharges and Frontal Paroxysmal EEG Abnormality Act as Predictive Marker for Subsequent Epilepsy in Children With Complex Febrile Seizures. *Clin. EEG Neurosci.* 45, 4, 299–303 (2014). <https://doi.org/10.1177/1550059413507568>.
352. Kumar, E.D., Annamalai, T.: Correlation of iron deficiency anemia and events of febrile seizures among children aged 6 months to 5 years. Presented at the IAIM (2017).
353. Kumar, N. et al.: Risk Factors of Recurrence of Febrile Seizures in Children in a Tertiary Care Hospital in Kanpur: A One Year Follow Up Study. *Ann. Indian Acad. Neurol.* 22, 1, 31–36 (2019). https://doi.org/10.4103/aiian.AIAN_472_17.
354. Kurt, A.N.C. et al.: Dynamic thiol/disulphide homeostasis in children with febrile seizure. *Seizure.* 59, 34–37 (2018). <https://doi.org/10.1016/j.seizure.2018.01.012>.
355. Kuturec, M. et al.: Febrile seizures: is the EEG a useful predictor of recurrences? *Clin. Pediatr. (Phila.).* 36, 1, 31–36 (1997). <https://doi.org/10.1177/000992289703600105>.
- 409
356. Kwak, B.O. et al.: Relationship between iron deficiency anemia and febrile seizures in children: A systematic review and meta-analysis. *Seizure.* 52, 27–34 (2017). <https://doi.org/10.1016/j.seizure.2017.09.009>.
357. Kwon, A. et al.: Cytokine levels in febrile seizure patients: A systematic review and meta-analysis. *Seizure.* 59, 5–10 (2018). <https://doi.org/10.1016/j.seizure.2018.04.023>.

358. Lahat, E. et al.: Interleukin-1beta levels in serum and cerebrospinal fluid of children with febrile seizures. *Pediatr. Neurol.* 17, 1, 34–36 (1997). [https://doi.org/10.1016/s0887-8994\(97\)00034-9](https://doi.org/10.1016/s0887-8994(97)00034-9).
359. Laino, D. et al.: Management of Pediatric Febrile Seizures. *Int. J. Environ. Res. Public. Health.* 15, 10, E2232 (2018). <https://doi.org/10.3390/ijerph15102232>.
360. de Lanerolle, N.C. et al.: A retrospective analysis of hippocampal pathology in human temporal lobe epilepsy: evidence for distinctive patient subcategories. *Epilepsia.* 44, 5, 677–687 (2003). <https://doi.org/10.1046/j.1528-1157.2003.32701.x>.
361. Lang, C.H., Dobrescu, C.: Gram-Negative Infection Increases Noninsulin-Mediated Glucose Disposal. *Endocrinology.* 128, 2, 645–653 (1991). <https://doi.org/10.1210/endo-128-2-645>.
362. de Lange, I.M. et al.: Mosaicism of de novo pathogenic SCN1A variants in epilepsy is a frequent phenomenon that correlates with variable phenotypes. *Epilepsia.* 59, 3, 690–703 (2018). <https://doi.org/10.1111/epi.14021>.
363. de Lange, I.M. et al.: Outcomes and comorbidities of SCN1A-related seizure disorders. *Epilepsy Behav.* EB. 90, 252–259 (2019). <https://doi.org/10.1016/j.yebeh.2018.09.041>.
364. Larsen, S.B. et al.: Platelet turnover in stable coronary artery disease - influence of thrombopoietin and low-grade inflammation. *PLoS One.* 9, 1, e85566 (2014). <https://doi.org/10.1371/journal.pone.0085566>.
365. Le Van Quyen, M. et al.: The dark side of high-frequency oscillations in the developing brain. *Trends Neurosci.* 29, 7, 419–427 (2006). <https://doi.org/10.1016/j.tins.2006.06.001>.
366. Lee, C.Y. et al.: Iron Deficiency Anemia: The Possible Risk Factor of Complex Febrile Seizure and Recurrence of Febrile Seizure. *J. Korean Child Neurol. Soc.* 26, 4, 210–214 (2018). <https://doi.org/2018.26.4.210>.
367. Lee, E.H., Chung, S.: A comparative study of febrile and afebrile seizures associated with mild gastroenteritis. *Brain Dev.* 35, 7, 636–640 (2013). <https://doi.org/10.1016/j.braindev.2012.09.014>.
368. Lee, J.-Y. et al.: Children Experiencing First-Time or Prolonged Febrile Seizure Are Prone to Stress Hyperglycemia. *J. Child Neurol.* 31, 4, 439–443 (2016). <https://doi.org/10.1177/0883073815597757>.
369. Lee, S.-J. et al.: Impact of varying levels of hyperglycemia on clinicoradiographic outcomes after endovascular reperfusion treatment. *Sci. Rep.* 8, 1, 9832 (2018). <https://doi.org/10.1038/s41598-018-28175-6>.
370. Lee, V., Shaikh, F.: Inflammation: Cause or Consequence of Epilepsy? Presented at the March 4 (2019). <https://doi.org/10.5772/intechopen.83428>.
371. Lee, Y.-J. et al.: Large-scale structural alteration of brain in epileptic children with SCN1A mutation. *NeuroImage Clin.* 15, 594–600 (2017). <https://doi.org/10.1016/j.nicl.2017.06.002>.
372. Lefranc, F. et al.: Targeting the alpha 1 subunit of the sodium pump to combat glioblastoma cells. *Neurosurgery.* 62, 1, 211–221; discussion 221-222 (2008). <https://doi.org/10.1227/01.NEU.0000311080.43024.0E>.
373. Leung, A.K. et al.: Febrile seizures: an overview. *Drugs Context.* 7, 212536 (2018). <https://doi.org/10.7573/dic.212536>.
374. Leung, A.K., Robson, W.L.: Febrile convulsions. How dangerous are they? *Postgrad. Med.* 89, 5, 217–218, 221–222, 224 (1991). <https://doi.org/10.1080/00325481.1991.11700905>.
375. Leung, A.K.C., Robson, W.L.M.: Febrile seizures. *J. Pediatr. Health Care Off. Publ. Natl. Assoc. Pediatr. Nurse Assoc. Pract.* 21, 4, 250–255 (2007). <https://doi.org/10.1016/j.pedhc.2006.10.006>.
376. Li, B.-M. et al.: Autism in Dravet syndrome: Prevalence, features, and relationship to the clinical characteristics of epilepsy and mental retardation. *Epilepsy Behav.* 21, 3, 291–295 (2011). <https://doi.org/10.1016/j.yebeh.2011.04.060>.
377. Li, J. et al.: Combination of Mean Platelet Volume/Platelet Count Ratio and the APACHE II Score Better Predicts the Short-Term Outcome in Patients with Acute Kidney Injury Receiving

Continuous Renal Replacement Therapy. *Kidney Blood Press. Res.* 43, 2, 479–489 (2018). <https://doi.org/10.1159/000488694>.

410

378. Li, J., Shen, X.: Oxidative stress and adipokine levels were significantly correlated in diabetic patients with hyperglycemic crises. *Diabetol. Metab. Syndr.* 11, 13 (2019). <https://doi.org/10.1186/s13098-019-0410-5>.
379. Li, T.: Genetic Defects of Voltage-Gated Sodium Channel α Subunit 1 in Dravet Syndrome and the Patients' Response to Antiepileptic Drugs. *Ion Channels Health Sick.* (2018). <https://doi.org/10.5772/intechopen.76390>.
380. Li, X. et al.: The Influence of Vaccine on Febrile Seizure. *Curr. Neuropharmacol.* 16, 1, 59–65 (2018). <https://doi.org/10.2174/1570159X15666170726115639>.
381. Li, Y. et al.: Nomograms for predicting prognostic value of inflammatory biomarkers in colorectal cancer patients after radical resection. *Int. J. Cancer.* 139, 1, 220–231 (2016). <https://doi.org/10.1002/ijc.30071>.
382. Liao, W.-P. et al.: Partial epilepsy with antecedent febrile seizures and seizure aggravation by antiepileptic drugs: associated with loss of function of Na(v) 1.1. *Epilepsia.* 51, 9, 1669–1678 (2010). <https://doi.org/10.1111/j.1528-1167.2010.02645.x>.
383. Lin, G.-W. et al.: GAPDH-mediated posttranscriptional regulations of sodium channel Scn1a and Scn3a genes under seizure and ketogenic diet conditions. *Neuropharmacology.* 113, Pt A, 480–489 (2017). <https://doi.org/10.1016/j.neuropharm.2016.11.002>.
384. Lipka, K., Bülow, H.-H.: Lactic acidosis following convulsions. *Acta Anaesthesiol. Scand.* 47, 5, 616–618 (2003). <https://doi.org/10.1034/j.1399-6576.2003.00115.x>.
385. Lippi, G. et al.: Relation between red blood cell distribution width and inflammatory biomarkers in a large cohort of unselected outpatients. *Arch. Pathol. Lab. Med.* 133, 4, 628–632 (2009). <https://doi.org/10.1043/1543-2165-133.4.628>.
386. Liu, Z. et al.: The role of Mean Platelet Volume/platelet count Ratio and Neutrophil to Lymphocyte Ratio on the risk of Febrile Seizure. *Sci. Rep.* 8, 1, 15123 (2018). <https://doi.org/10.1038/s41598-018-33373-3>.
387. Livingston, J.H. et al.: Cerebrospinal fluid nucleotide metabolites following short febrile convulsions. *Dev. Med. Child Neurol.* 31, 2, 161–167 (1989). <https://doi.org/10.1111/j.1469-8749.1989.tb03974.x>.
388. Lopatina, T. et al.: Platelet-derived growth factor regulates the secretion of extracellular vesicles by adipose mesenchymal stem cells and enhances their angiogenic potential. *Cell Commun. Signal. CCS.* 12, 26 (2014). <https://doi.org/10.1186/1478-811X-12-26>.
389. Lopez, E. et al.: Relationship between calcium mobilization and platelet α - and δ -granule secretion. A role for TRPC6 in thrombin-evoked δ -granule exocytosis. *Arch. Biochem. Biophys.* 585, 75–81 (2015). <https://doi.org/10.1016/j.abb.2015.09.012>.
390. Losser, M.-R. et al.: Bench-to-bedside review: Glucose and stress conditions in the intensive care unit. *Crit. Care Lond. Engl.* 14, 4, 231 (2010). <https://doi.org/10.1186/cc9100>.
391. Lossin, C. et al.: Epilepsy-associated dysfunction in the voltage-gated neuronal sodium channel SCN1A. *J. Neurosci. Off. J. Soc. Neurosci.* 23, 36, 11289–11295 (2003).
392. MacDonald, S.E. et al.: Risk of febrile seizures after first dose of measles-mumps-rubella-varicella vaccine: a population-based cohort study. *CMAJ Can. Med. Assoc. J. J. Assoc. Medicale Can.* 186, 11, 824–829 (2014). <https://doi.org/10.1503/cmaj.140078>.
393. Machlus, K., Italiano, J.: The incredible journey: From megakaryocyte development to platelet formation. *J. Cell Biol.* 201, 785–96 (2013). <https://doi.org/10.1083/jcb.201304054>.

394. Mackowiak, P.A. et al.: A critical appraisal of 98.6 degrees F, the upper limit of the normal body temperature, and other legacies of Carl Reinhold August Wunderlich. *JAMA*. 268, 12, 1578–1580 (1992).
395. Mackowiak, P.A. et al.: Concepts of fever: recent advances and lingering dogma. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 25, 1, 119–138 (1997). <https://doi.org/10.1086/514520>.
396. Madia, F. et al.: Cryptic chromosome deletions involving SCN1A in severe myoclonic epilepsy of infancy. *Neurology.* 67, 7, 1230–1235 (2006). <https://doi.org/10.1212/01.wnl.0000238513.70878.54>.
397. Magwenzi, S. et al.: Oxidized LDL activates blood platelets through CD36/NOX2-mediated inhibition of the cGMP/protein kinase G signaling cascade. *Blood.* 125, 17, 2693–2703 (2015). <https://doi.org/10.1182/blood-2014-05-574491>.
398. Mahmoud: High-mobility group box 1 protein serum level in children with febrile seizures, <https://www.mmj.eg.net/article.asp?issn=1110-2098;year=2018;volume=31;issue=3;spage=1005;epage=1010;aulast=Mahmoud>, last accessed 2021/03/13.
- 411
399. Mahyar, A. et al.: A Case-Control Study of the Association Between Serum Copper Level and Febrile Seizures in Children. *Iran. J. Child Neurol.* 6, 1, 23–27 (2012). <https://doi.org/10.22037/ijcn.v6i1.2928>.
400. Malfitano, C. et al.: Hyperglycaemia protects the heart after myocardial infarction: aspects of programmed cell survival and cell death. *Eur. J. Heart Fail.* 12, 7, 659–667 (2010). <https://doi.org/10.1093/eurjhf/hfq053>.
401. Maluf, C.B. et al.: Standardization and reference intervals of platelet volume indices: Insight from the Brazilian longitudinal study of adult health (ELSA-BRASIL). *Platelets.* 26, 5, 413–420 (2015). <https://doi.org/10.3109/09537104.2014.942620>.
402. Management, S.C. on Q.I. and, Seizures, S. on F.: Febrile Seizures: Clinical Practice Guideline for the Long-term Management of the Child With Simple Febrile Seizures. *Pediatrics.* 121, 6, 1281–1286 (2008). <https://doi.org/10.1542/peds.2008-0939>.
403. Manfredini, R. et al.: Circadian and seasonal variation of first febrile seizures. *J. Pediatr.* 145, 6, 838–839 (2004). <https://doi.org/10.1016/j.jpeds.2004.06.079>.
404. Maniu, I.: Tehnici de analiză a datelor: statistica. Ed. Univ. Lucian Blaga Sibiu (2014).
405. Mantegazza, M. et al.: Identification of an Nav1.1 sodium channel (SCN1A) loss-of-function mutation associated with familial simple febrile seizures. *Proc. Natl. Acad. Sci.* 102, 50, 18177–18182 (2005). <https://doi.org/10.1073/pnas.0506818102>.
406. Mantovani, A. et al.: Neutrophils in the activation and regulation of innate and adaptive immunity. *Nat. Rev. Immunol.* 11, 8, 519–531 (2011). <https://doi.org/10.1038/nri3024>.
407. Marcy, S.M. et al.: Fever as an adverse event following immunization: case definition and guidelines of data collection, analysis, and presentation. *Vaccine.* 22, 5–6, 551–556 (2004). <https://doi.org/10.1016/j.vaccine.2003.09.007>.
408. Marfella, R. et al.: Acute hyperglycemia induces an oxidative stress in healthy subjects. *J. Clin. Invest.* 108, 4, 635–636 (2001). <https://doi.org/10.1172/JCI13727>.
409. Margari, L. et al.: Association between SCN1A gene polymorphisms and drug resistant epilepsy in pediatric patients. *Seizure.* 55, 30–35 (2018). <https://doi.org/10.1016/j.seizure.2018.01.002>.
410. Margetic, S.: Inflammation and haemostasis. *Biochem. Medica.* 22, 1, 49–62 (2012).
411. Mariani, E. et al.: Platelet-rich plasma affects bacterial growth in vitro. *Cytotherapy.* 16, 9, 1294–1304 (2014). <https://doi.org/10.1016/j.jcyt.2014.06.003>.
412. Marik, P.E.: Precision Glycemic Control in the ICU. *Crit. Care Med.* 44, 7, 1433–1434 (2016). <https://doi.org/10.1097/CCM.0000000000001683>.

413. Marik, P.E., Bellomo, R.: Stress hyperglycemia: an essential survival response! Crit. Care Lond. Engl. 17, 2, 305 (2013). <https://doi.org/10.1186/cc12514>.
414. Marin, M. et al.: Use of combination measles, mumps, rubella, and varicella vaccine: recommendations of the Advisory Committee on Immunization Practices (ACIP). MMWR Recomm. Rep. Morb. Mortal. Wkly. Rep. Recomm. Rep. 59, RR-3, 1–12 (2010).
415. Marini, C. et al.: Childhood absence epilepsy and febrile seizures: a family with a GABA(A) receptor mutation. Brain J. Neurol. 126, Pt 1, 230–240 (2003). <https://doi.org/10.1093/brain/awg018>.
416. Marini, C. et al.: Idiopathic epilepsies with seizures precipitated by fever and SCN1A abnormalities. Epilepsia. 48, 9, 1678–1685 (2007). <https://doi.org/10.1111/j.1528-1167.2007.01122.x>.
417. Marini, C. et al.: SCN1A duplications and deletions detected in Dravet syndrome: implications for molecular diagnosis. Epilepsia. 50, 7, 1670–1678 (2009). <https://doi.org/10.1111/j.1528-1167.2009.02013.x>.
418. Marini, C. et al.: The genetics of Dravet syndrome. Epilepsia. 52 Suppl 2, 24–29 (2011). <https://doi.org/10.1111/j.1528-1167.2011.02997.x>.
419. Maroso, M. et al.: Toll-like receptor 4 and high-mobility group box-1 are involved in ictogenesis and can be targeted to reduce seizures. Nat. Med. 16, 4, 413–419 (2010). <https://doi.org/10.1038/nm.2127>.
420. Martinez-Losa, M. et al.: Nav1.1-Overexpressing Interneuron Transplants Restore Brain Rhythms and Cognition in a Mouse Model of Alzheimer’s Disease. Neuron. 98, 1, 75-89.e5 (2018). <https://doi.org/10.1016/j.neuron.2018.02.029>.
421. Martin-Garcia, A.C. et al.: Platelet count and mean platelet volume predict outcome in adults with Eisenmenger syndrome. Heart. 104, 1, 45–50 (2018). <https://doi.org/10.1136/heartjnl-2016-311144>.
- 412
422. Martinos, M.M. et al.: Recognition memory is impaired in children after prolonged febrile seizures. Brain J. Neurol. 135, Pt 10, 3153–3164 (2012). <https://doi.org/10.1093/brain/aws213>.
423. Marzouk, H.: Relevance of hypcapnia to febrile seizures in children. Egypt. Pediatr. Assoc. Gaz. 63, 3, 98–102 (2015). <https://doi.org/10.1016/j.epag.2015.08.002>.
424. Mashimo, T. et al.: A missense mutation of the gene encoding voltage-dependent sodium channel (Nav1.1) confers susceptibility to febrile seizures in rats. J. Neurosci. Off. J. Soc. Neurosci. 30, 16, 5744–5753 (2010). <https://doi.org/10.1523/JNEUROSCI.3360-09.2010>.
425. Mathern, G.W. et al.: Influence of the type of initial precipitating injury and at what age it occurs on course and outcome in patients with temporal lobe seizures. J. Neurosurg. 82, 2, 220–227 (1995). <https://doi.org/10.3171/jns.1995.82.2.0220>.
426. Maugeri, N. et al.: Neutrophils phagocytose activated platelets in vivo: a phosphatidylserine, P-selectin, and β 2 integrin-dependent cell clearance program. Blood. 113, 21, 5254–5265 (2009). <https://doi.org/10.1182/blood-2008-09-180794>.
427. Maytal, J. et al.: The value of early postictal EEG in children with complex febrile seizures. Epilepsia. 41, 2, 219–221 (2000). <https://doi.org/10.1111/j.1528-1157.2000.tb00143.x>.
428. Mazarati, A.M.: Respiratory alkalosis: “basic” mechanism of febrile seizures? Epilepsy Curr. 7, 1, 25–27 (2007). <https://doi.org/10.1111/j.1535-7511.2007.00158.x>.
429. McClelland, S. et al.: Epileptogenesis after prolonged febrile seizures: mechanisms, biomarkers and therapeutic opportunities. Neurosci. Lett. 497, 3, 155–162 (2011). <https://doi.org/10.1016/j.neulet.2011.02.032>.
430. McDonald, T. et al.: Impairments in Oxidative Glucose Metabolism in Epilepsy and Metabolic Treatments Thereof. Front. Cell. Neurosci. 12, 274 (2018). <https://doi.org/10.3389/fncel.2018.00274>.

431. Meira e Cruz, M. et al.: Hypothalamic Control of Sleep-Wake Circadian Cycle. In: Gordeladze, J.O. and Baloyannis, S.J. (eds.) Hypothalamus in Health and Diseases. (2018). <https://doi.org/10.5772/intechopen.79899>.
432. Meisler, M.H. et al.: Sodium channel gene family: epilepsy mutations, gene interactions and modifier effects. *J. Physiol.* 588, Pt 11, 1841–1848 (2010). <https://doi.org/10.1113/jphysiol.2010.188482>.
433. Meisler, M.H., Kearney, J.A.: Sodium channel mutations in epilepsy and other neurological disorders. *J. Clin. Invest.* 115, 8, 2010–2017 (2005). <https://doi.org/10.1172/JCI25466>.
434. Meng, H. et al.: The SCN1A mutation database: updating information and analysis of the relationships among genotype, functional alteration, and phenotype. *Hum. Mutat.* 36, 6, 573–580 (2015). <https://doi.org/10.1002/humu.22782>.
435. Mészáros, K. et al.: In vivo glucose utilization by individual tissues during nonlethal hypermetabolic sepsis. *FASEB J. Off. Publ. Fed. Am. Soc. Exp. Biol.* 2, 15, 3083–3086 (1988). <https://doi.org/10.1096/fasebj.2.15.3056766>.
436. Mikkonen, K. et al.: Diurnal and seasonal occurrence of febrile seizures. *Pediatr. Neurol.* 52, 4, 424–427 (2015). <https://doi.org/10.1016/j.pediatrneurol.2015.01.001>.
437. Miller-Delaney, S.F.C. et al.: Differential DNA methylation profiles of coding and non-coding genes define hippocampal sclerosis in human temporal lobe epilepsy. *Brain J. Neurol.* 138, Pt 3, 616–631 (2015). <https://doi.org/10.1093/brain/awu373>.
438. Millichap, J.G.: Studies in febrile seizures. I. Height of body temperature as a measure of the febrile-seizure threshold. *Pediatrics.* 23, 1 Pt 1, 76–85 (1959).
439. Millichap, J.G., Millichap, J.J.: Role of viral infections in the etiology of febrile seizures. *Pediatr. Neurol.* 35, 3, 165–172 (2006). <https://doi.org/10.1016/j.pediatrneurol.2006.06.004>.
440. Millichap, J.J., Millichap, J.G.: Diurnal and Seasonal Occurrence of Febrile Seizures. *Pediatr. Neurol. Briefs.* 29, 4, 29 (2015). <https://doi.org/10.15844/pedneurbriefs-29-4-4>.
441. Mishra, O.P. et al.: Cerebrospinal fluid zinc, magnesium, copper and gamma-aminobutyric acid levels in febrile seizures. *J. Pediatr. Neurol.* 5, 1, 39–44 (2007).
442. Mkhize, N.V.P. et al.: The Effect of Quercetin on Pro- and Anti-Inflammatory Cytokines in a Prenatally Stressed Rat Model of Febrile Seizures. *J. Exp. Neurosci.* 11, 1179069517704668 (2017). <https://doi.org/10.1177/1179069517704668>.
443. Mocan, I.: SPSS: introducere în analiza datelor. Ed. Univ. Lucian Blaga, Sibiu (2005).
444. Moghissi, E.S. et al.: American Association of Clinical Endocrinologists and American Diabetes Association consensus statement on inpatient glycemic control. *Diabetes Care.* 32, 6, 1119–1131 (2009). <https://doi.org/10.2337/dc09-9029>.
- 413
445. Mohammadi, M.: Febrile seizures: four steps algorithmic clinical approach. *Iran. J. Pediatr.* 20, 1, 5–15 (2010).
446. Mohammadpour Touserkani, F. et al.: HHV-6 and seizure: A systematic review and meta-analysis. *J. Med. Virol.* 89, 1, 161–169 (2017). <https://doi.org/10.1002/jmv.24594>.
447. Mollah, M.A.H. et al.: Zinc concentration in serum and cerebrospinal fluid simultaneously decrease in children with febrile seizure: findings from a prospective study in Bangladesh. *Acta Paediatr. Oslo Nor.* 1992. 97, 12, 1707–1711 (2008). <https://doi.org/10.1111/j.1651-2227.2008.01001.x>.
448. Momen, A. et al.: Comparing prevalence and characteristic of anemia in children with simple versus complex febrile seizures. *Fam. Med. Prim. Care Rev.* 20, 25–28 (2018). <https://doi.org/10.5114/fmpcr.2018.73700>.

449. Monteiro, P.F. et al.: Platelet hyperaggregability in high-fat fed rats: a role for intraplatelet reactive-oxygen species production. *Cardiovasc. Diabetol.* 11, 5 (2012). <https://doi.org/10.1186/1475-2840-11-5>.
450. Moorthy, V. et al.: Measurement of malaria vaccine efficacy in phase III trials: report of a WHO consultation. *Vaccine*. 25, 28, 5115–5123 (2007). <https://doi.org/10.1016/j.vaccine.2007.01.085>.
451. Moreno-Cabrera, J.M. et al.: Evaluation of CNV detection tools for NGS panel data in genetic diagnostics. *Eur. J. Hum. Genet. EJHG*. 28, 12, 1645–1655 (2020). <https://doi.org/10.1038/s41431-020-0675-z>.
452. Moser, E. et al.: Association between brain temperature and dentate field potentials in exploring and swimming rats. *Science*. 259, 5099, 1324–1326 (1993). <https://doi.org/10.1126/science.8446900>.
453. Mosili, P. et al.: The Pathogenesis of Fever-Induced Febrile Seizures and Its Current State. *Neurosci. Insights*. 15, 2633105520956973 (2020). <https://doi.org/10.1177/2633105520956973>.
454. Mueller, A. et al.: Low long-term efficacy and tolerability of add-on rufinamide in patients with Dravet syndrome. *Epilepsy Behav. EB*. 21, 3, 282–284 (2011). <https://doi.org/10.1016/j.yebeh.2011.04.057>.
455. Mukherjee, A., Mukherjee, A.: Febrile convulsion--an overview. *J. Indian Med. Assoc.* 100, 5, 317–319, 326 (2002).
456. Mulley, J.C. et al.: SCN1A mutations and epilepsy. *Hum. Mutat.* 25, 6, 535–542 (2005). <https://doi.org/10.1002/humu.20178>.
457. Munteanu, C., Iliuta, A.: The role of sodium in the body. *Balneo Res. J.* 2, 70–74 (2011). <https://doi.org/10.12680/balneo.2011.1015>.
458. Nabbout, R. et al.: Encephalopathy in children with Dravet syndrome is not a pure consequence of epilepsy. *Orphanet J. Rare Dis.* 8, 1, 176 (2013). <https://doi.org/10.1186/1750-1172-8-176>.
459. Nabbout, R. et al.: Spectrum of SCN1A mutations in severe myoclonic epilepsy of infancy. *Neurology*. 60, 12, 1961–1967 (2003). <https://doi.org/10.1212/01.wnl.0000069463.41870.2f>.
460. Nagao, Y. et al.: A family of generalized epilepsy with febrile seizures plus type 2—a new missense mutation of SCN1A found in the pedigree of several patients with complex febrile seizures. *Epilepsy Res.* 63, 2, 151–156 (2005). <https://doi.org/10.1016/j.eplepsyres.2004.11.005>.
461. Nakayama, Y. et al.: Usefulness of the neutrophil/lymphocyte ratio measured preoperatively as a predictor of peritoneal metastasis in patients with advanced gastric cancer. *Surg. Today*. 44, 11, 2146–2152 (2014). <https://doi.org/10.1007/s00595-014-0917-1>.
462. Namakin, K. et al.: Serum Trace Elements in Febrile Seizure: A Case-Control Study. *Iran. J. Child Neurol.* 10, 3, 57–60 (2016).
463. Nanjundagowda, V.: Dravet syndrome presenting as west syndrome secondary to SCN1A mutation- a rare report of two cases. Presented at the July 1 (2016).
464. Nasehi, M.M. et al.: Association between Iron Deficiency Anemia and Febrile Seizure: a Systematic Review and Meta-Analysis. *J. Pediatr. Rev.* 1, 2, 13–18 (2013).
465. Nass, R.D. et al.: The role of postictal laboratory blood analyses in the diagnosis and prognosis of seizures. *Seizure*. 47, 51–65 (2017). <https://doi.org/10.1016/j.seizure.2017.02.013>.
466. Navaeifar, M.R. et al.: Relation between Febrile Seizure Recurrence and Hyponatremia in Children: A Single-center Trial. *J. Pediatr. Neurosci.* 15, 1, 5–8 (2020). https://doi.org/10.4103/JPN.JPN_4_19.
467. Nelson, K.B., Ellenberg, J.H.: Predictors of epilepsy in children who have experienced febrile seizures. *N. Engl. J. Med.* 295, 19, 1029–1033 (1976). <https://doi.org/10.1056/NEJM197611042951901>.
- 414
468. Nelson, K.B., Ellenberg, J.H.: Prognosis in children with febrile seizures. *Pediatrics*. 61, 5, 720–727 (1978).

469. Neville, B.G.R., Gindner, D.: Febrile seizures - semiology in humans and animal models: evidence of focality and heterogeneity. *Brain Dev.* 32, 1, 33–36 (2010). <https://doi.org/10.1016/j.braindev.2009.09.013>.
470. Nicks, B. et al.: Acute Lactic Acidosis. (2018).
471. Niehusmann, P. et al.: Presence of human herpes virus 6 DNA exclusively in temporal lobe epilepsy brain tissue of patients with history of encephalitis. *Epilepsia.* 51, 12, 2478–2483 (2010). <https://doi.org/10.1111/j.1528-1167.2010.02741.x>.
472. Nigade, R., Khambalkar, D.: Iron deficiency anaemia and its association with febrile seizures. *Int. J. Contemp. Pediatr.* 5, 1120 (2018). <https://doi.org/10.18203/2349-3291.ijcp20181554>.
473. Nikkhah, A. et al.: Differences in Mean Platelet Volume and Platelet Count between Children with Simple and Complex Febrile Seizures. *Iran. J. Child Neurol.* 11, 2, 44–47 (2017).
474. Nikolaus, S. et al.: Increased secretion of pro-inflammatory cytokines by circulating polymorphonuclear neutrophils and regulation by interleukin 10 during intestinal inflammation. *Gut.* 42, 4, 470–476 (1998). <https://doi.org/10.1136/gut.42.4.470>.
475. Nording, H.M. et al.: Platelets in Inflammation and Atherogenesis. *Front. Immunol.* 6, (2015). <https://doi.org/10.3389/fimmu.2015.00098>.
476. Nordli, D.R. et al.: Acute EEG findings in children with febrile status epilepticus: results of the FEBSTAT study. *Neurology.* 79, 22, 2180–2186 (2012). <https://doi.org/10.1212/WNL.0b013e3182759766>.
477. Nordli, D.R.: Generalized Epilepsy with Febrile Seizures Plus (GEFS+). In: Panayiotopoulos, C.P. (ed.) *Atlas of Epilepsies.* pp. 861–864 Springer London, London (2010). https://doi.org/10.1007/978-1-84882-128-6_123.
478. Nørgaard, M. et al.: Febrile seizures and cognitive function in young adult life: a prevalence study in Danish conscripts. *J. Pediatr.* 155, 3, 404–409 (2009). <https://doi.org/10.1016/j.jpeds.2009.04.003>.
479. Oakley, J.C. et al.: Insights into pathophysiology and therapy from a mouse model of Dravet syndrome. *Epilepsia.* 52 Suppl 2, 59–61 (2011). <https://doi.org/10.1111/j.1528-1167.2011.03004.x>.
480. O'Dempsey, T.J. et al.: The effect of temperature reduction on respiratory rate in febrile illnesses. *Arch. Dis. Child.* 68, 4, 492–495 (1993). <https://doi.org/10.1136/adc.68.4.492>.
481. Offringa, M. et al.: Risk factors for seizure recurrence in children with febrile seizures: A pooled analysis of individual patient data from five studies. *J. Pediatr.* 124, 4, 574–584 (1994). [https://doi.org/10.1016/S0022-3476\(05\)83136-1](https://doi.org/10.1016/S0022-3476(05)83136-1).
482. Offringa, M. et al.: Seizure recurrence after a first febrile seizure: a multivariate approach. *Dev. Med. Child Neurol.* 34, 1, 15–24 (1992). <https://doi.org/10.1111/j.1469-8749.1992.tb08559.x>.
483. Offringa, M., Moyer, V.A.: Evidence based paediatrics: Evidence based management of seizures associated with fever. *BMJ.* 323, 7321, 1111–1114 (2001). <https://doi.org/10.1136/bmj.323.7321.1111>.
484. Ogihara, M. et al.: Diurnal variation in febrile convulsions. *Pediatr. Neurol.* 42, 6, 409–412 (2010). <https://doi.org/10.1016/j.pediatrneurol.2010.02.011>.
485. Ohlraun, S. et al.: CARbon DIoxide for the treatment of Febrile seizures: rationale, feasibility, and design of the CARDIF-study. *J. Transl. Med.* 11, 157 (2013). <https://doi.org/10.1186/1479-5876-11-157>.
486. Ohno, Y. et al.: Scn1a missense mutation causes limbic hyperexcitability and vulnerability to experimental febrile seizures. *Neurobiol. Dis.* 41, 2, 261–269 (2011). <https://doi.org/10.1016/j.nbd.2010.09.013>.
487. Okumura, A. et al.: Acute encephalopathy in children with Dravet syndrome. *Epilepsia.* 53, 1, 79–86 (2012). <https://doi.org/10.1111/j.1528-1167.2011.03311.x>.

488. Örnek, Z. et al.: Comparison of Hemogram Parameters in Febrile Seizures Types. Düzce Tıp Fakültesi Derg. 22, (2020). <https://doi.org/10.18678/dtfd.628239>.
489. Orringer, C.E. et al.: Natural history of lactic acidosis after grand-mal seizures. A model for the study of an anion-gap acidosis not associated with hyperkalemia. N. Engl. J. Med. 297, 15, 796–799 (1977). <https://doi.org/10.1056/NEJM197710132971502>.
490. Osselaer, J.C. et al.: Platelet distribution width for differential diagnosis of thrombocytosis. Clin. Chem. 43, 6 Pt 1, 1072–1076 (1997).
- 415
491. Osteen, J.D. et al.: Selective spider toxins reveal a role for the Nav1.1 channel in mechanical pain. Nature. 534, 7608, 494–499 (2016). <https://doi.org/10.1038/nature17976>.
492. Østergaard, J.R.: Febrile seizures. Acta Paediatr. Oslo Nor. 1992. 98, 5, 771–773 (2009). <https://doi.org/10.1111/j.1651-2227.2009.01200.x>.
493. Otmakhova, N.A., Lisman, J.E.: Contribution of Ih and GABAB to synaptically induced afterhyperpolarizations in CA1: a brake on the NMDA response. J. Neurophysiol. 92, 4, 2027–2039 (2004). <https://doi.org/10.1152/jn.00427.2004>.
494. Ottman, R. et al.: Genetic testing in the epilepsies--report of the ILAE Genetics Commission. Epilepsia. 51, 4, 655–670 (2010). <https://doi.org/10.1111/j.1528-1167.2009.02429.x>.
495. Ouss, L. et al.: Autism spectrum disorder and cognitive profile in children with Dravet syndrome: Delineation of a specific phenotype. Epilepsia Open. 4, 1, 40–53 (2018). <https://doi.org/10.1002/epi4.12281>.
496. Oyakhrome, S. et al.: Assessment of fever in African children: implication for malaria trials. Am. J. Trop. Med. Hyg. 82, 2, 215–218 (2010). <https://doi.org/10.4269/ajtmh.2010.09-0419>.
497. Ozaydin, E. et al.: Differences in iron deficiency anemia and mean platelet volume between children with simple and complex febrile seizures. Seizure. 21, 3, 211–214 (2012). <https://doi.org/10.1016/j.seizure.2011.12.014>.
498. Özkal, M. et al.: Çocuklarda trombosit belirteçleri ve febril konvülzyon arasındaki ilişki. Cukurova Med. J. Çukurova Üniversitesi Tıp Fakültesi Derg. 41, 23861, 695–701 (2016). <https://doi.org/10.17826/cutf.254196>.
499. Öztürk, Z.A. et al.: Could platelet indices be new biomarkers for inflammatory bowel diseases? Eur. Rev. Med. Pharmacol. Sci. 17, 3, 334–341 (2013).
500. Pagani, A. et al.: Hepcidin and Anemia: A Tight Relationship. Front. Physiol. 10, 1294 (2019). <https://doi.org/10.3389/fphys.2019.01294>.
501. Panayiotopoulos, C.P.: Benign Childhood Focal Seizures and Related Epileptic Syndromes. Bladon Medical Publishing (2005).
502. Panayiotopoulos, C.P.: The Epilepsies: Seizures, Syndromes and Management. Bladon Medical Publishing, Oxfordshire (UK) (2005).
503. Papageorgiou, V. et al.: Association between iron deficiency and febrile seizures. Eur. J. Paediatr. Neurol. EJPN Off. J. Eur. Paediatr. Neurol. Soc. 19, 5, 591–596 (2015). <https://doi.org/10.1016/j.ejpn.2015.05.009>.
504. Papale, L.A. et al.: Altered sleep regulation in a mouse model of SCN1A-derived genetic epilepsy with febrile seizures plus (GEFS+). Epilepsia. 54, 4, 625–634 (2013). <https://doi.org/10.1111/epi.12060>.
505. Parihar, R., Ganesh, S.: The SCN1A gene variants and epileptic encephalopathies. J. Hum. Genet. 58, 9, 573–580 (2013). <https://doi.org/10.1038/jhg.2013.77>.
506. Patel, A.D., Vidaurre, J.: Complex febrile seizures: a practical guide to evaluation and treatment. J. Child Neurol. 28, 6, 762–767 (2013). <https://doi.org/10.1177/0883073813483569>.

507. Patel, H.C. et al.: Neurodegenerative actions of interleukin-1 in the rat brain are mediated through increases in seizure activity. *J. Neurosci. Res.* 83, 3, 385–391 (2006). <https://doi.org/10.1002/jnr.20735>.
508. Patel, N. et al.: Febrile seizures. *BMJ*. 351, h4240 (2015). <https://doi.org/10.1136/bmj.h4240>.
509. Patterson, J.L. et al.: Febrile seizures. *Pediatr. Ann.* 42, 12, 249–254 (2013). <https://doi.org/10.3928/00904481-20131122-09>.
510. Pavlidou, E., Panteliadis, C.: Prognostic factors for subsequent epilepsy in children with febrile seizures. *Epilepsia*. 54, 12, 2101–2107 (2013). <https://doi.org/10.1111/epi.12429>.
511. Pedersen, M. et al.: Human GABRG2 generalized epilepsy: Increased somatosensory and striatothalamic connectivity. *Neurol. Genet.* 5, 4, e340 (2019). <https://doi.org/10.1212/NXG.0000000000000340>.
512. Pedespan, L.: [Febrile seizures]. *Arch. Pediatr. Organe Off. Soc. Francaise Pediatr.* 14, 4, 394–398 (2007). <https://doi.org/10.1016/j.arcped.2007.02.005>.
513. Pereira, M. et al.: Acute Iron Deprivation Reprograms Human Macrophage Metabolism and Reduces Inflammation In Vivo. *Cell Rep.* 28, 2, 498–511.e5 (2019). <https://doi.org/10.1016/j.celrep.2019.06.039>.
514. Pernot, F. et al.: Inflammatory changes during epileptogenesis and spontaneous seizures in a mouse model of mesiotemporal lobe epilepsy. *Epilepsia*. 52, 12, 2315–2325 (2011). <https://doi.org/10.1111/j.1528-1167.2011.03273.x>.
- 416
515. Pino, J.M.V. et al.: Iron-Restricted Diet Affects Brain Ferritin Levels, Dopamine Metabolism and Cellular Prion Protein in a Region-Specific Manner. *Front. Mol. Neurosci.* 10, 145 (2017). <https://doi.org/10.3389/fnmol.2017.00145>.
516. Pisacane, A. et al.: Iron deficiency anaemia and febrile convulsions: case-control study in children under 2 years. *BMJ*. 313, 7053, 343 (1996). <https://doi.org/10.1136/bmj.313.7053.343>.
517. Pitkänen, A., Sutula, T.P.: Is epilepsy a progressive disorder? Prospects for new therapeutic approaches in temporal-lobe epilepsy. *Lancet Neurol.* 1, 3, 173–181 (2002). [https://doi.org/10.1016/s1474-4422\(02\)00073-x](https://doi.org/10.1016/s1474-4422(02)00073-x).
518. Pittau, F. et al.: Prognostic factors in patients with mesial temporal lobe epilepsy. *Epilepsia*. 50 Suppl 1, 41–44 (2009). <https://doi.org/10.1111/j.1528-1167.2008.01969.x>.
519. Poduri, A.: HCN1 Gain-Of-Function Mutations – A New Cause of Epileptic Encephalopathy. *Epilepsy Curr.* 14, 6, 348–349 (2014). <https://doi.org/10.5698/1535-7597-14.6.348>.
520. Poffers, M. et al.: Sodium Channel Nav1.3 Is Expressed by Polymorphonuclear Neutrophils during Mouse Heart and Kidney Ischemia In Vivo and Regulates Adhesion, Transmigration, and Chemotaxis of Human and Mouse Neutrophils In Vitro. *Anesthesiology*. 128, 6, 1151–1166 (2018). <https://doi.org/10.1097/ALN.0000000000002135>.
521. Ponomarev, E.D.: Fresh Evidence for Platelets as Neuronal and Innate Immune Cells: Their Role in the Activation, Differentiation, and Deactivation of Th1, Th17, and Tregs during Tissue Inflammation. *Front. Immunol.* 9, 406 (2018). <https://doi.org/10.3389/fimmu.2018.00406>.
522. Portales-Cervantes, L. et al.: The His155Tyr (489C>T) single nucleotide polymorphism of P2RX7 gene confers an enhanced function of P2X7 receptor in immune cells from patients with rheumatoid arthritis. *Cell. Immunol.* 276, 1–2, 168–175 (2012). <https://doi.org/10.1016/j.cellimm.2012.05.005>.
523. Poryo, M. et al.: Dravet syndrome: a new causative SCN1A mutation? *Clin. Case Rep.* 5, 5, 613–615 (2017). <https://doi.org/10.1002/ccr3.787>.
524. Prasad, A.N., Seshia, S.S.: Susceptibility to febrile seizures: more than just a faulty thermostat! *Can. J. Neurol. Sci. J. Can. Sci. Neurol.* 36, 3, 277–279 (2009). <https://doi.org/10.1017/s031716710000696x>.

525. Principi, N., Esposito, S.: Vaccines and febrile seizures. *Expert Rev. Vaccines.* 12, 8, 885–892 (2013). <https://doi.org/10.1586/14760584.2013.814781>.
526. Purnak, T. et al.: Mean platelet volume could be a promising biomarker to monitor dietary compliance in celiac disease. *Ups. J. Med. Sci.* 116, 3, 208–211 (2011). <https://doi.org/10.3109/03009734.2011.581399>.
527. Puskarjov, M. et al.: A variant of KCC2 from patients with febrile seizures impairs neuronal Cl⁻ extrusion and dendritic spine formation. *EMBO Rep.* 15, 6, 723–729 (2014). <https://doi.org/10.1002/embr.201438749>.
528. Qu, L. et al.: Hyperthermia decreases GABAergic synaptic transmission in hippocampal neurons of immature rats. *Neurobiol. Dis.* 27, 3, 320–327 (2007). <https://doi.org/10.1016/j.nbd.2007.06.003>.
529. Qu, Y. et al.: Nonclassical IL-1 beta secretion stimulated by P2X7 receptors is dependent on inflammasome activation and correlated with exosome release in murine macrophages. *J. Immunol. Baltim. Md 1950.* 179, 3, 1913–1925 (2007). <https://doi.org/10.4049/jimmunol.179.3.1913>.
530. Radzicki, D. et al.: Temperature-Sensitive Cav1.2 Calcium Channels Support Intrinsic Firing of Pyramidal Neurons and Provide a Target for the Treatment of Febrile Seizures. *J. Neurosci.* 33, 24, 9920–9931 (2013). <https://doi.org/10.1523/JNEUROSCI.5482-12.2013>.
531. Rantala, H. et al.: Factors triggering the first febrile seizure. *Acta Paediatr. Oslo Nor.* 1992. 84, 4, 407–410 (1995). <https://doi.org/10.1111/j.1651-2227.1995.tb13660.x>.
532. Rantala, H., Uhari, M.: Risk factors for recurrences of febrile convulsions. *Acta Neurol. Scand.* 90, 3, 207–210 (1994). <https://doi.org/10.1111/j.1600-0404.1994.tb02707.x>.
533. Raparelli, V. et al.: Low-grade endotoxemia and platelet activation in cirrhosis. *Hepatology.* 65, 2, 571–581 (2017). <https://doi.org/10.1002/hep.28853>.
534. Rathinam, V.A.K. et al.: Regulation of inflammasome signaling. *Nat. Immunol.* 13, 4, 333–342 (2012). <https://doi.org/10.1038/ni.2237>.
535. Ravizza, T., Vezzani, A.: Status epilepticus induces time-dependent neuronal and astrocytic expression of interleukin-1 receptor type I in the rat limbic system. *Neuroscience.* 137, 1, 301–308 (2006). <https://doi.org/10.1016/j.neuroscience.2005.07.063>.
- 417
536. Reid, C.A. et al.: Synaptic Zn²⁺ and febrile seizure susceptibility. *Br. J. Pharmacol.* 174, 2, 119–125 (2017). <https://doi.org/10.1111/bph.13658>.
537. Rich, S.S. et al.: Complex segregation analysis of febrile convulsions. *Am. J. Hum. Genet.* 41, 2, 249–257 (1987).
538. Richards, S. et al.: Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet. Med.* 17, 5, 405–423 (2015). <https://doi.org/10.1038/gim.2015.30>.
539. Ricobaraza, A. et al.: Epilepsy and neuropsychiatric comorbidities in mice carrying a recurrent Dravet syndrome SCN1A missense mutation. *Sci. Rep.* 9, 1, 14172 (2019). <https://doi.org/10.1038/s41598-019-50627-w>.
540. Rizzi, M. et al.: Glia activation and cytokine increase in rat hippocampus by kainic acid-induced status epilepticus during postnatal development. *Neurobiol. Dis.* 14, 3, 494–503 (2003). <https://doi.org/10.1016/j.nbd.2003.08.001>.
541. Rodríguez-Muñoz, M. et al.: Fenfluramine diminishes NMDA receptor-mediated seizures via its mixed activity at serotonin 5HT2A and type 1 sigma receptors. *Oncotarget.* 9, 34, 23373–23389 (2018). <https://doi.org/10.18632/oncotarget.25169>.
542. Rodríguez-Núñez, A. et al.: Purine metabolites and pyrimidine bases in cerebrospinal fluid of children with simple febrile seizures. *Dev. Med. Child Neurol.* 33, 10, 908–911 (1991). <https://doi.org/10.1111/j.1469-8749.1991.tb14801.x>.

543. Ronco, C. et al. eds: Critical care nephrology. Elsevier Inc, Philadelphia, PA (2018).
544. Rose, A.B.: Introns as Gene Regulators: A Brick on the Accelerator. *Front. Genet.* 9, 672 (2018). <https://doi.org/10.3389/fgene.2018.00672>.
545. Rosman, N.P. et al.: A controlled trial of diazepam administered during febrile illnesses to prevent recurrence of febrile seizures. *N. Engl. J. Med.* 329, 2, 79–84 (1993). <https://doi.org/10.1056/NEJM199307083290202>.
546. Rossi, M.A.: SCN1A and Febrile Seizures in Mesial Temporal Epilepsy: An Early Signal to Guide Prognosis and Treatment? *Epilepsy Curr.* 14, 4, 189–190 (2014). <https://doi.org/10.5698/1535-7597-14.4.189>.
547. Rouault, T.A., Cooperman, S.: Brain iron metabolism. *Semin. Pediatr. Neurol.* 13, 3, 142–148 (2006). <https://doi.org/10.1016/j.spen.2006.08.002>.
548. Rubinstein, M. et al.: Dissecting the phenotypes of Dravet syndrome by gene deletion. *Brain J. Neurol.* 138, Pt 8, 2219–2233 (2015). <https://doi.org/10.1093/brain/awv142>.
549. Sachan, D., Goyal, S.: Association of Hypocapnia in Children with Febrile Seizures. *J. Pediatr. Neurosci.* 13, 4, 388–391 (2018). https://doi.org/10.4103/JPN.JPN_73_18.
550. Sachdev, R. et al.: Establishing biological reference intervals for novel platelet parameters (immature platelet fraction, high immature platelet fraction, platelet distribution width, platelet large cell ratio, platelet-X, plateletrit, and platelet distribution width) and their correlations among each other. *Indian J. Pathol. Microbiol.* 57, 2, 231–235 (2014). <https://doi.org/10.4103/0377-4929.134676>.
551. Sadleir, L.G. et al.: Not all SCN1A epileptic encephalopathies are Dravet syndrome: Early profound Thr226Met phenotype. *Neurology.* 89, 10, 1035–1042 (2017). <https://doi.org/10.1212/WNL.0000000000004331>.
552. Sadleir, L.G., Scheffer, I.E.: Febrile seizures. *BMJ.* 334, 7588, 307–311 (2007). <https://doi.org/10.1136/bmj.39087.691817.AE>.
553. Safak, S. et al.: Association between mean platelet volume levels and inflammation in SLE patients presented with arthritis. *Afr. Health Sci.* 14, 4, 919–924 (2014). <https://doi.org/10.4314/ahs.v14i4.21>.
554. Saghazadeh, A. et al.: Genetic background of febrile seizures. *Rev. Neurosci.* 25, 1, 129–161 (2014). <https://doi.org/10.1515/revneuro-2013-0053>.
555. Saghazadeh, A. et al.: Possible role of trace elements in epilepsy and febrile seizures: a meta-analysis. *Nutr. Rev.* 73, 11, 760–779 (2015). <https://doi.org/10.1093/nutrit/nuv026>.
556. Sahib, A.M. et al.: The Risk of Iron Deficiency Anemia Contribution To Febrile Seizures Among Children (6-60) Months. *Int. J. Sci. Res. Publ. IJSRP.* 10, 9, 8–11 (2020). <https://doi.org/10.29322/IJSRP.10.09.2020.p10503>.
557. Sahli, M. et al.: Clinical exome sequencing identifies two novel mutations of the SCN1A and SCN2A genes in Moroccan patients with epilepsy: a case series. *J. Med. Case Reports.* 13, 1, 266 (2019). <https://doi.org/10.1186/s13256-019-2203-8>.
- 418
558. Sakauchi, M. et al.: Mortality in Dravet syndrome: search for risk factors in Japanese patients. *Epilepsia.* 52 Suppl 2, 50–54 (2011). <https://doi.org/10.1111/j.1528-1167.2011.03002.x>.
559. Sakr: Serum zinc status in febrile seizures, <https://www.azmj.eg.net/article.asp?issn=1687-1693;year=2018;volume=16;issue=2;spage=156;epage=159;aulast=Sakr>, last accessed 2021/03/13.
560. Salehi, B. et al.: Comparison of Relation between Attention Deficit Hyperactivity Disorder in Children with and without Simple Febrile Seizure Admitted in Arak Central Iran. *Iran. J. Child Neurol.* 10, 4, 56–61 (2016).
561. Salehiomran, M.R., Mahzari, M.: Zinc status in febrile seizure: a case-control study. *Iran. J. Child Neurol.* 7, 4, 20–23 (2013).

562. Salzmann, A. et al.: Carboxypeptidase A6 gene (CPA6) mutations in a recessive familial form of febrile seizures and temporal lobe epilepsy and in sporadic temporal lobe epilepsy. *Hum. Mutat.* 33, 1, 124–135 (2012). <https://doi.org/10.1002/humu.21613>.
563. Sanders, S.J. et al.: De novo mutations revealed by whole-exome sequencing are strongly associated with autism. *Nature*. 485, 7397, 237–241 (2012). <https://doi.org/10.1038/nature10945>.
564. San-Millán, I., Brooks, G.A.: Reexamining cancer metabolism: lactate production for carcinogenesis could be the purpose and explanation of the Warburg Effect. *Carcinogenesis*. 38, 2, 119–133 (2017). <https://doi.org/10.1093/carcin/bgw127>.
565. Sawyer, M. et al.: Vaccines and Febrile Seizures: Quantifying the Risk. *PEDIATRICS*. 138, (2016). <https://doi.org/10.1542/peds.2016-0976>.
566. Scheffer, I.E. et al.: Locus for febrile seizures. *Ann. Neurol.* 47, 6, 840–841 (2000). [https://doi.org/10.1002/1531-8249\(200006\)47:6<840::aid-ana28>3.0.co;2-v](https://doi.org/10.1002/1531-8249(200006)47:6<840::aid-ana28>3.0.co;2-v).
567. Scheffer, I.E. et al.: Temporal lobe epilepsy and GEFS+ phenotypes associated with SCN1B mutations. *Brain J. Neurol.* 130, Pt 1, 100–109 (2007). <https://doi.org/10.1093/brain/awl272>.
568. Scheffer, I.E., Berkovic, S.F.: Generalized epilepsy with febrile seizures plus. A genetic disorder with heterogeneous clinical phenotypes. *Brain J. Neurol.* 120 (Pt 3), 479–490 (1997). <https://doi.org/10.1093/brain/120.3.479>.
569. Schoonjans, A.-S. et al.: More daytime sleepiness and worse quality of sleep in patients with Dravet Syndrome compared to other epilepsy patients. *Eur. J. Paediatr. Neurol. EJPN Off. J. Eur. Paediatr. Neurol. Soc.* 23, 1, 61–69 (2019). <https://doi.org/10.1016/j.ejpn.2018.09.012>.
570. Schubert, J. et al.: Mutations in STX1B, encoding a presynaptic protein, cause fever-associated epilepsy syndromes. *Nat. Genet.* 46, 12, 1327–1332 (2014). <https://doi.org/10.1038/ng.3130>.
571. Schuchmann, S. et al.: Experimental febrile seizures are precipitated by a hyperthermia-induced respiratory alkalosis. *Nat. Med.* 12, 7, 817–823 (2006). <https://doi.org/10.1038/nm1422>.
572. Schuchmann, S. et al.: Respiratory alkalosis in children with febrile seizures. *Epilepsia*. 52, 11, 1949–1955 (2011). <https://doi.org/10.1111/j.1528-1167.2011.03259.x>.
573. Schutte, S.S. et al.: Model systems for studying cellular mechanisms of SCN1A-related epilepsy. *J. Neurophysiol.* 115, 4, 1755–1766 (2016). <https://doi.org/10.1152/jn.00824.2015>.
574. Seizures, S. on F.: Febrile Seizures: Guideline for the Neurodiagnostic Evaluation of the Child With a Simple Febrile Seizure. *Pediatrics*. 127, 2, 389–394 (2011). <https://doi.org/10.1542/peds.2010-3318>.
575. Selmer, K.K. et al.: SCN1A mutation screening in adult patients with Lennox-Gastaut syndrome features. *Epilepsy Behav. EB*. 16, 3, 555–557 (2009). <https://doi.org/10.1016/j.yebeh.2009.08.021>.
576. Sfaihi, L. et al.: Febrile seizures: an epidemiological and outcome study of 482 cases. *Childs Nerv. Syst. ChNS Off. J. Int. Soc. Pediatr. Neurosurg.* 28, 10, 1779–1784 (2012). <https://doi.org/10.1007/s00381-012-1789-6>.
577. Shah, M.M.: Cortical HCN channels: function, trafficking and plasticity. *J. Physiol.* 592, 13, 2711–2719 (2014). <https://doi.org/10.1113/jphysiol.2013.270058>.
578. Shah, P.B. et al.: EEG for children with complex febrile seizures. *Cochrane Database Syst. Rev.* 12, CD009196 (2015). <https://doi.org/10.1002/14651858.CD009196.pub3>.
579. Shaikh, A.M. et al.: Association of iron deficiency states and febrile seizures in children-a case control study. *Int. J. Res. Med. Sci.* 6, 3, 869–877 (2018). <https://doi.org/10.18203/2320-6012.ijrms20180606>.
580. Sharafi, R. et al.: Circadian Rhythm and the Seasonal Variation in Childhood Febrile Seizure. *Iran. J. Child Neurol.* 11, 3, 27–30 (2017).
581. Sharif, M.R. et al.: The Relationship Between Iron Deficiency and Febrile Convulsion: A Case-Control Study. *Glob. J. Health Sci.* 8, 2, 185–189 (2015). <https://doi.org/10.5539/gjhs.v8n2p185>.

582. Sherjil, A. et al.: Iron deficiency anaemia--a risk factor for febrile seizures in children. *J. Ayub Med. Coll. Abbottabad JAMC.* 22, 3, 71–73 (2010).
583. Shi, L. et al.: SCN1A and SCN2A polymorphisms are associated with response to valproic acid in Chinese epilepsy patients. *Eur. J. Clin. Pharmacol.* 75, 5, 655–663 (2019). <https://doi.org/10.1007/s00228-019-02633-0>.
584. Shibasaki, K. et al.: Effects of Body Temperature on Neural Activity in the Hippocampus: Regulation of Resting Membrane Potentials by Transient Receptor Potential Vanilloid 4. *J. Neurosci.* 27, 7, 1566–1575 (2007). <https://doi.org/10.1523/JNEUROSCI.4284-06.2007>.
585. Shin, D.H. et al.: An Increase in Mean Platelet Volume/Platelet Count Ratio Is Associated with Vascular Access Failure in Hemodialysis Patients. *PLOS ONE.* 12, 1, e0170357 (2017). <https://doi.org/10.1371/journal.pone.0170357>.
586. Shinnar, S. et al.: MRI abnormalities following febrile status epilepticus in children: the FEBSTAT study. *Neurology.* 79, 9, 871–877 (2012). <https://doi.org/10.1212/WNL.0b013e318266fcc5>.
587. Shinnar, S., Glauser, T.A.: Febrile seizures. *J. Child Neurol.* 17 Suppl 1, S44–52 (2002). <https://doi.org/10.1177/08830738020170010601>.
588. Shokrzade, M. et al.: Serum zinc and copper levels in children with febrile convulsion. *Pharm. Biomed. Res.* 2, 19–24 (2016). <https://doi.org/10.18869/acadpub.pbr.2.3.19>.
589. Simpson, H. et al.: Cerebrospinal fluid acid-base status and lactate and pyruvate concentrations after short (less than 30 minutes) first febrile convulsions in children. *Arch. Dis. Child.* 52, 11, 836–843 (1977). <https://doi.org/10.1136/adc.52.11.836>.
590. Singh, N.A. et al.: A role of SCN9A in human epilepsies, as a cause of febrile seizures and as a potential modifier of Dravet syndrome. *PLoS Genet.* 5, 9, e1000649 (2009). <https://doi.org/10.1371/journal.pgen.1000649>.
591. Singh, R. et al.: Generalized epilepsy with febrile seizures plus: a common childhood-onset genetic epilepsy syndrome. *Ann. Neurol.* 45, 1, 75–81 (1999). [https://doi.org/10.1002/1531-8249\(199901\)45:1<75::aid-art13>3.0.co;2-w](https://doi.org/10.1002/1531-8249(199901)45:1<75::aid-art13>3.0.co;2-w).
592. Singh, R. et al.: Severe myoclonic epilepsy of infancy: extended spectrum of GEFS+? *Epilepsia.* 42, 7, 837–844 (2001). <https://doi.org/10.1046/j.1528-1157.2001.042007837.x>.
593. Sisodiya, S.: Feverish prospects for seizure genetics. *Nat. Genet.* 46, 12, 1255–1256 (2014). <https://doi.org/10.1038/ng.3150>.
594. Skovbjerg, S. et al.: Gram-positive and gram-negative bacteria induce different patterns of cytokine production in human mononuclear cells irrespective of taxonomic relatedness. *J. Interferon Cytokine Res. Off. J. Int. Soc. Interferon Cytokine Res.* 30, 1, 23–32 (2010). <https://doi.org/10.1089/jir.2009.0033>.
595. Small, E., Clements, C.: Defining fever: likelihood of infection diagnosis as a function of body temperature in the emergency department. *Crit. Care.* 18, Suppl 2, P42 (2014). <https://doi.org/10.1186/cc14045>.
596. Smirnova, T. et al.: Assignment of the human syntaxin 1B gene (STX) to chromosome 16p11.2 by fluorescence in situ hybridization. *Genomics.* 36, 3, 551–553 (1996). <https://doi.org/10.1006/geno.1996.0506>.
597. Sobaniec, W. et al.: Evaluation of the influence of antiepileptic therapy on antioxidant enzyme activity and lipid peroxidation in erythrocytes of children with epilepsy. *J. Child Neurol.* 21, 7, 558–562 (2006). <https://doi.org/10.1177/0883073806210070501>.
598. Soeters, M.R., Soeters, P.B.: The evolutionary benefit of insulin resistance. *Clin. Nutr. Edinb. Scotl.* 31, 6, 1002–1007 (2012). <https://doi.org/10.1016/j.clnu.2012.05.011>.
599. Sofijanov, N. et al.: Febrile seizures: clinical characteristics and initial EEG. *Epilepsia.* 33, 1, 52–57 (1992). <https://doi.org/10.1111/j.1528-1157.1992.tb02282.x>.

600. Spagnoli, C. et al.: Early infantile SCN1A epileptic encephalopathy: Expanding the genotype-phenotype correlations. *Seizure*. 65, 62–64 (2019). <https://doi.org/10.1016/j.seizure.2019.01.002>.
601. Spagnolo, E. et al.: Gait patterns in Dravet syndrome: Preliminary data of a multicentric longitudinal prospective study. *Gait Posture*. 49, S10–S11 (2016). <https://doi.org/10.1016/j.gaitpost.2016.07.037>.
602. Spampinato, J. et al.: Increased neuronal firing in computer simulations of sodium channel mutations that cause generalized epilepsy with febrile seizures plus. *J. Neurophysiol.* 91, 5, 2040–2050 (2004). <https://doi.org/10.1152/jn.00982.2003>.
603. Steel, D. et al.: Dravet syndrome and its mimics: Beyond SCN1A. *Epilepsia*. 58, 11, 1807–1816 (2017). <https://doi.org/10.1111/epi.13889>.
- 420
604. Stein, R. et al.: Hippocampal deletion of Na V 1.1 channels in mice causes thermal seizures and cognitive deficit characteristic of Dravet Syndrome. *Proc. Natl. Acad. Sci.* 116, 201906833 (2019). <https://doi.org/10.1073/pnas.1906833116>.
605. Steinberg, B.E.: Neutrophils: A Therapeutic Target of Local Anesthetics? *Anesthesiology*. 128, 6, 1060–1061 (2018). <https://doi.org/10.1097/ALN.0000000000002205>.
606. Stenhouse, S.A.R. et al.: SCN1A Genetic Test for Dravet Syndrome (Severe Myoclonic Epilepsy of Infancy and its Clinical Subtypes) for use in the Diagnosis, Prognosis, Treatment and Management of Dravet Syndrome. *PLoS Curr.* 5, (2013). <https://doi.org/10.1371/currents.eogt.c553b83d745dd79bfb61eaf35e522b0b>.
607. Stores, G.: When does an EEG contribute to the management of febrile seizures? *Arch. Dis. Child.* 66, 4, 554–557 (1991). <https://doi.org/10.1136/adc.66.4.554>.
608. Stranneheim, H., Lundeberg, J.: Stepping stones in DNA sequencing. *Biotechnol. J.* 7, 9, 1063–1073 (2012). <https://doi.org/10.1002/biot.201200153>.
609. Straussberg, R. et al.: Pro- and anti-inflammatory cytokines in children with febrile convulsions. *Pediatr. Neurol.* 24, 1, 49–53 (2001). [https://doi.org/10.1016/s0887-8994\(00\)00234-4](https://doi.org/10.1016/s0887-8994(00)00234-4).
610. Strehl, U.: What learning theories can teach us in designing neurofeedback treatments. *Front. Hum. Neurosci.* 8, 894 (2014). <https://doi.org/10.3389/fnhum.2014.00894>.
611. Striano, P. et al.: Brain MRI findings in severe myoclonic epilepsy in infancy and genotype-phenotype correlations. *Epilepsia*. 48, 6, 1092–1096 (2007). <https://doi.org/10.1111/j.1528-1167.2007.01020.x>.
612. van Stuijvenberg, M. et al.: Characteristics of the initial seizure in familial febrile seizures. *Arch. Dis. Child.* 80, 2, 178–180 (1999). <https://doi.org/10.1136/adc.80.2.178>.
613. van Stuijvenberg, M. et al.: Temperature, age, and recurrence of febrile seizure. *Arch. Pediatr. Adolesc. Med.* 152, 12, 1170–1175 (1998). <https://doi.org/10.1001/archpedi.152.12.1170>.
614. Suega, K., Widiana, G.R.: Predicting hepcidin level using inflammation markers and iron indicators in patients with anemia of chronic disease. *Hematol. Transfus. Cell Ther.* 41, 4, 342–348 (2019). <https://doi.org/10.1016/j.htct.2019.03.011>.
615. Suga, S. et al.: Clinical and virological analyses of 21 infants with exanthem subitum (roseola infantum) and central nervous system complications. *Ann. Neurol.* 33, 6, 597–603 (1993). <https://doi.org/10.1002/ana.410330607>.
616. Sugai, K.: Current management of febrile seizures in Japan: an overview. *Brain Dev.* 32, 1, 64–70 (2010). <https://doi.org/10.1016/j.braindev.2009.09.019>.
617. Suls, A. et al.: Four generations of epilepsy caused by an inherited microdeletion of the SCN1A gene. *Neurology*. 75, 1, 72–76 (2010). <https://doi.org/10.1212/WNL.0b013e3181e62088>.
618. Suls, A. et al.: Microdeletions involving the SCN1A gene may be common in SCN1A-mutation-negative SMEI patients. *Hum. Mutat.* 27, 9, 914–920 (2006). <https://doi.org/10.1002/humu.20350>.

619. Sun, G. et al.: Carbamazepine and topiramate modulation of transient and persistent sodium currents studied in HEK293 cells expressing the Na(v)1.3 alpha-subunit. *Epilepsia*. 48, 4, 774–782 (2007). <https://doi.org/10.1111/j.1528-1167.2007.01001.x>.
620. Sun, Y. et al.: Risk of febrile seizures and epilepsy after vaccination with diphtheria, tetanus, acellular pertussis, inactivated poliovirus, and Haemophilus influenzae type B. *JAMA*. 307, 8, 823–831 (2012). <https://doi.org/10.1001/jama.2012.165>.
621. Symonds, J.D. et al.: Incidence and phenotypes of childhood-onset genetic epilepsies: a prospective population-based national cohort. *Brain J. Neurol.* 142, 8, 2303–2318 (2019). <https://doi.org/10.1093/brain/awz195>.
622. Syndi Seinfeld, D.O., Pellock, J.M.: Recent Research on Febrile Seizures: A Review. *J. Neurol. Neurophysiol.* 4, 165, (2013). <https://doi.org/10.4172/2155-9562.1000165>.
623. Taghizadegan, N. et al.: The Evaluation of Serum Selenium Concentration in Children with Febrile Convulsion. *Sci. J. Ilam Univ. Med. Sci.* 24, 4, 95–103 (2016). <https://doi.org/10.18869/acadpub.sjimu.24.4.95>.
624. Takano, T. et al.: Seizure susceptibility due to antihistamines in febrile seizures. *Pediatr. Neurol.* 42, 4, 277–279 (2010). <https://doi.org/10.1016/j.pediatrneurol.2009.11.001>.
625. Takao, T. et al.: Interleukin-1 receptors in mouse brain: characterization and neuronal localization. *Endocrinology*. 127, 6, 3070–3078 (1990). <https://doi.org/10.1210/endo-127-6-3070>.
626. Takeuchi, O., Akira, S.: Pattern recognition receptors and inflammation. *Cell*. 140, 6, 805–820 (2010). <https://doi.org/10.1016/j.cell.2010.01.022>.
- 421
627. Takeyama, H. et al.: Platelet Activation Markers Are Associated with Crohn's Disease Activity in Patients with Low C-Reactive Protein. *Dig. Dis. Sci.* 60, 11, 3418–3423 (2015). <https://doi.org/10.1007/s10620-015-3745-2>.
628. Tan, N.C.K. et al.: Genetic association studies in epilepsy: “the truth is out there.” *Epilepsia*. 45, 11, 1429–1442 (2004). <https://doi.org/10.1111/j.0013-9580.2004.22904.x>.
629. Tang, J. et al.: Relationship between common viral upper respiratory tract infections and febrile seizures in children from Suzhou, China. *J. Child Neurol.* 29, 10, 1327–1332 (2014). <https://doi.org/10.1177/0883073813515074>.
630. Tang, Y.-Q. et al.: Antimicrobial peptides from human platelets. *Infect. Immun.* 70, 12, 6524–6533 (2002). <https://doi.org/10.1128/iai.70.12.6524-6533.2002>.
631. Taşoğlu, I. et al.: Usefulness of Neutrophil/Lymphocyte Ratio as a Predictor of Amputation after Embolectomy for Acute Limb Ischemia. *Ann. Vasc. Surg.* 28, 3, 606–613 (2014). <https://doi.org/10.1016/j.avsg.2012.12.009>.
632. Templeton, A.J. et al.: Prognostic Role of Neutrophil-to-Lymphocyte Ratio in Solid Tumors: A Systematic Review and Meta-Analysis. *JNCI J. Natl. Cancer Inst.* 106, dju124, (2014). <https://doi.org/10.1093/jnci/dju124>.
633. Thachil, J.: Platelets in Inflammatory Disorders: A Pathophysiological and Clinical Perspective. *Semin. Thromb. Hemost.* 41, (2015). <https://doi.org/10.1055/s-0035-1556589>.
634. Thébault-Dagher, F. et al.: Age at first febrile seizure correlates with perinatal maternal emotional symptoms. *Epilepsy Res.* 135, 95–101 (2017). <https://doi.org/10.1016/j.eplepsyres.2017.06.001>.
635. Thilothammal, N. et al.: Risk factors for recurrence of febrile convulsions. *Indian J. Pediatr.* 59, 6, 749–754 (1992). <https://doi.org/10.1007/BF02859414>.
636. Thoman, J.E. et al.: Do serum sodium levels predict febrile seizure recurrence within 24 hours? *Pediatr. Neurol.* 31, 5, 342–344 (2004). <https://doi.org/10.1016/j.pediatrneurol.2004.05.013>.
637. Thompson, C.B., Jakubowski, J.A.: The Pathophysiology and Clinical Relevance of Platelet Heterogeneity. *Blood*. 72, 1, 1–8 (1988). <https://doi.org/10.1182/blood.V72.1.1.1>.

638. Thoresen, M. et al.: Does a sedative dose of chloral hydrate modify the EEG of children with epilepsy? *Electroencephalogr. Clin. Neurophysiol.* 102, 2, 152–157 (1997). [https://doi.org/10.1016/s0921-884x\(96\)96509-1](https://doi.org/10.1016/s0921-884x(96)96509-1).
639. Threatte, G.A.: Usefulness of the Mean Platelet Volume. *Clin. Lab. Med.* 13, 4, 937–950 (1993). [https://doi.org/10.1016/S0272-2712\(18\)30418-9](https://doi.org/10.1016/S0272-2712(18)30418-9).
640. Tian, Y. et al.: A novel inherited STX1B mutation associated with generalized epilepsy with febrile seizures plus: a family analysis and literature review]. *Zhonghua Er Ke Za Zhi Chin. J. Pediatr.* 57, 3, 206–210 (2019). <https://doi.org/10.3760/cma.j.issn.0578-1310.2019.03.010>.
641. Tilgen, N. et al.: Association analysis between the human interleukin 1beta (-511) gene polymorphism and susceptibility to febrile convulsions. *Neurosci. Lett.* 334, 1, 68–70 (2002). [https://doi.org/10.1016/s0304-3940\(02\)01069-8](https://doi.org/10.1016/s0304-3940(02)01069-8).
642. Todd, E. et al.: GABAA receptor biogenesis is impaired by the γ 2 subunit febrile seizure-associated mutation, GABRG2(R177G). *Neurobiol. Dis.* 69, 215–224 (2014). <https://doi.org/10.1016/j.nbd.2014.05.013>.
643. Tolner, E.A. et al.: Five percent CO₂ is a potent, fast-acting inhalation anticonvulsant. *Epilepsia.* 52, 1, 104–114 (2011). <https://doi.org/10.1111/j.1528-1167.2010.02731.x>.
644. Tosun, A. et al.: Ratios of nine risk factors in children with recurrent febrile seizures. *Pediatr. Neurol.* 43, 3, 177–182 (2010). <https://doi.org/10.1016/j.pediatrneurol.2010.05.007>.
645. Toth, Z. et al.: Seizure-induced neuronal injury: vulnerability to febrile seizures in an immature rat model. *J. Neurosci. Off. J. Soc. Neurosci.* 18, 11, 4285–4294 (1998).
646. Traynelis, S.F., Cull-Candy, S.G.: Proton inhibition of N-methyl-D-aspartate receptors in cerebellar neurons. *Nature.* 345, 6273, 347–350 (1990). <https://doi.org/10.1038/345347a0>.
647. Trinka, E. et al.: Childhood febrile convulsions--which factors determine the subsequent epilepsy syndrome? A retrospective study. *Epilepsy Res.* 50, 3, 283–292 (2002). [https://doi.org/10.1016/s0920-1211\(02\)00083-9](https://doi.org/10.1016/s0920-1211(02)00083-9).
648. Tsuboi, T., Okada, S.: Exogenous causes of seizures in children: A population study. *Acta Neurol. Scand.* 71, 2, 107–113 (1985). <https://doi.org/10.1111/j.1600-0404.1985.tb03174.x>.
649. Tsuboi, T., Okada, S.: Seasonal variation of febrile convolution in Japan. *Acta Neurol. Scand.* 69, 5, 285–292 (1984). <https://doi.org/10.1111/j.1600-0404.1984.tb07814.x>.
650. Tu, Y.-F. et al.: Febrile convulsions increase risk of Tourette syndrome. *Seizure.* 23, 8, 651–656 (2014). <https://doi.org/10.1016/j.seizure.2014.05.005>.
- 422
651. Tu, Y.-F. et al.: Postnatal Steroids and Febrile Seizure Susceptibility in Preterm Children. *Pediatrics.* 137, 4, (2016). <https://doi.org/10.1542/peds.2015-3404>.
652. Unger, E.L. et al.: Dopamine D2 Receptor Expression Is Altered by Changes in Cellular Iron Levels in PC12 Cells and Rat Brain Tissue. *J. Nutr.* 138, 12, 2487–2494 (2008). <https://doi.org/10.3945/jn.108.095224>.
653. Vadlamudi, L. et al.: Timing of De Novo Mutagenesis — A Twin Study of Sodium-Channel Mutations. *N. Engl. J. Med.* 363, 14, 1335–1340 (2010). <https://doi.org/10.1056/NEJMoa0910752>.
654. Vagdatli, E. et al.: Platelet distribution width: a simple, practical and specific marker of activation of coagulation. *Hippokratia.* 14, 1, 28–32 (2010).
655. Vahidnia, F. et al.: Maternal smoking, alcohol drinking, and febrile convulsion. *Seizure.* 17, 4, 320–326 (2008). <https://doi.org/10.1016/j.seizure.2007.10.003>.
656. Valerio, G. et al.: High prevalence of stress hyperglycaemia in children with febrile seizures and traumatic injuries. *Acta Paediatr. Oslo Nor.* 1992. 90, 6, 618–622 (2001).
657. Vamvakopoulos, J. et al.: Genetic control of IL-1 β bioactivity through differential regulation of the IL-1 receptor antagonist. *Eur. J. Immunol.* 32, 10, 2988–2996 (2002). [https://doi.org/10.1002/1521-4141\(2002010\)32:10<2988::AID-IMMU2988>3.0.CO;2-9](https://doi.org/10.1002/1521-4141(2002010)32:10<2988::AID-IMMU2988>3.0.CO;2-9).

658. Van Poppel, K. et al.: Mesial temporal sclerosis in a cohort of children with SCN1A gene mutation. *J. Child Neurol.* 27, 7, 893–897 (2012). <https://doi.org/10.1177/0883073811435325>.
659. Vanoye, C.G. et al.: Single-channel properties of human NaV1.1 and mechanism of channel dysfunction in SCN1A-associated epilepsy. *J. Gen. Physiol.* 127, 1, 1–14 (2006). <https://doi.org/10.1085/jgp.200509373>.
660. Verburgh, M.E. et al.: Incidence of febrile seizures in The Netherlands. *Neuroepidemiology*. 11, 4–6, 169–172 (1992). <https://doi.org/10.1159/000110928>.
661. Verity, C.M. et al.: Febrile convulsions in a national cohort followed up from birth. I—Prevalence and recurrence in the first five years of life. *Br. Med. J. Clin. Res. Ed.* 290, 6478, 1307–1310 (1985). <https://doi.org/10.1136/bmj.290.6478.1307>.
662. Verity, C.M. et al.: Long-term intellectual and behavioral outcomes of children with febrile convulsions. *N. Engl. J. Med.* 338, 24, 1723–1728 (1998). <https://doi.org/10.1056/NEJM199806113382403>.
663. Verity, C.M., Golding, J.: Risk of epilepsy after febrile convulsions: a national cohort study. *BMJ*. 303, 6814, 1373–1376 (1991). <https://doi.org/10.1136/bmj.303.6814.1373>.
664. Vestergaard, M. et al.: Prenatal exposure to cigarettes, alcohol, and coffee and the risk for febrile seizures. *Pediatrics*. 116, 5, 1089–1094 (2005). <https://doi.org/10.1542/peds.2004-2210>.
665. Vestergaard, M. et al.: Risk factors for febrile convulsions. *Epidemiol. Camb. Mass.* 13, 3, 282–287 (2002). <https://doi.org/10.1097/00001648-200205000-00008>.
666. Vestergaard, M. et al.: The long-term risk of epilepsy after febrile seizures in susceptible subgroups. *Am. J. Epidemiol.* 165, 8, 911–918 (2007). <https://doi.org/10.1093/aje/kwk086>.
667. Vestergaard, M., Christensen, J.: Register-based studies on febrile seizures in Denmark. *Brain Dev.* 31, 5, 372–377 (2009). <https://doi.org/10.1016/j.braindev.2008.11.012>.
668. Vezyroglou, A. et al.: Focal epilepsy in SCN1A -mutation carrying patients: is there a role for epilepsy surgery? *Dev. Med. Child Neurol.* 62, (2020). <https://doi.org/10.1111/dmcn.14588>.
669. Vezzani, A. et al.: Powerful anticonvulsant action of IL-1 receptor antagonist on intracerebral injection and astrocytic overexpression in mice. *Proc. Natl. Acad. Sci. U. S. A.* 97, 21, 11534–11539 (2000). <https://doi.org/10.1073/pnas.190206797>.
670. Vezzani, A. et al.: The role of inflammation in epilepsy. *Nat. Rev. Neurol.* 7, 1, 31–40 (2011). <https://doi.org/10.1038/nrneurol.2010.178>.
671. Vezzani, A., Granata, T.: Brain inflammation in epilepsy: experimental and clinical evidence. *Epilepsia*. 46, 11, 1724–1743 (2005). <https://doi.org/10.1111/j.1528-1167.2005.00298.x>.
672. Viana, M.V. et al.: Assessment and treatment of hyperglycemia in critically ill patients. *Rev. Bras. Ter. Intensiva*. 26, 1, 71–76 (2014). <https://doi.org/10.5935/0103-507X.20140011>.
673. Villeneuve, N. et al.: Cognitive and adaptive evaluation of 21 consecutive patients with Dravet syndrome. *Epilepsy Behav.* EB. 31C, 143–148 (2014). <https://doi.org/10.1016/j.yebeh.2013.11.021>.
674. Virta, M. et al.: Increased frequency of interleukin-1beta (-511) allele 2 in febrile seizures. *Pediatr. Neurol.* 26, 3, 192–195 (2002). [https://doi.org/10.1016/s0887-8994\(01\)00380-0](https://doi.org/10.1016/s0887-8994(01)00380-0).
- 423
675. Virta, M. et al.: Increased plasma levels of pro- and anti-inflammatory cytokines in patients with febrile seizures. *Epilepsia*. 43, 8, 920–923 (2002). <https://doi.org/10.1046/j.1528-1157.2002.02002.x>.
676. Visser, A.M. et al.: Febrile seizures and behavioural and cognitive outcomes in preschool children: the Generation R study. *Dev. Med. Child Neurol.* 54, 11, 1006–1011 (2012). <https://doi.org/10.1111/j.1469-8749.2012.04405.x>.
677. Visser, A.M. et al.: Fetal growth retardation and risk of febrile seizures. *Pediatrics*. 126, 4, e919–925 (2010). <https://doi.org/10.1542/peds.2010-0518>.

678. Viviani, B. et al.: Interleukin-1beta enhances NMDA receptor-mediated intracellular calcium increase through activation of the Src family of kinases. *J. Neurosci. Off. J. Soc. Neurosci.* 23, 25, 8692–8700 (2003).
679. Wagner, S., Yarom, Y.: Excitation by GABA in the SCN reaches its time and place (Commentary on Irwin & Allen). *Eur. J. Neurosci.* 30, 8, 1461 (2009). <https://doi.org/10.1111/j.1460-9568.2009.07011.x>.
680. Wahlestedt, C.: Targeting long non-coding RNA to therapeutically upregulate gene expression. *Nat. Rev. Drug Discov.* 12, 6, 433–446 (2013). <https://doi.org/10.1038/nrd4018>.
681. Wallace, A. et al.: Pharmacotherapy for Dravet Syndrome. *Paediatr. Drugs.* 18, 3, 197–208 (2016). <https://doi.org/10.1007/s40272-016-0171-7>.
682. Wallace, R.H. et al.: Febrile seizures and generalized epilepsy associated with a mutation in the Na⁺-channel beta1 subunit gene SCN1B. *Nat. Genet.* 19, 4, 366–370 (1998). <https://doi.org/10.1038/1252>.
683. Wallace, R.H. et al.: Sodium channel α1-subunit mutations in severe myoclonic epilepsy of infancy and infantile spasms. *Neurology.* 61, 6, 765–769 (2003). <https://doi.org/10.1212/01.WNL.0000086379.71183.78>.
684. Wang, J. et al.: Distribution and function of voltage-gated sodium channels in the nervous system. *Channels Austin Tex.* 11, 6, 534–554 (2017). <https://doi.org/10.1080/19336950.2017.1380758>.
685. Wang, J. et al.: Gene mutational analysis in a cohort of Chinese children with unexplained epilepsy: Identification of a new KCND3 phenotype and novel genes causing Dravet syndrome. *Seizure.* 66, 26–30 (2019). <https://doi.org/10.1016/j.seizure.2019.01.025>.
686. Wang, P. et al.: Association of interleukin-1 gene polymorphisms with gastric cancer: A meta-analysis. *Int. J. Cancer.* 120, 3, 552–562 (2007). <https://doi.org/10.1002/ijc.22353>.
687. Wang, Z.J. et al.: Association between SCN1A polymorphism rs3812718 and valproic acid resistance in epilepsy children: a case-control study and meta-analysis. *Biosci. Rep.* 38, 6, (2018). <https://doi.org/10.1042/BSR20181654>.
688. Waqar Rabbani, M. et al.: Serum Zinc Level in Children Presenting with Febrile Seizures. *Pak. J. Med. Sci.* 29, 4, 1008–1011 (2013).
689. Waruiru, C., Appleton, R.: Febrile seizures: an update. *Arch. Dis. Child.* 89, 8, 751–756 (2004). <https://doi.org/10.1136/adc.2003.028449>.
690. Weber, Y.G. et al.: Genetic biomarkers in epilepsy. *Neurother. J. Am. Soc. Exp. Neurother.* 11, 2, 324–333 (2014). <https://doi.org/10.1007/s13311-014-0262-5>.
691. Weiss, L.A. et al.: Sodium channels SCN1A, SCN2A and SCN3A in familial autism. *Mol. Psychiatry.* 8, 2, 186–194 (2003). <https://doi.org/10.1038/sj.mp.4001241>.
692. Weiss, S.L. et al.: Extreme stress hyperglycemia during acute illness in a pediatric emergency department. *Pediatr. Emerg. Care.* 26, 9, 626–632 (2010). <https://doi.org/10.1097/PEC.0b013e3181ef0488>.
693. Whitaker, W.R.J. et al.: Comparative distribution of voltage-gated sodium channel proteins in human brain. *Mol. Brain Res.* 88, 1, 37–53 (2001). [https://doi.org/10.1016/S0169-328X\(00\)00289-8](https://doi.org/10.1016/S0169-328X(00)00289-8).
694. Wipfler, P. et al.: The Viral Hypothesis of Mesial Temporal Lobe Epilepsy – Is Human Herpes Virus-6 the Missing Link? A systematic review and meta-analysis. *Seizure - Eur. J. Epilepsy.* 54, 33–40 (2018). <https://doi.org/10.1016/j.seizure.2017.11.015>.
695. Wirrell, E.C. et al.: Optimizing the Diagnosis and Management of Dravet Syndrome: Recommendations From a North American Consensus Panel. *Pediatr. Neurol.* 68, 18-34.e3 (2017). <https://doi.org/10.1016/j.pediatrneurol.2017.01.025>.
696. Wirrell, E.C.: Treatment of Dravet Syndrome. *Can. J. Neurol. Sci. J. Can. Sci. Neurol.* 43 Suppl 3, S13-18 (2016). <https://doi.org/10.1017/cjn.2016.249>.

697. Wirrell, E.C., Nabbout, R.: Recent Advances in the Drug Treatment of Dravet Syndrome. *CNS Drugs.* 33, 9, 867–881 (2019). <https://doi.org/10.1007/s40263-019-00666-8>.
698. Wiwanitkit, V.: Plateleterit, mean platelet volume, platelet distribution width: its expected values and correlation with parallel red blood cell parameters. *Clin. Appl. Thromb. Off. J. Int. Acad. Clin. Appl. Thromb.* 10, 2, 175–178 (2004). <https://doi.org/10.1177/107602960401000208>.
699. Wo, S.B. et al.: Risk for developing epilepsy and epileptiform discharges on EEG in patients with febrile seizures. *Brain Dev.* 35, 4, 307–311 (2013). <https://doi.org/10.1016/j.braindev.2012.07.014>.
700. Wyers, L. et al.: Gait deviations in patients with dravet syndrome: A systematic review. *Eur. J. Paediatr. Neurol. EJPN Off. J. Eur. Paediatr. Neurol. Soc.* 23, 3, 357–367 (2019). <https://doi.org/10.1016/j.ejpn.2019.03.003>.
701. Xing, D. et al.: Expression of neonatal Nav1.5 in human brain astrocytoma and its effect on proliferation, invasion and apoptosis of astrocytoma cells. *Oncol. Rep.* 31, 6, 2692–2700 (2014). <https://doi.org/10.3892/or.2014.3143>.
702. Xu, X. et al.: Amplicon Resequencing Identified Parental Mosaicism for Approximately 10% of “de novo” SCN1A Mutations in Children with Dravet Syndrome. *Hum. Mutat.* 36, 9, 861–872 (2015). <https://doi.org/10.1002/humu.22819>.
703. Yakoub, M. et al.: Early diagnosis of severe myoclonic epilepsy in infancy. *Brain Dev.* 14, 5, 299–303 (1992). [https://doi.org/10.1016/S0387-7604\(12\)80147-1](https://doi.org/10.1016/S0387-7604(12)80147-1).
704. Yamada, K. et al.: Protective role of ATP-sensitive potassium channels in hypoxia-induced generalized seizure. *Science.* 292, 5521, 1543–1546 (2001). <https://doi.org/10.1126/science.1059829>.
705. Yang, A. et al.: Mean platelet volume as marker of restenosis after percutaneous transluminal coronary angioplasty in patients with stable and unstable angina pectoris. *Thromb. Res.* 117, 4, 371–377 (2006). <https://doi.org/10.1016/j.thromres.2005.04.004>.
706. Yang, H. et al.: Glycolysis in energy metabolism during seizures. *Neural Regen. Res.* 8, 14, 1316–1326 (2013). <https://doi.org/10.3969/j.issn.1673-5374.2013.14.008>.
707. Yazar, A. et al.: Mean Platelet Volume and Neutrophil-to-Lymphocyte Ratio May Be Used as Predictors in Febrile Seizures. *J. Pediatr. Infect. Dis.* 13, 04, 283–286 (2018). <https://doi.org/10.1055/s-0038-1668534>.
708. Yazici, S. et al.: The platelet functions in patients with ankylosing spondylitis: anti-TNF-alpha therapy decreases the mean platelet volume and platelet mass. *Platelets.* 21, 2, 126–131 (2010). <https://doi.org/10.3109/09537100903470306>.
709. Yeşil, A. et al.: Red cell distribution width: a novel marker of activity in inflammatory bowel disease. *Gut Liver.* 5, 4, 460–467 (2011). <https://doi.org/10.5009/gnl.2011.5.4.460>.
710. Yigit, Y. et al.: The role of neutrophil-lymphocyte ratio and red blood cell distribution width in the classification of febrile seizures. *Eur. Rev. Med. Pharmacol. Sci.* 21, 3, 554–559 (2017).
711. Yıldızdaş, D. et al.: Correlation of simultaneously obtained capillary, venous, and arterial blood gases of patients in a paediatric intensive care unit. *Arch. Dis. Child.* 89, 2, 176–180 (2004). <https://doi.org/10.1136/adc.2002.016261>.
712. Yoshikawa, T. et al.: Human herpesvirus-6 DNA in cerebrospinal fluid of a child with exanthem subitum and meningoencephalitis. *Pediatrics.* 89, 5 Pt 1, 888–890 (1992).
713. Youn, Y. et al.: The role of cytokines in seizures: interleukin (IL)-1 β , IL-1Ra, IL-8, and IL-10. *Korean J. Pediatr.* 56, 7, 271–274 (2013). <https://doi.org/10.3345/kjp.2013.56.7.271>.
714. Yu, M.-J. et al.: Milder phenotype with SCN1A truncation mutation other than SMEI. *Seizure.* 19, 7, 443–445 (2010). <https://doi.org/10.1016/j.seizure.2010.06.010>.

715. Yu, X. et al.: Polymorphisms in the interleukin-1 β (IL-1B) and interleukin-1 α (IL-1A) genes on risk of febrile seizures: a meta-analysis. *Neurol. Sci. Off. J. Ital. Neurol. Soc. Ital. Soc. Clin. Neurophysiol.* 39, 9, 1529–1536 (2018). <https://doi.org/10.1007/s10072-018-3449-4>.
716. Yücel, O. et al.: Role of early EEG and neuroimaging in determination of prognosis in children with complex febrile seizure. *Pediatr. Int. Off. J. Jpn. Pediatr. Soc.* 46, 4, 463–467 (2004). <https://doi.org/10.1111/j.1328-8067.2003.01799.x>.
717. Zancan, P. et al.: Clotrimazole inhibits and modulates heterologous association of the key glycolytic enzyme 6-phosphofructo-1-kinase. *Biochem. Pharmacol.* 73, 10, 1520–1527 (2007). <https://doi.org/10.1016/j.bcp.2007.01.018>.
718. Zancan, P., Sola-Penna, M.: Regulation of human erythrocyte metabolism by insulin: cellular distribution of 6-phosphofructo-1-kinase and its implication for red blood cell function. *Mol. Genet. Metab.* 86, 3, 401–411 (2005). <https://doi.org/10.1016/j.ymgme.2005.06.011>.
- 425
719. Zareifar, S. et al.: Association between iron status and febrile seizures in children. *Seizure.* 21, 8, 603–605 (2012). <https://doi.org/10.1016/j.seizure.2012.06.010>.
720. Zeng, S. et al.: Long-read sequencing identified intronic repeat expansions in SAMD12 from Chinese pedigrees affected with familial cortical myoclonic tremor with epilepsy. *J. Med. Genet.* 56, 4, 265–270 (2019). <https://doi.org/10.1136/jmedgenet-2018-105484>.
721. Zeng, T. et al.: A novel variant in the 3' UTR of human SCN1A gene from a patient with Dravet syndrome decreases mRNA stability mediated by GAPDH's binding. *Hum. Genet.* 133, 6, 801–811 (2014). <https://doi.org/10.1007/s00439-014-1422-8>.
722. Zhang, F. et al.: Combination of platelet count and mean platelet volume (COP-MPV) predicts postoperative prognosis in both resectable early and advanced stage esophageal squamous cell cancer patients. *Tumour Biol. J. Int. Soc. Oncodevelopmental Biol. Med.* 37, 7, 9323–9331 (2016). <https://doi.org/10.1007/s13277-015-4774-3>.
723. Zhang, S. et al.: Use of Platelet Indices for Determining Illness Severity and Predicting Prognosis in Critically Ill Patients. *Chin. Med. J. (Engl.)* 128, 15, 2012–2018 (2015). <https://doi.org/10.4103/0366-6999.161346>.
724. Zhang, T. et al.: Are afebrile seizures associated with minor infections a single seizure category? A hospital-based prospective cohort study on outcomes of first afebrile seizure in early childhood. *Epilepsia.* 55, 7, 1001–1008 (2014). <https://doi.org/10.1111/epi.12651>.
725. Zhang, Y. et al.: Association of SCN1A gene polymorphisms with infantile spasms and adrenocorticotropic hormone responsiveness. *Eur. Rev. Med. Pharmacol. Sci.* 18, 17, 2500–2506 (2014).
726. Zhang, Y.-H. et al.: Genetic epilepsy with febrile seizures plus: Refining the spectrum. *Neurology.* 89, 12, 1210–1219 (2017). <https://doi.org/10.1212/WNL.0000000000004384>.
727. Zhu, G. et al.: Effects of interleukin-1 beta on hippocampal glutamate and GABA releases associated with Ca²⁺-induced Ca²⁺ releasing systems. *Epilepsy Res.* 71, 107–16 (2006). <https://doi.org/10.1016/j.eplepsyres.2006.05.017>.
728. Ziemann, A.E. et al.: Seizure termination by acidosis depends on ASIC1a. *Nat. Neurosci.* 11, 7, 816–822 (2008). <https://doi.org/10.1038/nn.2132>.
729. Zolaly, M.A.: Histamine H1 antagonists and clinical characteristics of febrile seizures. *Int. J. Gen. Med.* 5, 277–281 (2012). <https://doi.org/10.2147/IJGM.S29320>.
730. Zou, F. et al.: Expanding the phenotypic spectrum of GABRG2 variants: a recurrent GABRG2 missense variant associated with a severe phenotype. *J. Neurogenet.* 31, 1–2, 30–36 (2017). <https://doi.org/10.1080/01677063.2017.1315417>.
731. Zuberi, S.M. et al.: Genotype-phenotype associations in SCN1A-related epilepsies. *Neurology.* 76, 7, 594–600 (2011). <https://doi.org/10.1212/WNL.0b013e31820c309b>.

732. acidbase.org, <https://acidbase.org/>, last accessed 2021/07/23.
733. ClinicalTrials.gov, <https://clinicaltrials.gov/>, last accessed 2020/10/16.
734. Dravet syndrome gene therapy, <http://www.draccon.com/dracaena-report/2019/2/25/dravet-syndrome-gene-therapy>, last accessed 2021/03/14.
735. EpilepsyDiagnosis.org, <https://www.epilepsydiagnosis.org/>, last accessed 2020/03/26.
736. Gene: SCN1A (ENSG00000144285) - Summary - Homo_sapiens - Ensembl genome browser 103, http://www.ensembl.org/Homo_sapiens/Gene/Summary?g=ENSG00000144285;r=2:165984641-166149214, last accessed 2021/03/14.
737. Guidelines for epidemiologic studies on epilepsy. Commission on Epidemiology and Prognosis, International League Against Epilepsy. *Epilepsia*. 34, 4, 592–596 (1993). <https://doi.org/10.1111/j.1528-1157.1993.tb00433.x>.
738. MLPA Technique - MRC Holland, <https://www.mrcholland.com/technology/mlpa/technique>, last accessed 2021/03/14.
739. OMIM - Online Mendelian Inheritance in Man, <https://www.omim.org/>, last accessed 2021/03/08.
740. OMIM Phenotypic Series - PS604233, <https://www.omim.org/phenotypicSeries/PS604233>, last accessed 2021/03/14.
741. Our Science – Stoke Therapeutics, <https://www.stoketherapeutics.com/our-science/>, last accessed 2021/03/14.
742. Paediatric Reference Ranges – Microscopic 3e Haematology, <https://microscopic-haematology.com/paediatric-reference-ranges/>, last accessed 2020/07/05.
- 426
743. Practice parameter: the neurodiagnostic evaluation of the child with a first simple febrile seizure. American Academy of Pediatrics. Provisional Committee on Quality Improvement, Subcommittee on Febrile Seizures. *Pediatrics*. 97, 5, 769–772; discussion 773-775 (1996).
744. SCN1A – this is what you need to know | Beyond the Ion Channel, <http://epilepsygenetics.net/the-epilepsiome/scn1a-this-is-what-you-need-to-know/>, (2020).
745. SCN1A mutation database!, <http://scn1a.caae.org.cn/index.php>, last accessed 2021/03/07.
746. Venous blood gases and other alternatives to arterial blood gases - UpToDate, <https://www.uptodate.com/contents/venous-blood-gases-and-other-alternatives-to-arterial-blood-gases>, (2019).